
1 Exposure

*David J. Spurgeon, Hana R. Pohl, Susana Loureiro,
Hans Løkke, and Cornelis A. M. van Gestel*

CONTENTS

1.1	Introduction	2
1.2	Emission Scenarios	4
1.2.1	Major Emission Sources	4
1.2.2	Emission Estimation Methods	5
1.2.3	Prioritization	9
1.2.4	Validation Studies	10
1.3	Interactions Affecting Availability and Exposure to Chemical Mixtures	10
1.3.1	Characteristics of the Major Environmental Compartments	10
1.3.2	Environmental Fate Affecting Mixture Composition	11
1.3.2.1	Single Compounds as Chemical Mixtures	11
1.3.2.2	Chemical Fate Effects on Mixture Composition	12
1.3.3	Availability	14
1.3.3.1	Availability and Bioavailability	14
1.3.3.2	Influence of Medium Physical–Chemical Properties on Chemical Availability	16
1.3.3.3	Metal Speciation Determines Bioavailability	17
1.3.3.4	Species Specificity	19
1.3.3.5	Formulating Agents	20
1.3.3.6	Analytical–Chemical Procedures	20
1.3.4	Chemical–Chemical Interactions in Mixtures	21
1.4	Environmental Fate Modeling	21
1.5	Exposure Scenarios and Monitoring	23
1.5.1	Human Exposure	24
1.5.1.1	Environmental Exposures Excluding Food	24
1.5.1.2	Food	28
1.5.1.3	Human Exposure in Different Life Stages	28
1.5.1.4	Modeling and Measuring Human Exposure	32
1.5.1.5	Human Biobanks and Human Volunteer Monitoring of Exposure	33
1.5.2	Exposure in Ecosystems	34
1.5.2.1	Air	35
1.5.2.2	Water	36
1.5.2.3	Sediment	38
1.5.2.4	Soil	38

1.5.2.5	Monitoring of Food Chain Transfer.....	39
1.5.2.6	Multimedia Exposure Scenarios.....	41
1.5.2.7	Critique on Biomonitoring Studies for Complex Exposure Assessment.....	42
1.5.2.8	Effect-Directed Assessment.....	42
1.6	Summary and Conclusions	43
1.7	Recommendations	43
	Acknowledgments.....	45

1.1 INTRODUCTION

In the environment, organisms including man are exposed to mixtures of chemicals rather than single compounds. Examples of mixtures are food and feedstuff, pesticide and medical products, dyes, cosmetics, and alloys. Many other commercial products, such as printing inks, contain a mixture of substances, possibly up to 60 individual chemicals in 1 formulation. Preparation of these chemicals may involve the use of several hundred other substances in upstream processes.

As a first step in the risk assessment of chemicals, it is essential to have an insight into the magnitude and duration of exposure. Following the toxicological principle that dose determines the effect, one may assume that no exposure means no risk. In the case of chemical mixtures, a proper assessment of exposure assists in adequately interpreting the interacting effects of chemicals. So, exposure assessment is an essential component of any risk assessment study of mixtures, since it can be used to reduce uncertainty and provide data.

The exposure of organisms includes man-made chemicals as well as natural compounds. Natural compounds are, for example, toxins in plants, ozone, or natural occurring metals. The total number of man-made chemicals is vast. To assess exposure, the ambient concentrations of chemicals resulting from man-made sources need to be known or estimated. Chemical Abstracts, covering more than 8000 journals since 1907, registers more than 20 million entries. This section focuses on man-made chemicals. In Europe, around 30,000 chemicals are commonly used and thereby emitted to the environment (EC 2001).

In human health risk assessment, “direct” and “indirect” methods of exposure assessment are distinguished. The direct method involves measurements of exposure at the point of contact or uptake, for instance, by monitoring chemical concentrations in humans or the environments they are exposed to (food, air, water). The indirect methods use modeling and extrapolation techniques to estimate exposure levels (Fryer et al. 2006). Also in environmental exposure assessment, these 2 ways to assess exposure may be applied.

Indirect exposure assessment, both human and environmental, starts with emission data and a prediction of the fate of chemicals in the environment and the resulting concentrations in different environmental compartments. Foster et al. (2005) outlined 5 steps in a strategy to conduct exposure assessment of complex mixtures, consisting of many different components, such as gasoline. These steps, as outlined below, are also relevant when assessing exposure to less complex mixtures.

- 1) Determination of mixture composition. Composition of the mixture may vary spatially and temporally. Measurements at the source (point of emission) may help in identifying (variations in) mixture composition.
- 2) Selection of component groups (optional). Within a mixture, different (groups of) components may be identified. These components may be grouped on the basis of properties that affect their fate in the environment.
- 3) Compilation of relevant property data for each group. This step consists of collecting properties relevant for predicting the fate of the different (groups of) components in the environment.
- 4) Assessment of the environmental fate of each group. Fate models may be used to predict environmental fate of mixture components on different spatial scales. Such models may yield a predicted distribution over air, water, soil, and sediment.
- 5) Assessment of environmental and human exposure. As a final step, concentrations can be calculated for each of the (groups of) mixture components in different exposure media (inhaled air, ingested water, food items) or environmental compartments (soil, sediment, air, and surface or groundwater). This information may not, however, represent the complete picture: often only part of the total concentration in an environmental compartment is biologically available for uptake by organisms. In addition, species habits and individual behavior may affect the nature of exposure. Finally, life-stage-specific aspects may be highly important in determining exposure to mixtures; this aspect is best studied for human exposure, but is also relevant to ecological assessment for some taxa.

For exposure assessment of ecosystems, direct exposure assessment involves taking field samples at the site and time of exposure and measuring chemical concentrations in these samples or in the organisms exposed at the site. Direct assessment of (potential) exposure also is possible by performing bioassays in which selected test organisms are exposed to the environmental sample, in the laboratory or in the field. The latter approach is discussed in more detail in Chapter 4.

In this chapter, the different steps in the assessment of mixture exposure are discussed. The chapter starts from emission scenarios and subsequently discusses transformation processes taking place in the environment and their effects on mixture composition. Next, bioavailability is discussed, and exposure scenarios for both humans and biota in the environment are described. These descriptions also consider methods to assess exposure to mixtures. Most data available on mixture exposure are restricted to North America and Europe, but we recognize that there are emerging problems in other regions of the world. We restrict our discussion to man-made chemicals and those natural chemicals subject to regulation (metals, polycyclic aromatic hydrocarbons (PAHs)), because these represent the most well-studied group and the current priorities for risk assessment.

1.2 EMISSION SCENARIOS

An emission is defined as the amount of chemical discharged or transferred per unit time, or it is the amount of chemical per unit volume of gas or liquid emitted. The emission can be characterized by the following attributes (OECD 2006):

- 1) Pollutant type
- 2) Release medium
- 3) Source type
- 4) Spatial scale
- 5) Temporal scale

Normally, an emission assessment deals with a single chemical or a group of chemicals, which have similar properties, such as PAHs, metals, ozone-depleting substances, or chlorinated biphenyls. In the environment, organisms and man are exposed to mixtures of chemicals with different properties rather than single chemicals or chemicals with similar properties. As an example, many commercial products (e.g., inks, oils, lubricants) contain a mixture of substances in a single formulation and so may be released simultaneously to environmental media, including land, surface water, groundwater, and indoor and outdoor air.

1.2.1 MAJOR EMISSION SOURCES

Emission sources are generally divided between point, diffuse, and mobile sources (OECD 2006). Point sources, such as industrial plants, power stations, waste incinerators, and sewage treatment plants, may play a major role as sources of mixtures of chemicals. Emissions from such sources are frequently of multiple chemicals; even in cases where the emission is dominated by a single chemical, overlap of plumes from other nearby point sources for different chemicals means that the surrounding areas are subject to combined exposure. Diffuse emissions from the application of pesticides and biocides and the domestic and widespread commercial use of chemicals can make a major contribution to the release of chemical mixtures into air, soils, and waters. In the case of pesticides, these biologically active compounds are applied as mixtures, or the application is repeated with other types of active ingredients within a short period, so that more than 1 chemical is present. As for local sources, even when there is not deliberate combined release, overlapping release and transport mechanisms in the atmospheric and the aquatic environments result in the widespread presence of chemical mixtures in different environmental compartments. Emissions from mobile sources, such as vehicles, may be regarded in effect as diffuse emission and in the same way can contribute to the widespread contamination of the environment with chemical mixtures. Thus, diffuse emission can comprise contributions from several emission sources and product emissions. In addition to exposure through environmental media, such as air, soil, and water, the indoor conditions of private households can be relevant for many airborne mixtures in relation to human health due to a large variety of products that are used indoors, where the ventilation can be limited. Also, food intake can be considered

for potential relevant mixture exposures for humans and for species in the higher tier of ecological food webs.

For the terrestrial environment, waste sites may act as major emission sources of mixtures. In the United States, the Agency for Toxic Substances and Disease Registry (ATSDR) has performed a trend analysis to identify priority chemical mixtures associated with hazardous waste sites (De Rosa et al. 2001, 2004; Fay 2005). The information was extracted from the Hazardous Substance Release/Health Effects Database (HazDat) (ATSDR 1997). The HazDat contains data from hundreds of hazardous waste sites in the United States. A trend analysis was completed for frequently co-occurring chemicals in binary or ternary combinations found in air, water, and soil at or around hazardous waste sites (Fay and Mumtaz 1996; De Rosa et al. 2001, 2004). Table 1.1 gives an overview of frequently occurring substances at hazardous waste sites in the United States.

1.2.2 EMISSION ESTIMATION METHODS

In the work of OECD (2006) on assessment of emissions, a distinction is made between Emission Scenario Documents (ESDs) and Pollution Release and Transfer Registers (PRTRs). An ESD provides a description of activities related to emissions and methods to estimate these emissions. A PRTR is an environmental database of potentially harmful chemicals released to air, water, and soil (on-site releases) and transported to treatment and disposal sites (off-site transfers). PRTRs contain data on releases or transfers, by source, and are publicly available in many countries, including Australia, Canada, Japan, several European countries, and the United States. An OECD study identified the similarities and differences between the emission estimation methods used in ESDs and PRTRs, showing that PRTR mass balance and emission factor methods yielded more conservative estimates than the ESD fixation-based method (OECD 2006). The PRTR mass balance method was found to account for a more thorough analysis of parameters, such as substance sources and recycles, which could impact emissions. Both ESD and PRTR methods might be applied to complex chemical mixtures, although no studies are available at present.

The emission estimation methods of the PRTR approach are described by OECD (2002a, 2002b, 2002c) and include direct monitoring, mass balance, emission factor, and engineering calculations and judgment. These methods are all feasible for the estimation of mixture emissions. The mass balance approach is based on the principle of mass conservation. Emissions from a system can be estimated by knowing the amount of a substance going into the system and the amount that is created or removed (dissipated or released to other compartments, degraded, transformed, or bound):

$$\Sigma(\text{output}) = \Sigma(\text{input}) - \Sigma(\text{removed}) + \Sigma(\text{generated})$$

For mixtures of chemicals, this equation should be used to estimate the concentration of each component under steady-state conditions, or under dynamic conditions when data are available to describe temporal conditions. These calculations lead to the (constant or varying) composition of the mixture over time.

Table 1.1 Frequencies of single substances and combination of substances at hazardous waste sites in the United States

Rank	Percent of sites	Single substance	Percent of sites	Binary combination		Percent of sites	Ternary (tertiary) combinations		
Water									
1	42.4	TCE	23.5	TCE	Perc	11.6	1,1,1-TCA	TCE	Perc
2	38.4	Lead	18.9	Lead	Chromium	10.6	Benzene	TCE	Perc
3	27.3	Perc	17.9	1,1,1-TCA	TCE	10.6	Lead	Cadmium	Chromium
4	25.8	Benzene	17.3	TCE	Lead	9.8	1,1,1-TCA	1,1-DCA	TCE
5	25.8	Chromium	17.3	Lead	Cadmium	9.7	Lead	Arsenic	Cadmium
6	23.9	Arsenic	17.0	Benzene	TCE	9.7	TCE	Perc	Lead
7	20.8	1,1,1-TCA	16.3	Lead	Arsenic	9.6	Lead	Arsenic	Chromium
8	20.3	Toluene	14.5	TCE	Trans-1,2-DCE	9.4	Benzene	TCE	Toluene
9	19.8	Cadmium	13.6	TCE	Toluene	9.3	TCE	Perc	Trans-1,2-DCE
10	17.7	MeCl	13.5	Benzene	Lead	9.1	TCE	Lead	Chromium
Soil									
1	37.7	Lead	20.5	Lead	Chromium	12.0	Lead	Cadmium	Chromium
2	25.3	Chromium	17.8	Lead	Arsenic	11.6	Lead	Arsenic	Chromium
3	23.0	Arsenic	17.6	Lead	Cadmium	10.9	Lead	Arsenic	Cadmium
4	19.7	Cadmium	13.3	Arsenic	Chromium	8.4	Arsenic	Cadmium	Chromium
5	19.1	TCE	12.9	Cadmium	Chromium	8.1	Lead	Nickel	Chromium
6	16.0	Toluene	11.6	Arsenic	Cadmium	7.9	Lead	Chromium	Zinc
7	14.8	Perc	10.9	TCE	Perc	7.7	Lead	Copper	Zinc
8	13.6	PCBs	10.9	Lead	Zinc	7.6	Toluene	Lead	Chromium
9	13.0	Xylenes	10.4	Ethylbenzene	Toluene	7.5	Ethylbenzene	Toluene	Xylenes
10	12.8	Ethylbenzene	10.4	Lead	Nickel	7.5	Lead	Nickel	Cadmium

Air									
1	6.0	Benzene	3.5	Benzene	Toluene	2.2	Benzene	TCE	Perc
2	4.7	Toluene	2.7	Benzene	TCE	1.9	Benzene	Ethylbenzene	Toluene
3	3.8	TCE	2.6	Benzene	Perc	1.8	Benzene	Toluene	Perc
4	3.4	Perc	2.6	TCE	Perc	1.8	Benzene	TCE	Toluene
5	3.1	1,1,1-TCA	2.3	Toluene	Perc	1.8	TCE	Toluene	Perc
6	2.6	Lead	2.1	Ethylbenzene	Toluene	1.4	1,1,1-TCA	Toluene	Perc
7	2.5	Ethylbenzene	2.1	TCE	Toluene	1.4	1,1,1-TCA	TCE	Perc
8	2.4	MeCl	1.9	1,1,1-TCA	TCE	1.3	Benzene	1,1,1-TCA	Perc
9	2.4	Xylenes	1.9	Toluene	Xylenes	1.3	Benzene	Toluene	Xylenes
10	1.8	Chloroform	1.9	1,1,1-TCA	Perc	1.3	1,1,1-TCA	TCE	Toluene

Source: Adapted from De Rosa CT, El-Masri HE, Pohl H, Cibulas W, Mumtaz MM. 2004. J. Toxicol. Environ. Health 7:339–350.

Note: MeCl = methylene chloride, PCBs = polychlorinated biphenyls, Perc = perchloroethylene (tetrachloroethylene), 1,1,1-TCA = 1,1,1-trichloroethane, TCE = trichloroethylene, Trans-1,2-DCE = trans-1,2-dichloroethylene, 1,1-DCA = 1,1-dichloroethane.

An emission factor is defined as a constant that relates the intensity of an activity to an emission (OECD 2002a). Emission factors can be used to estimate releases from nearly any source that generates emissions with a strong proportional dependence on the extent. Emission factors are used for specific cases where no release information is available, or if the release is only given for 1 specific compartment. The complementary release estimates can be obtained using the OECD approach or from the European Technical Guidance Document (ECB 2003a). The release of a chemical to an environmental compartment a is calculated as

$$\text{Release}_a = F_a \times \text{Prodvol}$$

That is, the release to the compartment (e.g., freshwater or air) is equal to the product of the fraction of the produced volume released, F_a , for example, during production, and the produced volume of the chemical (Prodvol). F_a is the emission factor. Emission factors can be applied to essentially any pollution or source (OECD 2006). They may be derived by many different techniques, but often are developed by taking the average emission rate during a representative time interval and relating it to the extent of the activity in question (OECD 1999). Emission factors can be used to estimate emissions of mixtures when data on the components are available. If the ratio between concentrations of different chemicals in the emission is stable, periodic monitoring of certain pollutants can be used to represent other pollutants by applying average ratios (OECD 2006).

More complex calculations can be made based on mathematical relationships between variables within a system, the outcome being dependent on the quality of data and the validity of assumptions. No specific models are available for chemical mixtures; however, modeling tools might be useful to estimate the varying composition of mixtures in time and space.

In Europe, ESDs are commonly used to facilitate the risk assessment of single substances, and they often deal with groups of chemicals. The methods used in ESDs are designed to deal with broader emissions than those used in PRTRs by dealing with emissions from clusters of facilities. This provides information on data on a local or regional basis. Although the ESDs are directed toward single chemicals, they may provide data for estimation of the emission of mixtures. Recently such information on emissions of single chemicals has been collected by the European Chemicals Bureau (ECB 2003b). The data are currently available for the different industrial categories (ICs) and biocidal product types. This documentation has been developed by different competent authorities and by industry. In most cases the documents are based on in-depth studies of the environmental release of substances used in different industrial categories and of different biocidal product types. Some documents describe environmental releases from specific use categories under an industrial category. Data are not available for all industrial categories and biocidal product types; some documents are still under preparation. It is anticipated that this set of emission scenario documents will be expanded continuously in the future. For industrial chemicals 9 areas have been developed so far: chemical, leather processing, metal extraction, photographic, textile processing, rubber, and paint industries, as well as personal or domestic and public domains.

Data on the emission of (mixtures of) chemicals may also be obtained from the European Pollutant Emission Register (EPER), which is the first European-wide register of industrial emissions into air and water (<http://eper.ec.europa.eu/> (last accessed November 2009)). EPER gives access to information on the annual emissions of approximately 9200 industrial facilities in the member states of the European Union as well as Norway mostly for the year 2001, and approximately 12,000 facilities for the year 2004. It has the option to group information by pollutant, activity (sector), air and water (direct or via a sewerage system), or country, and even gives access to data on individual facilities. Such information thus has value for developing realistic emission scenarios for diffuse release and also at the local scale.

Prediction of the composition of a chemical mixture that organisms are exposed to in a certain area requires considering all emission sources in that area or contributing to the input of chemicals in that area. The European Chemicals Bureau has described the relevant factors for estimating the release or emission of chemicals, including their intermediates and degradation products (ECB 2003a):

- the emission factor (release fraction) for processing of the intermediate,
- local production volume per time unit,
- the emission factor (release fraction) for production of the intermediate,
- the elimination in on-site treatment facilities, and
- the elimination in biological wastewater treatment facilities.

As an example, emission to surface water (in g/s) was estimated in a German scenario. The local concentration in rivers was calculated from that emission and the river flow (in m³/s), taking into account adsorption processes in the surface water. This approach was based on a statistically evaluated database. Volumes of wastewater flows from the production or processing facility were not taken into account. Although for the time being the database is restricted to a set of 29 substances and to German conditions, it can be regarded as a realistic worst-case situation since it combines two 90th percentiles (discharge × river flow) (ECB 2003a).

1.2.3 PRIORITIZATION

As described, living organisms are constantly exposed to vast amounts of chemicals, that is, to 1 big chemical mixture. We lack the information on how to properly capture the entire exposure, on how to address the toxicity, and how to evaluate the associated risk. Therefore, the initial approach is to properly define mixtures on a smaller scale. These are the “mixtures of concern” and are usually associated with a specific exposure scenario and possible health implications. Exposures to chemical mixtures of concern can range from simple and well-defined mixtures to complex and poorly defined mixtures. For example, morphine in combination with other epidural anesthetics is used in hospital settings for pain relief. The mixture can be characterized as simple (<10 chemicals), and in some cases such mixtures can be well defined because it is easier to identify the chemicals involved and know their dose, toxicity, and potential interactions. In contrast, complex mixtures are composed of many (>10) chemicals. Their composition may be largely known or at least reproducible (e.g.,

fumes from a specific paint), or the mixture may not be fully characterized either qualitatively or quantitatively, and may even vary from one similar exposure scenario to another (e.g., diesel fuels or gasoline from different sources). The sheer number of interactions means that the complete set of individual and interactive effects can never be established. Such cases thus present a particular challenge, and Chapter 4 discusses tools available to assessing the toxicity of such complex mixtures.

In the prioritization of chemicals for setting scenarios that are the most realistic and probable for chemical mixtures, tools are needed to overcome the almost infinite number of chemicals or combinations of chemicals and of differing concentrations in emissions. The global production of chemicals has increased from 1 million tons in 1930 to 400 million tons today. There are about 100,000 different substances registered in the EU market, of which 10,000 are marketed in volumes of more than 10 tons (produced per manufacturer or imported per importer per annum), and a further 20,000 are marketed at 1 to 10 tons (EC 2001). In the selection of chemicals for mixture scenarios, the volume of marketing is an important parameter; however, other factors, such as emission pattern (spatial and temporal), degradation, and toxicity, are also very important.

1.2.4 VALIDATION STUDIES

In many countries, emissions are determined by direct monitoring. In these cases the measurements should be subject to quality assessment, and the sampling plan should be evaluated to estimate the uncertainty in all steps of the procedure. To assess emissions of mixtures of chemicals, concentrations of chemicals should be measured simultaneously, but the validation procedures would be the same as for single chemicals. When emissions are estimated from data on produced, processed, or used amounts of the individual chemicals in mixtures, the calculations should be validated by measurements in the field.

1.3 INTERACTIONS AFFECTING AVAILABILITY AND EXPOSURE TO CHEMICAL MIXTURES

1.3.1 CHARACTERISTICS OF THE MAJOR ENVIRONMENTAL COMPARTMENTS

Identification of the most probable mixture exposure scenarios is a first step to establishing the nature of multiple chemical exposures, but even when such information is available, the interaction of the chemical constituents of the mixture with the environment needs to be considered. The physical and chemical nature of different environmental compartments has a large influence on the magnitude, duration, and stability of exposure (Table 1.2). On the one hand, chemicals in air are highly mobile and can travel over long distances, but also can be subject to rapid dilution through mixing. On the other hand, soils and sediments are immobile, and so the chemical released into these media can remain patchily distributed and dilution through mixing (e.g., by bioturbation) occurs at a very slow rate. In soils, sediments, and waters, the chemical properties of the media (such as pH, percentage of total and dissolved organic matter, cation exchange capacity, and concentrations of some ionic species) can have a large

Table 1.2 Physical characteristics influencing the duration and magnitude of exposure for the major environmental compartments

Media or compartment	Characteristics of the exposure
Soil	Immobile; hard to dilute; exposure, especially to persistent compounds, can be temporally quite constant, but spatially patchy; soil properties (pH, organic matter content, cat ion exchange capacity) have large effects on exposure (bioavailability) and can show spatial and temporal variability
Sediment	Immobile; hard to dilute; exposure, especially to persistent compounds, can be temporally quite constant, but spatially patchy; sediment characteristics can affect exposure; anoxia common and can affect exposure
Water	Mobile; can be diluted; pollutants can disperse within water column over moderate distances, pulsed exposure common; chemical characteristics can affect exposure, although these are less variable than, for example, soil
Air	Highly mobile, so pollutants can travel long distances (transboundary issues arise); composition quite stable at given altitude, and so local effect on bioavailability not such an important issue; rapid dissolution through mixing can occur; pulsed exposure common
Food chain for higher organism	The physiology of the species involved (both predator and prey) strongly influences the nature of exposure; exact exposure of an individual depends on the home range and dietary composition of the species or individual involved

influence on chemical availability. In ecotoxicology, a great deal of research effort has been focused on understanding the relationships between chemicals and soil, sediment, and water properties. The models and methods developed from this work are likely to be broadly applicable to exposure assessments for mixtures, although work is needed to validate this assumption. The importance of food chain transfer as an exposure route is greatly influenced by the properties of the chemical involved (e.g., persistence) and the physiology of the species within the particular food web. For example, Hendriks et al. (2001) modeled bioaccumulation of organic chemicals and Hendriks and Heikens (2001) modeled metal accumulation and demonstrated the need to include both chemical properties ($\log K_{ow}$ in case of organic chemicals) and species characteristics (body size, metabolic rate, trophic position, route of exposure) in order to allow for an effective assessment of bioaccumulation.

1.3.2 ENVIRONMENTAL FATE AFFECTING MIXTURE COMPOSITION

1.3.2.1 Single Compounds as Chemical Mixtures

Once released into the environment, selected single compounds can be subject to a range of transformations that may change the chemical identity of part or the entire amount of the original chemical released. Thus over time, even release of a single compound will result in the presence of a chemical mixture. The most obvious example of cases where a single chemical release results ultimately in the presence of a complex and changing mixture is in organic molecule (especially pesticide and biocide) degradation. There is an extensive literature that describes in detail many of the catabolic processes that can transform organic molecules in the environment, and it is not realistic to include anything other than a sample of such information.

When considering how such degradation affects exposure and potential effect, it is obviously tempting to assume that degradation of biologically active molecules such as pesticides will result in a lower toxicity of the metabolite-parent compound mixture than for the parent compound alone. While this is often the case, it certainly cannot be seen as a universal law. Thus, dichlorodiphenyl trichloroethane (DDT), which is already a highly persistent and bioaccumulative insecticide, is converted in organisms into the even more stable metabolites dichlorodiphenyl dichloroethylene (DDE) and dichlorodiphenyl dichloroethane (DDD). Thus, for this compound, the metabolite can represent an important component of the resulting mixture. The same principles apply for the pesticide aldrin, which is degraded in soil by microbial epoxidation to form the more persistent dieldrin (O'Halloran 2006). Such effects can be media specific depending on the prevalent processes. Thus, the organophosphate pesticide chlorpyrifos is degraded in air by reaction with OH^\bullet radicals to form different products, including chlorpyrifos oxon, which has a higher toxicity than the parent compound. The oxon may subsequently be converted into other products by side chain oxidations. In water, soil, and sediment, degradation of chlorpyrifos by photolysis or hydrolysis results in the formation of 3,5,6-trichloro-pyridinol (TCP). The importance of the metabolite as a component of the mixture depends on the speed at which conversion reactions occur. In the case of water, soil, or sediment, hydrolysis of the oxon metabolite leading to the formation of TCP is expected to take place faster than that of the thiol (parent) molecule, meaning that the metabolite is only likely to be present at low concentrations, and so have only a limited contribution to overall toxicity compared to other components (Cahill et al. 2003).

As well as organic molecules, metals can also be the subject of chemical and biological transformations that can change chemical identity and result in altered toxicity. The most well-known and well-studied example of this is the conversion of mercury (Hg^0) into methylated and ionic species (such as Hg^{2+} , CH_3Hg^+ , and $(\text{CH}_3)_2\text{Hg}$). Mercury is emitted to the environment from many different sources, usually as phenyl mercury, metallic mercury, and bivalent inorganic mercury. In anoxic sediments, the dominating form is sulfide (HgS), which is almost insoluble in water. Different bacteria (especially methane bacteria) as well as several species of fungi are capable of forming methyl mercury (CH_3Hg^+) and dimethyl mercury ($(\text{CH}_3)_2\text{Hg}$), which are soluble in water (Regnell 1994; King et al. 2002; Bisinoti and Jardim 2003).

These examples show that also in the case of single chemicals, such as pesticides and metals, exposure assessment should not only focus on the parent chemical but also include the metabolites and transformation products produced either in the environment or upon biotransformation in the organism.

1.3.2.2 Chemical Fate Effects on Mixture Composition

Once a mixture of chemicals is introduced into the environment, processes may take place that affect the composition of the mixture in space and time (see, e.g., Foster et al. 2005; Haws et al. 2006). When (complex) mixtures are released, differing fate properties of the individual chemicals can lead to a change in the exposure over time, and exposure in different environmental compartments and through different routes. An example is the distribution of gasoline over different environmental compartments:

depending on the compartment of first introduction and the properties of the different (groups of) components in the gasoline mixture, their distribution over the environment may differ (Foster et al. 2005). Properties of single chemicals determining their distribution and fate in the environment include physical–chemical properties, such as molar mass, boiling point, density, vapor pressure, water solubility, Henry's law constant, partition coefficients like the octanol–water partition coefficient (K_{ow}) and the octanol–air partition coefficient (K_{oa}), and degradation half-lives in different environmental compartments (air, water, soil, and sediment). In addition, bioaccumulation factors may be needed to determine partitioning in biological compartments (Foster et al. 2005).

The residence time of a compound in a mixture is not only determined by its fugacity (Mackay et al. 1992a), but also by its susceptibility for degradation. Persistency (usually indicated by its half-life) not only depends on the properties of the compound, but also on those of the environmental compartment (Haws et al. 2006). Since persistency of most organic compounds in soil, water, and sediment mainly relies on biological activity, it is important to focus on processes affecting biodegradation. It is assumed that biodegradation only takes place in the bulk aqueous phase of soil (or sediment). Processes affecting the sorption of chemicals (see above) determine the availability of chemicals in solution, and therefore play a key role in biodegradation. In addition, biological factors, such as microbial abundance and activity, and affinity for the contaminant, determine biodegradation rate. Such interactions can have both negative and positive effects on the degradation rates for particular compounds.

Two types of interaction with the potential to negatively affect degradation rates of chemicals in mixtures compared to single chemicals are competition and toxicity. When substrates are present, this can lead to the inhibition of biodegradation simply because more substrate molecules are competing for the same (and limited) number of active enzyme sites. This inhibition can be competitive in case of homologous chemicals, but also noncompetitive when 2 chemicals independently bind to the same enzyme, leading to a reduction of its overall utilization rate. It is also possible that the second chemical (inhibitor) binds to the enzyme complex but not to the free enzyme (Haws et al. 2006).¹ Toxicity to the microbial community can change biodegradation potential in cases where the species responsible for the catabolism of one compound are sensitive to the presence of another that is present in a mixture. Such interactions can potentially affect the ratio at which metabolites are formed or change the nature of degradation pathways (Haws et al. 2006).

As well as inhibition, some interactions in mixtures have the potential to stimulate degradation processes for a particular chemical. One example of an increased biodegradation rate in a mixture is as a result of increased biomass growth. This can occur, for example, when an easily metabolized substrate is present in conjunction with a more recalcitrant chemical. The rapid breakdown of the easily metabolized chemical supports the expansion of the degradative community, and this larger community is then better able to metabolize the less readily catabolized compound. This

¹ Similar principles of competitive, noncompetitive, and uncompetitive inhibition of metabolism will also be discussed in Chapter 2 in relation to toxicokinetics.

was the case with a mixture of naphthalene, phenanthrene, and pyrene. Compared to single compound studies, the mineralization rate of the easily degraded naphthalene was decreased compared to rates found for the single chemical alone, while that of the more persistent phenanthrene and pyrene was increased (Guha et al. 1999). The presence of solubilizing agents, such as biosurfactants, is a further means by which the degradation of chemicals present in combination with such chemicals may be increased. This effect has been shown for a number of PAHs (Guha et al. 1998). These biosurfactants, which enhance the bioavailability of the PAH, may, however, be toxic to the microorganisms responsible for PAH biodegradation, leading to a reduced degradation rate. This has been shown for phenanthrene (Shin et al. 2005) and demonstrates the potential complexities of the interactions that may occur.

1.3.3 AVAILABILITY

1.3.3.1 Availability and Bioavailability

Bioavailability is an important issue, not only regarding the transfer of contaminants within food chains, but also for a robust effect assessment of chemical mixtures in the environment. Pathways and mechanisms on how chemicals interact with each other within a mixture, how they enter the organism, and how they accumulate under defined circumstances need to be unraveled (CSTEE 2000). It is generally accepted that the total concentration of chemicals and chemical mixtures in all environmental compartments is not enough to predict biological or ecosystem effects. Bioavailability can be defined as the fraction of a chemical compound in a specific environmental compartment that, within a given time span, is or can be made available for uptake by organisms or is made available at the site of physiological activity (cf. Peijnenburg and Jager 2003). Within the same scope other words are often used in human health issues, such as availability and bioaccessibility; the latter one usually is defined as the fraction of a chemical that is capable of being used by a living organism. These 3 concepts can be considered synonyms, and from now on in this section the word “bioavailability” is used.

For mixtures, like for single chemicals, bioavailability is a key factor governing the magnitude and duration of exposure. Processes affecting the sorption of chemicals in, for example, soils and waters are vital in determining the availability of chemicals that enter the solution phase. Physical and chemical characteristics of the sorbent (particle size, organic matter content, pH) and sorbates (K_{ow} , ionization) are the major determinants of the bioavailability of single chemicals. Additionally, in cases where mixtures of chemicals are found, the presence of other chemicals may affect bioavailability (Haws et al. 2006). Competitive sorption especially may occur when chemicals have an affinity for similar sorption sites. This has been reported for phenanthrene and pyrene, with phenanthrene desorption increasing in the presence of pyrene (White and Pignatello 1999). Also, in case of metal mixtures, competitive sorption may occur: nickel sorption to soils decreased when other cations (including H^+) were present and was highest at neutral pH (Staunton 2004). And desorption of cadmium was increased by the presence of zinc, but zinc desorption was not increased by the presence of cadmium (Van Gestel and Hensbergen 1997).

In principle, sorption of chemicals in a mixture can be predicted from isotherms for sorption of the single chemicals, but such predictions may become less accurate for mixtures with significant interactions of the chemicals (Haws et al. 2006). Another aspect hampering a proper prediction of the sorption of single chemicals, as well as of mixtures, is the fact that sorption may change with time. Due to the process of aging, sorption of organic chemicals increases with time and is stronger than expected on the basis of physical-chemical properties (such as K_{ow}) or laboratory sorption experiments (Alexander 1995; Hatzinger and Alexander 1995; Kelsey and Alexander 1997). This also is the case for metals (Smit and Van Gestel 1998; Lock and Janssen 2003). Also, in case of changing redox conditions, sorption, of especially metals may no longer be directly predictable, due to the formation of sulfides that have a very low solubility (Lee and Lee 2005).

Although the effects of chemical mixtures in the environment on humans are thought to have important health implications, they are not widely studied or especially well understood. Their potential risks, therefore, have to be considered as a crucial environmental health problem requiring clear and rigorous future investigation. Examples pertaining to human exposure to chemical mixtures can be associated with emissions from point sources and diffuse and mobile sources, although because they often represent the most severe absolute exposure concentrations, the majority of published work is focused on local scale studies associated with industrial and mining releases. One such example is the study of Pereira et al. (2004) in an abandoned mine in the southeast of Portugal (S. Domingos mine) on integrated human and environmental risk assessment. Hair samples from the scalp were analyzed for metal presence (As, Cd, Cr, Cu, Mn, and Zn) in the human population living nearby the mine. High concentrations of Cd, Cu, and As were recorded in individuals living near the mine compared to individuals that live apart. The concentrations reported in the hair of this group of people were above the reference values. It was also concluded in this study that metal concentrations were related to concentrations in soil, probably related to the consumption of milk and cheese from cattle from the region. In a further local scale study around an industrial facility, Cui et al. (2005) showed that Cd intake through vegetables was higher in samples from one village closer to a factory (500 m) than in samples from another village located at 1500 m. Cd concentration in urine and serum from residents was, however, much lower in the village closer to the factory. The authors suggested that the possible reason for this was the higher intake of Fe, Ca, and Pb from vegetables of the far-away village, suggesting also that high intake of these ions could lead to a decrease in Cd body burdens. This therefore indicates clearly that, like for environmental species, it is not the absolute concentration of a chemical that is present in the exposure medium, but rather the concentration that is actually available for uptake that is the primary determinant of exposure.

Oomen et al. (2003) defined 4 steps in the oral bioavailability of chemicals present in contaminated soils to man: soil ingestion, mobilization from soil during digestion (i.e., bioaccessibility), absorption from the intestinal lumen, and first-pass effect. An *in vitro* model of the human digestive system was used to study the uptake of chemicals from ingested soil. When an artificial soil, spiked

with a mixture of polychlorinated biphenyls (PCBs) and lindane, was introduced into the system, approximately 35% of PCBs and 57% of the lindane appeared to be accessible. Bioaccessibility was explained from the distribution of these chemicals over the soil solid phase, bile salt micelles, and proteins (Oomen et al. 2000). It turned out that the first step in the uptake process, the mobilization of the chemicals from soil, was most important in determining the uptake flux, and therefore the bioavailability of these contaminants in ingested soil particles (Oomen et al. 2001). Other studies demonstrated that from an artificial soil dosed at 530 mg Pb/kg, only 23% of the Pb was bioaccessible, and that only a part of the bioaccessible soil-borne lead was actually taken up (Oomen et al. 2003). These studies demonstrate that the composition of a mixture of chemicals originally present in ingested soil particles may change in the course of the uptake process, affecting exposure.

This example suggests that organisms may also affect the sorption equilibrium of chemicals in soil or sediment. Another example of that was found in the uptake of cadmium in *Hyallela azteca*. In water-only exposure, cadmium uptake and toxicity in this organism were not affected by phenanthrene. When exposed in sediment, however, cadmium uptake was increased when the animals were simultaneously exposed to phenanthrene. This most likely is a result of associated feeding, causing alterations in ingestion or digestive processes, leading to an increased cadmium uptake. So, from a toxicological point of view, the synergism seen cannot be explained from an interaction in the animals but rather is exposure related (Gust and Fleeger 2005).

1.3.3.2 Influence of Medium Physical–Chemical Properties on Chemical Availability

In case of metal mixtures in soils and sediments, bioavailability is controlled by the strength of binding of metal ions to the soil or sediment particles. The partitioning of metals to the soil or sediment reduces the availability for mobilization and uptake by plants, animals, and microbes. The binding strength depends highly on properties that control the partitioning process, such as pH, dissolved organic matter (DOM), and other organic ligands, calcium, organic matter content, inorganic ligands, and the solid-phase metal oxide (Allen 2002).

The partitioning of metal ions can be modified in the digestive track of soil-dwelling organisms or in the rhizosphere. Depending on the organism, bioavailability can be related to several key factors. In plants and microorganisms, it has been correlated to the activity of the free metal ion and its presence and diffusion in the soil pore water. Bioavailability of metals in invertebrates can be influenced by the organic matter content that is digested in their guts, and might be physiologically moderated by pH and competing cations (e.g., Ca^{2+}).

Soil or water pH is one of the most important parameters when dealing with metal bioavailability. Also for ionizable organic chemicals, like chlorophenols, pH may affect bioavailability. Changes of conditions in environmental compartments may depend on their surrounding environments and climatic changes, and pH is one of those properties that can show larger changes.

In aquatic systems, in addition to the complexation of metal ions by natural organic matter, metal bioavailability, bioaccumulation, and toxicity are highly affected by water hardness and alkalinity (Banks et al. 2003). This is also applicable to metal mixtures where complexation of metals can occur even at higher rates than when single chemical compounds are present.

Bioavailability of organic chemicals is strongly dependent on aqueous solubility. The equilibrium partitioning theory has been applied to sediment toxicity studies, and it was concluded that uptake from sediment as well as from (pore) water is possible at the same time; however, the exposure route in equilibrium is not necessarily important. For substances with $\log K_{ow} < 5$, the equilibrium partitioning theory is considered acceptable to assess the risk. For substances with $\log K_{ow} > 5$, a safety factor of 10 is applied, in order to include the additional uptake by sediment ingestion (Loonen et al. 1997).

Bioavailability can vary with the contact time of chemicals with soil or sediment particle constituents. Chemicals in newly deposited sediments may become more bioavailable than older buried materials. In soils the same can happen. In a study where a 70-year-old and a freshly copper-contaminated soil were compared, in the soil contaminated 70 years ago no copper toxicity to *Folsomia fimetaria* was observed for concentrations of copper as high as 2911 mg/kg. The newly spiked soil, however, caused a 10% decrease in reproduction at 337 mg Cu/kg (Scott Fordsmand et al. 2000). In the work of Smit and Van Gestel (1998), aging was also shown to be important to understanding the toxicity of Zn to *F. candida*. Within time soil pH can change, which also induces a change of Zn toxicity by altering Zn sorption to soil particles (e.g., an increase of soil pH leads to an increased Zn sorption). For chemical mixtures the same may happen due to the effect of soil characteristics on chemical adsorption to soil particles and also to chemical interactions through aging. For organic chemicals, it was suggested that molecules slowly become sequestered within the soil matrix and therefore become less available to organisms. Besides abiotic parameters, biotic interactions also may play a role. Bacteria, fungi, and soil invertebrates may alter the behavior of chemicals in soils, altering also their persistence and sorption to soil particles. These facts are crucial when evaluating soil toxicity and risk assessment for decision makers and cleanup purposes (Alexander 1995).

When exposed to mixtures, chemicals in the exposure medium may affect each other's uptake by humans in a manner that is analogous to some of the bioavailability effects outlined here for environmental species. This was, for instance, shown for the neurotoxicity of EPN (O-ethyl-O-4-nitrophenyl phenylphosphonothionate), which was enhanced by aliphatic hydrocarbons due in part to increased dermal absorption (Abou-Donia et al. 1985). It was also shown that dietary zinc inhibits some aspects of lead toxicity, which could in part be explained by decreasing dietary lead absorption (Cerklewski and Forbes 1976). Other examples of interactions of chemicals at the uptake phase in humans, which may in part be related to bioavailability interactions, are summarized in Table 1.3.

1.3.3.3 Metal Speciation Determines Bioavailability

To understand chemical exposure and bioavailability the biotic ligand model (BLM) was developed for single (cationic) metal species, assuming that the amount of a

Table 1.3 Examples of interactions of chemicals at the uptake and absorption level in humans

Antimicrobial agent	Other drug	Mechanism
Decreased absorption		
Lincomycin	Kaolin-pectin	Irreversible adsorption to kaolin-pectin
Lincomycin	Cyclamate	Possibly complexation
Ritampicin	PAS “granulates”	Adsorption to bentonite granules
Tetracycline	Sodium bicarbonate, cimetidine, bi- and trivalent metal cat ions (including anatides), atropine	Alkaline pH inhibits dissolution, probably reduced dissolution, chelation, slowed gut peristalsis
Pivampicillin PAS	Diphenhydramine	Slowed gut peristalsis
Neomycin	Digoxin	Induced malabsorption
Neomycin	Warfarin	Induced malabsorption
Neomycin	Penicillin V	Induced malabsorption
Increased absorption		
Tetracycline	Metoclopramine	Increased gut peristalsis
Pivampicillin	Metoclopramine	Increased gut peristalsis

Source: Based on Calabrese, EJ. 1991. Multiple Chemical interactions, Part 4: Drugs; Part 5: The drug–pollutant interface. Chelsea (MI): Lewis Publishers, p 389–578.

metal binding to a sensitive biological ligand determines its toxicity. This fraction binding to the biological membrane is considered the bioavailable fraction. In addition to metals, other cationic elements may bind to the same target sites on the biological membrane. On the other hand, concentration and activity of free metal ions is determined by the presence of organic and inorganic ligands. The BLM incorporates the competition and affinity to the site of toxic action on the organism of the free metal ion, other naturally occurring cations, and also the possible complexation by abiotic ligands (e.g., dissolved organic matter, chloride, carbonates, sulfide). On this basis, the model includes ion uptake pathways that can quantify chemical characteristics like metal affinity and capacity in vivo. In general, the greater the affinity to the binding sites, the higher the toxicity of a particular metal. The BLM was also applied to mixtures to evaluate how it would respond, using the classic toxic unit’s concept of additivity. Ion competition has been included from the beginning in the BLM because cat ions like Ca^{2+} and H^{+} are known to decrease metal accumulation at the ligand (Playle 2004).

As an example, in the study of Sanchez-Dardon et al. (1999) exposure of the rainbow trout (*Oncorhynchus mykiss*) to Zn decreased Hg and Cd toxicity, probably by competition at the entry sites to gills’ cells, through competition at intracellular binding sites or through induced synthesis of metallothionein. Another example is with *Daphnia magna* exposed to Zn that increased their tolerance toward Cd, probably through a competition at the ligand site in the gut or through an induction of metallothionein synthesis (Barata et al. 2002). These studies are 2 examples of possible

competition between ions to bind in the uptake and intracellular sites in organisms. See also Chapter 2 for a discussion of BLM-type interactions in relation to toxicokinetics and toxicodynamics.

1.3.3.4 Species Specificity

Bioavailability is also considered species and organ or tissue specific because what is available to one species might not be the same for the other, and the same counts for different organs or tissues inside a living organism. Barahona et al. (2005) found that physiological differences among oat, wheat, and sunflower roots, such as root waxes, might be responsible for differences in permeability for nonpolar compounds and therefore for differences in sensitivity of the root elongation process.

Another example for species-specific differences in bioavailability of chemical mixtures is the study of Loureiro et al. (2005), where avoidance behavior of earthworms and isopods was studied in 2 soils from the vicinities of an abandoned gold mine. Isopods (*Porcellionides pruinosus*) turned out to show avoidance behavior, whereas earthworms (*Eisenia andrei*) did not, suggesting isopods to be more sensitive than earthworms for these soils that contained a mixture of metals. It remains unclear which factors explain this difference, but routes of exposure and bioavailability difference may have played a role.

Since bioavailability is an integral factor in the estimation of the internal dose (or dose at target tissue) of the chemical, it is important in human studies to consider the environmental and physiological characteristics of uptake sites and their interactions since these are clearly important for defining the extent of exposure. When exposure is through food, the gastrointestinal tract and its physiology has an important effect on the amount of different chemicals that are taken up. It has been suggested, for example, that gut uptake of 2,3,7,8-tetrachlorodibenzodioxin (TCDD) and related compounds is variable, incomplete, and congener and vehicle specific, and that more lipid-soluble congeners, such as 2,3,7,8-tetrachlorodibenzofuran, are almost completely absorbed, while the extremely insoluble octachlorodibenzodioxin is less well absorbed, depending on the dosing regimen. The fact that high doses may be absorbed at a lower rate, whereas low repetitive doses may be absorbed at a greater rate, thus has the potential to alter internal exposure from the assumptions based on external exposure concentrations. To date, the only study of TCDD bioavailability in humans was reported by Poiger and Schlatter (1986) and was based on a single male in which the gastrointestinal absorption was >87% when TCDD was administered in corn oil. Laboratory data suggest that there are no major interspecies differences in the gastrointestinal absorption of dioxins and dibenzofurans. However, absorption of TCDD is dependent on conditions and characteristics of the soil medium. In animals, absorption of TCDD from different soils ranged from 0.5% (Umbreit et al. 1986a, 1986b) to 50% (Lucier et al. 1986). Absorption from a diet was 50 to 60% in rats (Fries and Marrow 1992). Therefore, exposure with food rather than with oil as a vehicle relates more closely to exposure from soil. Bioavailability has to be considered when calculating the hypothetical ingestion dose.

1.3.3.5 Formulating Agents

Another important issue on the bioavailability of chemical mixtures is the use of formulating agents for several purposes (e.g., agriculture, veterinary, human health). Some studies have been performed to compare the toxicity of pure compounds and formulations. Azadirachtin, a plant-derived extract, is often used as a repellent, anti-feedant, and molt regulator for several insect species and can be applied as several formulations, like Azantin-EC, Bioneem, or Neemix. When exposed to the pure compound and 2 formulations, the cladoceran *Daphnia pulex* showed clear differences in sensitivity. LC50 values for the formulations Bioneem and Neemix were 0.07 and 0.03 $\mu\text{g/L}$, respectively, while a 50- to 100-fold higher value was found for the toxicity of the pure compound (0.382 $\mu\text{g/L}$). The higher toxicity of both formulations suggests that azadirachtin is not the only active compound in the formulations, or that the inert ingredients significantly enhanced the toxicity of azadirachtin. However, it remains unclear whether this is due to a higher bioavailability in the formulation, although it is suggested that the formulation may also affect fate (sorption, biodegradation) of azadirachtin (Goktepe and Plhak 2002).

Garcia-Ortega et al. (2006) reported conflicting results for the fate and effect of pure (Pestanal) propetamphos and its formulation Ectomort Centenary (8% active ingredient) in sediments. Sorption of propetamphos to sediments was stronger in the formulation. Biodegradation rate decreased with increasing sorption but was not affected by the formulation. Toxicity to sediment microbial communities was, however, significantly greater for the commercial formulation than for the pure compound. This cannot be explained from a higher availability of propetamphos in the formulation. Garcia-Ortega et al. (2006) therefore assume that ingredients of the formulation enhance toxicity of the active ingredient.

These studies do not confirm the effect of formulations on the bioavailability of compounds, but they do emphasize the need to test formulations (next to or instead of active ingredients) when performing a risk assessment of commercial pesticides (Garcia-Ortega et al. 2006).

1.3.3.6 Analytical–Chemical Procedures

Several methodologies have been developed for bioavailability assessment using chemical-analytical procedures. To determine the bioavailable fraction of metals or organic compounds in soils and sediments, several methods can be applied, such as extraction techniques. Usually desorption procedures are applied, using an extracting agent to desorb chemicals from the soil or sediment solid phase. An aqueous extraction might be considered the simplest alternative, although with some limitations. Deionized water has a very low ionic strength compared to natural (pore) water. That is why a weak salt extraction, for example, using 0.01 M CaCl_2 (Houba et al. 1996), is sometimes preferred, because it better simulates the soil solution. Such water and neutral salt extractions can be used to determine partition coefficients (K_d).

Specific extractants and sequential fractionation are also widely used procedures to estimate metal or nutrient availability to plants (see, e.g., Houba et al. 1996). Several extractants can be considered, like diethylene triamine penta acetate (DTPA), ethylene diamine tetra acetic acid (EDTA), acetic acid, HNO_3 , HCl , or other

mineral acids (Allen 2002). Each reagent might be considered specific for extracting a certain fraction of metals. As an example, EDTA and DTPA are often used as extractants of exchangeable and organically bound trace metals and also to dissolve metal precipitates. These procedures can also be considered as chelating extractions, showing a correlation between water and total digestion extractions. Another technique also used for the identification of the solid phase associated with metals is an x-ray absorption fine structure methodology (EXAFS) (Manceau et al. 2003).

Recently, attempts have been made to develop biomimetic methods, simulating plant uptake of metals. An example of such a method is DGT (diffusive gradients in thin films), developed by Zhang et al. (2001), for measuring metal availability to plants. In this case, metal accumulation in a chelex layer is measured. By taking into account thickness of the diffusive layer covering the chelex layer and contact time with the soil sample, it is possible to estimate the available metal concentration in the soil solution. The DGT method may also be used to estimate metal speciation in surface water (Zhang 2004).

Similar biomimetic methods have been developed for assessing the (bio)availability of organic chemicals in water, sediments, and soil (Mayer et al. 2003; Ter Laak et al. 2006). The main advantage of these nondepletion techniques, such as the solid-phase microextraction methods (SPMEs), is that they may be used without disturbing the chemical distribution in surface water or soils or sediments.

1.3.4 CHEMICAL–CHEMICAL INTERACTIONS IN MIXTURES

Another form of interaction that may change the nature of exposure for chemicals in mixtures is through direct chemical-chemical interaction. One example is the formation of nitrosamines (which are carcinogenic) from noncarcinogenic nitrates and amines in the stomach (Klaassen 1996). The formation of N-nitrosoatrazine from atrazine and nitrite has been demonstrated in vitro in human gastric juice (pH 1.5 to 2.0) during 1.5 to 12 hours of incubation at 37 °C (Cova et al. 1996). The percent formation peaked at 3 hours, and gradually declined thereafter, due to degradation of N-nitrosoatrazine to atrazine. Peak formation of N-nitrosoatrazine was 2% from 0.05 mM atrazine and 0.5 mM nitrite, 23% from 0.05 mM atrazine and 3 mM nitrite, and 53% from 1 mM atrazine and 3 mM nitrite. The formation of N-nitrosoatrazine from atrazine and nitrite also has been demonstrated in vivo, the amount formed being dependent on the ratio of atrazine and nitrite concentrations and on pH (Krull et al. 1980).

1.4 ENVIRONMENTAL FATE MODELING

Environmental fate models make use of chemical properties to describe transfer, partitioning, and degradation (Mackay et al. 1992a; Cahill et al. 2003). For organic chemicals, quantitative structure-property relationships (QSPRs) may be used to predict partitioning from physical–chemical properties, such as K_{ow} and K_{oa} . Such properties may also allow for a prediction of the transfer of chemicals between compartments. Recently, some successful attempts have also been made to predict persistency of chemicals (Raymond et al. 2001), although this mainly concerns

degradation rate under standardized conditions (Posthumus et al. 2005). But in most cases, specific knowledge on degradation pathways is required to predict the formation of metabolites. OECD (2004) presented an overview of multimedia fate models to predict overall environmental persistence (P_{ov}) and the potential for long-range transport (LRTP) of organic chemicals. P_{ov} and LRTP derive from both chemical properties and environmental conditions. Multimedia fate models can be used during exposure assessment to identify spatial extent of exposure, environmental partitioning (media of concern), and residence time of chemicals in a certain environmental compartment. Four levels of model complexity were identified, ranging from closed systems at equilibrium to dynamic open systems. OECD (2004) identifies generic multimedia models, region-specific multimedia fate models, and multizone multimedia models (Figure 1.1). These models may be helpful in describing distribution on a global scale (Toose et al. 2004), on a regional scale (Mackay et al. 1992a), or on a smaller scale, such as in surface water (e.g., the Exposure Analysis Modeling System (EXAMS); Schramm 1990).

The input required by multimedia fate models includes properties of the chemicals (such as distribution over compartments air, water, and soil or sediment), properties of the environment or landscape receiving the contaminants, and emission patterns and mode of entry of chemicals into the environment (OECD 2004) (Figure 1.1). Fenner et al. (2005) compared the outcome of 9 multimedia fate models by applying them to a set of 3175 hypothetical chemicals covering a range of 25 half-life combinations (in water, air, or soil or sediment) and 127 combinations of partition coefficients (air-water (K_{aw}), K_{ow} , and K_{oa}). Results show great similarities between the model outputs for P_{ov} predictions, but less for LRTP. P_{ov} and, to a lesser extent,

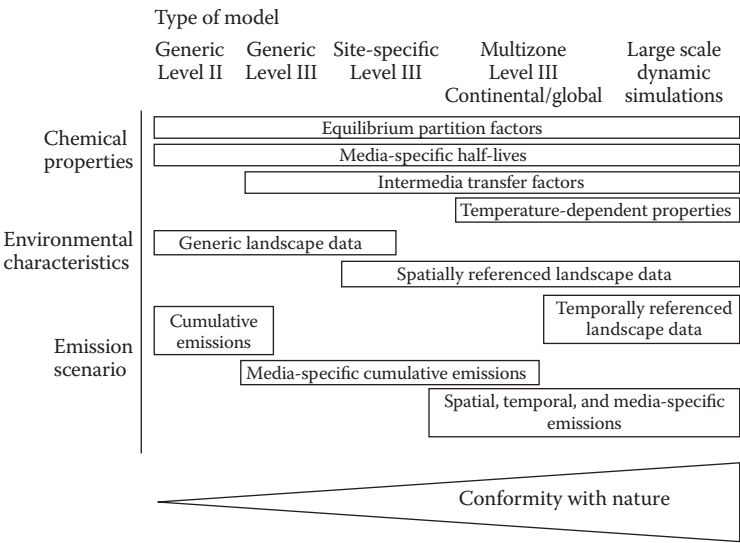


Figure 1.1 The continuum of multimedia fate models available for estimating overall persistence (P_{ov}) and potential for long-range transport of chemicals (Redrawn from OECD [2004]).

L RTP depend mainly on chemical properties. Models show significant differences in certain regions of the chemical space, with model uncertainty being higher than parameter uncertainty in case of L RTP predictions, and vice versa for P_{ov} . On the basis of this analysis, Fenner et al. (2005) conclude that it always is best to not rely on just 1 model but apply more than 1 model. Model selection is not an arbitrary task but requires careful consideration of the question and context of the assessment. Fenner et al. (2005) therefore provide guidance for selecting the most suitable model for a certain task.

So far, the main focus of multimedia fate models has been on single chemicals, but extensions may become available to include fate of transformation products. This may open the way to making the models applicable to mixtures (OECD 2004). Initially such development may simply be made through the serial analysis of the fate of individual chemicals, and from this a derivation of probable concentrations of each, assuming no interaction. Such analysis is, for example, feasible for many of the most widely used “down the drain” and is at present being extended to other product types, such as personal care chemicals and human pharmaceuticals. Such combined analysis would in fact represent a considerable step forward in addressing the nature of likely mixture exposures; however, if the interactions with the environment and between chemicals as outlined above are to be considered, then this would require a considerable effort to understand and include the major processes involved within existing models.

1.5 EXPOSURE SCENARIOS AND MONITORING

An important issue in pollution management is the identification of exposure scenarios that pose relatively high risks to humans or the environment (Thomsen et al. 2006). The regulation of these high-risk scenarios should be given priority in order to realize an effective reduction of the overall risks. An exposure scenario can be broadly defined as a set of parameters that together determine the risk level. This may include parameters such as the emission load, the emission compartment, fate parameters (persistence, biodegradability, vapor pressure), and toxic properties. The exposure scenario is thus the set of parameter values that needs to be known as a condition to estimate the risk. Thomsen et al. (2006) have described a new methodology for the identification of exposure scenarios that cause high risks for the environment or human health. They define a scenario type as the set of criteria that is considered important after analysis of a problem tree. A criterion has to be described by data, and thus empirical knowledge, before it can be taken into account as a condition for scenario selection. A data type that describes a specific criterion is denoted a descriptor. A specific scenario is defined as a combination of realistic descriptor values. In this procedure, descriptors for emission are consulted (descriptors for different product groups), as this can be seen as first screening for potential mixtures of relevance for further studies.

According to this new system, a harmful mixture consists of substances that together reach the target on 1 side and also have a toxic action that has potential to yield harmful effects. Realistic mixtures can thus be defined based on 1 of 2 principal general approaches:

- 1) toxicity-driven approach watching for common harmful toxicity of mixtures and thus based on the mode of action, where the combination of substances is identified that yields a high combined toxicity as a result of either additive or synergistic mixture toxicity mechanisms; and
- 2) exposure-driven approach watching for simultaneous exposure, where substances that can cause simultaneously exposure are investigated for combined toxicity.

It is not possible to claim one of these approaches as best because they are complementary to each other and a combination of them is ideal if possible. The method takes concentration addition (CA) as the starting point for mixture toxicity, so that the same risk descriptors can indicate toxic actions that add to each other between different substances (Thomsen et al. 2006).

1.5.1 HUMAN EXPOSURE

The mere presence of any single chemical or chemical mixture in the environment does not indicate that a health threat exists. An important step of mixture risk assessment is the evaluation of completed exposure pathways. Completed exposure pathways link together the source of contamination, environmental medium, point of exposure, route of exposure, and a receptor population. It means that without the potential for chemicals actually entering (or contacting) the human body, no threat is present.

Personal behavior and practices also add a further level of complexity to the estimation of individual total exposure for humans. Many publications pointed out harmful effects of high-dose alcohol consumption, smoking, drugs, or the use of mercurials in religious practices, just to name a few. All these chemicals contribute to overall exposures and may affect the toxicity of other chemicals entering the human body (Calabrese 1991). Such personal exposure patterns can also be overlaid on top of more regionally based environmental exposure resulting from diffuse environmental pollution. In this section, first, different pathways of human exposure are considered as well as life-stage-related exposures. Second, the use of monitoring data to assess mixture exposure is discussed.

1.5.1.1 Environmental Exposures Excluding Food

Environmental exposures are present through the human lifetime. However, they may vary considerably over time at the same location, for example, because of the local or global changes in emission and environmental pollution levels. Environmental exposures of humans consist of exposures outdoors and indoors as well as at workplaces; these environments may significantly differ. The exposure media include air, water, and soil and dust. Historically, research on human exposures to chemicals and associated health effects has been conducted mostly on single chemicals. In addition, several studies have dealt with complex mixtures, such as diesel fuel and gasoline, by-products from coal combustion, and tobacco smoke. A common problem of complex mixtures is that the composition may vary from one exposure to another and, as a result, the associated toxicity may vary. For a better understanding

of joint toxic action of chemicals and their effects on human health, it is important to identify combinations of chemicals that represent the most frequently occurring simple mixtures.

Some studies have attempted to address this need. For example, further analyses of the ATSDR data on mixtures at and around hazardous waste sites (see Section 1.2) considered completed exposure pathways (Table 1.4) (De Rosa et al. 2004; Fay 2005). The results show that the number of sites with mixtures in completed exposure pathways is lower than the number of sites for which only the frequency of co-occurrence was analyzed. Data from 1706 hazardous waste sites indicated that completed exposure pathways exist at 743 (44%) of the sites (Fay 2005). Of these, 588 had 2 or more chemicals in the completed exposure pathway. That means that exposure to mixtures occurred at 79% of the sites with exposure. As indicated in Table 1.4, mixtures of inorganic chemicals were found predominantly in soil, mixtures of organic chemicals were detected in air, and combinations of both in water.

A study of the US Geological Survey identified mixtures of chemicals in groundwater used for drinking water in the United States (Squillace et al. 2002). Samples were analyzed from 1255 domestic drinking water wells and 242 public supply wells between 1992 and 1999. Water in 11.6% samples did not meet current drinking water standards or human health criteria established by the US Environmental Protection Agency (USEPA). Volatile organic compounds (VOCs) were detected in 44% of the samples, pesticides in 38%, and nitrate in 28%. Many mixtures (i.e., possible combinations of chemicals) were found in the samples; however, only 402 mixtures were detected at least 15 times (>1% detection frequency). From all the samples, 47% contained at least 2 analyzed compounds and 33% contained at least 3 compounds. A list of the top 25 most frequently detected mixtures is shown in Table 1.5. Since the study was done on drinking water, human exposure can be assumed. In addition to these compounds in groundwater, drinking water may also contain a complex mixture of products resulting from disinfection, including trihalomethanes, haloacetic acids, haloacetonitriles, and bromate (Teuschler et al. 2000).

The USEPA's Total Exposure Assessment Methodology (TEAM) studies found levels of about a dozen common organic pollutants to be 2 to 5 times higher inside homes than outside, regardless of whether the homes were located in rural or highly industrial areas (USEPA 2006b). Evidence for the contribution of indoor air pollution to human exposure to mixtures of chemicals has also been obtained for other areas in the world, for instance, through the project Towards Healthy Air in Dwellings in Europe (THADE), which was sponsored by the European Union (Franchi et al. 2006). While the consumers are using products containing organic and inorganic chemicals indoors, not only can they expose themselves to high chemical concentrations, but increased concentrations can also persist in the air long after the activity is completed. Among the chemicals often found inside are carbon monoxide, nitrogen dioxide, formaldehyde, methylene chloride, and tetrachloroethylene. Carbon monoxide is generated as a product of incomplete combustion from sources, which include home furnaces and fireplaces. Similarly, nitrogen dioxide may be found in houses with poorly vented fireplaces and furnaces. Formaldehyde is found in many products used around the house, such as antiseptics, medicines, cosmetics, dishwashing liquids, fabric softeners, shoe-care agents, carpet cleaners, glues, adhesives, lacquers,

Table 1.4 Chemical mixtures in completed exposure pathways at and around hazardous waste sites in the United States

Rank	No. sites	Binary combinations		Rank	No. sites	Binary combinations	
Water							
1	120	TCE	Perc	10	45	Chloroform	TCE
2	64	1,1,1-TCA	TCE	12	42	TCE	1,2,-DCA
3	58	1,1,-DCE	TCE	13	40	1,1,1-TCA	1,2,-DCA
4	55	Benzene	TCE	13	40	1,1,1-TCA	1,2,-DCE
5	54	TCE	Lead	13	40	TCE	Trans-1,2-DCE
6	51	1,1,1-TCA	Perc	16	39	Lead	Cadmium
7	49	1,1-DCA	TCE	16	39	Perc	Lead
8	47	1,1,-DCE	Perc	18	38	Vinyl Chloride	TCE
9	46	TCE	Toluene	19	37	1,1-DCA	Perc
10	13	Lead	Arsenic	19	37	MeCl	TCE
Soil							
1	60	Lead	Arsenic	10	34	Lead	Nickel
2	56	Lead	Chromium	12	33	Copper	Zinc
3	52	Lead	Cadmium	13	32	Arsenic	Zinc
4	47	Arsenic	Chromium	13	32	PCBs	Lead
5	46	Arsenic	Cadmium	15	30	Cadmium	Copper
6	44	Lead	Zinc	16	29	Nickel	Chromium
7	39	Cadmium	Chromium	17	28	Antimony	Arsenic
8	38	Cadmium	Zinc	17	28	Arsenic	Copper
9	36	Lead	Copper	17	28	Chromium	Copper
10	34	Chromium	Zinc	27	28	Lead	Antimony
Air							
1	18	Benzene	Toluene	11	10	1,1,1-TCA	Toluene
2	16	Benzene	TCE	11	10	1,1,1-TCA	TCE
3	15	Benzene	Perc	13	9	Benzene	Xylenes
4	15	TCE	Perc	13	9	Ethylbenzene	Perc
5	14	Benzene	Ethylbenzene	15	8	1,1,1-TCA	Perc
6	13	Ethylbenzene	Toluene	15	8	Benzene	Chlorobenzene
7	12	Benzene	1,1,1-TCA	15	8	Benzene	MeCl
7	12	TCE	Toluene	15	8	Ethylbenzene	Chlorobenzene
9	11	Toluene	Perc	15	8	TCE	Ethylbenzene
10	11	Toluene	Xylenes	20	7	1,1,1-TCA	Ethylbenzene

Source: Adapted from De Rosa CT, El-Masri HE, Rohl, H, Cibulas W, Mumtaz, MM. 2004. J. Toxicol. Environ. Health 7:339–350.

Note: Binary combinations at the 1188 sites surveyed. MeCl = methylene chloride, PCBs = polychlorinated biphenyls, Perc = perchloroethylene (tetrachloroethylene), 1,1,1-TCA = 1,1,1-trichloroethane, TCE = trichloroethylene, Trans-1,2-DCE = trans-1,2-dichloroethylene, 1,1-DCA = 1,1-dichloroethane, 1,1-DCE = 1,1-dichloroethene.

Table 1.5 Top 25 most frequently detected mixtures in groundwater used for drinking water in the United States

Rank	Compounds in mixture				No. samples (out of 1497) with mixture
1	Atrazine	Deethylatrazine			284
2	Deethylatrazine	Nitrate			214
3	Atrazine	Nitrate			198
4	Atrazine	Deethylatrazine	Nitrate		179
5	Atrazine	Simazine			138
6	Deethylatrazine	Simazine			127
7	Atrazine	Deethylatrazine	Simazine		120
8	Nitrate	Simazine			111
9	Atrazine	Metolachlor			103
10	Deethylatrazine	Metolachlor			99
11	Deethylatrazine	Trichloromethane			97
12	Atrazine	Prometon			96
13	Atrazine	Deethylatrazine	Metolachlor		95
14	Atrazine	Nitrate	Simazine		92
15	Deethylatrazine	Nitrate	Simazine		92
16	Deethylatrazine	Prometon			90
17	Atrazine	Deethylatrazine	Prometon		87
18	Nitrate	Trichloromethane			86
19	Tetrachloroethene	Trichloromethane			86
20	Atrazine	Deethylatrazine	Nitrate	Simazine	86
21	Atrazine	Trichloromethane			78
22	Metolachlor	Nitrate			76
23	Nitrate	Prometon			73
24	Deethylatrazine	Metolachlor	Nitrate		71
25	Atrazine	Metolachlor	Nitrate		70

Source: Adapted from Squillace PJ, Scott JC, Moran MJ Nolan T, Koplin DW. 2002. Environ. Sci. Technol 36:1923–1930.

plastics, and some types of wood products. Methylene chloride is widely used as an industrial solvent and as a paint stripper and can also be found in certain aerosol and pesticide products, some spray paints, and automotive cleaners. Tetrachloroethylene may be found in the home environment as a result of dry cleaning of textiles. Another important group of indoor contaminants consists of pesticides. For example, chlorpyrifos, an organophosphorus insecticide, is the most widely used insecticide for indoor and outdoor residential applications in the United States (ATSDR 1997). Based on the monitoring of different outdoor and indoor media, a study indicated that indoor dust and air were the primary exposure media for the residents (Whyatt et al. 2002). Similar results were obtained in a large group of pregnant women; the chlorpyrifos body burden that did not exhibit any seasonal variations was thought to come primarily from indoor exposures (Berkowitz et al. 2003). Metals can be also found in the households; for example, lead is generated from deteriorating lead paint

and is a major concern in regards to children exposed to lead-contaminated house dust (ATSDR 1999). Further, radon can be found in many homes all over the United States (ATSDR 1998b).

1.5.1.2 Food

Another major exposure route for humans is via contaminated food. For example, North America's Great Lakes, which are the largest body of freshwater in the world, are polluted with about 362 contaminants that were found in quantifiable amounts in the water, sediment, and biota (IJC 1983; USEPA 1994). The critical pollutants were identified as PCBs, DDT, dieldrin, toxaphene, mirex, methyl mercury, benzo(*a*) pyrene, hexachlorobenzene, polychlorinated dibenzodioxins (PCDDs), polychlorinated dibenzofurans (PCDFs), and alkylated lead. Some of these pollutants biomagnify in the aquatic food chain and can be detected in increased levels in cooked Great Lakes fish. Consequently, the blood serum levels of these chemicals are significantly increased in consumers of contaminated Great Lakes sport fish compared to people who do not eat such fish (Humphrey 1983; Fiore et al. 1989; Sonzogni et al. 1991).

Another example is human exposure to mixtures of PCDDs, PCDFs, and PCBs. The primary route of exposure for the general population is the food supply (ATSDR 1998a). When the USEPA and US Department of Agriculture (USDA) completed the first statistically designed survey of the occurrence and concentrations of PCDDs and Fs in beef fat (Ferrario et al. 1996; Winters et al. 1996), pork fat (Lorber et al. 1997), poultry fat (Ferrario et al. 1997) and US milk supply (Lorber et al. 1998), the total TEQ values¹ were the highest for pork and the lowest for chicken and milk. This exposure results in a background body burden of about 5 ng TEQ/kg body weight for a mixture of dioxin-like PCDDs, PCDFs, and PCBs (USEPA 2000a). Analysis of fish oil dietary supplements showed exceedance of the WHO-TEQ limit of 2 ng/kg for dioxins in approximately 30% of the samples. When combined with whole diet intake, exposure to dioxin-like compounds was estimated at 1.8 to 8.9 pg WHO-TEQ per kilogram body weight per day for adults and 1.4 to 14 pg WHO-TEQ per kilogram body weight per day for children (Fernandes et al. 2006).

1.5.1.3 Human Exposure in Different Life Stages

Most environmental exposures are similar for a given population at a given location and time period; however, human exposures to chemicals also have some distinctive characteristics related to the life stages (Figure 1.2).

¹ Toxicity of dioxin-like chemicals is expressed in toxicity equivalents (TEQs). TEQ is defined as the product of the concentration, C_i , of an individual "dioxin-like compound" in a complex environmental mixture and the corresponding TCDD (2,3,7,8-tetrachlorodibenzo-*p*-dioxin) toxicity equivalency factor (TEF_{*i*}) for that compound. TEFs are based on congener-specific data. The TEF scheme compares the relative toxicity of individual dioxin-like compounds to that of TCDD, which had been traditionally assigned a toxicity of one (ATSDR 1998a). In 1998, the World Health Organization (WHO) released an updated system where a TEF of one was assigned not only to TCDD but also to 1,2,3,7,8-pentaCDD (Van den Berg et al. 1998).

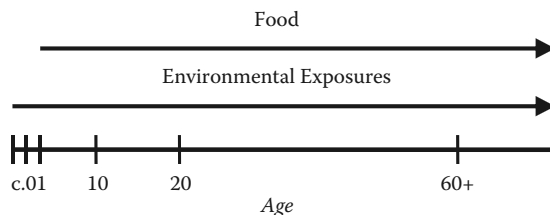


Figure 1.2 Exposures in life stages. c = conception.

1.5.1.3.1 *Fetuses*

In utero exposures represent the first contact of the developing organism with environmental pollutants. Less than half of all human conceptions result in the birth of a completely normal, healthy infant. For example, approximately 60% of spontaneous abortions are thought to be related to genetic, infectious, hormonal, and immunological factors (Bulletti et al. 1996). However, the role of the environment in the etiology of spontaneous abortion remains poorly understood. The placenta plays a key role in influencing fetal exposure by helping to regulate blood flow, by offering a transport barrier, and by metabolizing chemicals (Shiverick et al. 2003). However, the placenta never acts as a complete barrier; virtually any substance present in the maternal plasma is transported to some extent by the placenta. There is a distinction between direct and indirect developmental toxicants. Direct chemicals such as thalidomide and retinoids induce developmental toxicity without maternal toxicity. Indirect chemicals such as ethanol and cocaine mostly affect the fetus at levels also toxic to the mother. Epidemiological studies indicated that environmental exposures to low concentrations of some chemicals may cause subtle neurodevelopmental changes (Jacobson and Jacobson 1996) or may disrupt the endocrine system (Kavlock et al. 1996).

1.5.1.3.2 *Infants*

Breast-feeding is recognized as providing the developing infant the benefits of balanced nutrition and passive immunization against infections; however, exposure of infants to anthropogenic chemicals via breast milk is of concern. Many chemicals persist in the environment for a long time; they bioaccumulate in the organism and biomagnify through the food chain. Among the environmental pollutants found in breast milk are PCDDs, PCDFs, PCBs, metals, and pesticides. Considering the relatively short period of breast-feeding and relatively high daily intake, the exposure may be substantial. For example, Schecter and Gasiewicz (1987a, 1987b) estimated the daily intake of PCDDs and PCDFs in TEQs by nursing infants (10 kg) in the United States to be 83 pg TEQs/kg body weight/day. Later, Schecter et al. (1994) estimated lower intakes of 35 to 53 pg TEQs/kg body weight/day for infants (7.3 kg) (see exposure changes under Section 1.5.1.2). In contrast, intake for infants who were fed soy formula was substantially lower, ranging from 0.07 to 0.16 pg TEQ/kg body weight/day. In comparison, Schecter et al. (1994) estimated the daily intake of PCDDs and PCDFs for adults (70 kg) as 0.26 to 2.75 pg TEQs/kg body weight/day. Koppe (1995) reported that daily dietary intake of PCDDs during lactation represents only 14% of the daily secretion of PCDDs in breast milk; the rest (about

Table 1.6 Levels of various chemicals in human breast milk samples from general populations

Chemical	Range of mean or median concentrations (ng/g lipid)	Newborn intake via breast milk ^a (µg/kg/day)	Region	Reference
PCDDs and PCDFs	0.013–0.028 ^b	0.00009–0.00057 ^c	United States, Canada, Germany, New Zealand, Japan, Russia	Pohl and Hibbs (1996)
Mercury (total)	130–793 ^c	0.922–5.625	Japan, Germany, Sweden	Abadin et al. (1997)
Hexachlorobenzene	5–63	0.035–0.447	New Zealand, Brazil, Arkansas, Australia, Canada, Mexico, Quebec Caucasians	Pohl and Tylanda (2000)
Hexachlorobenzene	100 to >1000	0.709–7.094	France, Spain, Quebec Inuits, Slovak Republic, Czech Republic	Pohl and Tylanda (2000)
<i>p,p'</i> -DDE	300 to >3000	2.128–21.281	New Zealand, Brazil, France, Australia, Quebec Caucasians and Inuits, Arkansas, Canada, Slovak Republic, Czech Republic, Germany, North Carolina	Pohl and Tylanda (2000), Rogan et al. (1986)
PCBs	167–1770	1.185–12.556	Japan, Quebec Caucasians and Inuits, New York, Michigan, the Netherlands, Poland, Finland, Croatia, North Carolina	DeKoning and Karmaus (2000)

Source: Adapted from Pohl HR, McClure P, De Rosa CT. 2004. Environ Toxicol Pharmacol 18: 259–266.

^a Converted from 0.6 to 3.6 µg Hg/dL, using a conversion factor of 45.4 g lipid/10 dL milk (DeKoning and Karmaus 2000). Organic forms accounted for about 7 to 50% of total mercury (Abadin et al. 1997).

^b Measured in 2,3,7,8-TCDD toxicity equivalents (TEQs).

^c Calculated, based on assumptions of 3.2 kg body weight, 45.4 g fat/L milk, and 0.5 L milk/day (DeKoning and Karmaus 2000), as follows: 5 ng/g fat × 45.4 g fat/L × 0.5 L/day × 1/3.2 kg × 1 µg/1000 ng = 0.035 µg/kg/day.

86%) is derived from PCDDs stored in adipose tissue. Levels of selected chemicals found in breast milk around the world are presented in Table 1.6.

1.5.1.3.3 *Children*

There are many differences between children and adults. The first obvious difference is in size; children consume more food and water per kilogram of body weight, they have higher inhalation rates, and they have larger surface-area-to-volume ratios than adults. For example, Schecter and Li (1997) conducted a congener-specific analysis of PCDDs, PCDFs, and dioxin-like PCBs in US fast foods. They reported TEQ values from 0.03 to 0.28 pg/g wet weight for McDonald's big mac, 0.03 to 0.29 pg/g for Pizza Hut's personal pan supreme pizza with all toppings, 0.01 to 0.49 pg/g for Kentucky Fried Chicken's 3-piece original recipe meal, and 0.3 to 0.31 pg/g for Häagen-Dazs' chocolate–chocolate chip ice cream. Daily TEQ consumption per kilogram body weight assuming a 65 kg adult, from 1 serving of each of the fast foods tested, ranged between 0.046 and 1.556 pg/kg. The daily intake from 1 serving of each of the fast foods tested, assuming a 20 kg child (6-year-old), ranged from 0.15 to 5.05 pg TEQs/kg. A child on average consumes 3 times more TEQs on a per kilogram body weight basis than adults eating any one of the fast foods tested.

Children may also be more sensitive to harmful environmental chemicals because of differences in absorption, excretion, and metabolism (see also Chapter 2). Immaturity of some systems (e.g., immune, nervous systems) also contributes to children's vulnerability to chemicals.

Another difference between children and adults is in their behavior. Children spend more time outside and play in the dirt. Associated with this activity is soil ingestion, where hand-to-mouth ingestion has been recognized as a major exposure route (Clark et al. 1996; Hemond and Solo-Gabriele 2004). The soil ingestion value for children is based on a number of studies estimating the average soil ingestion in populations of normal children (Binder et al. 1986; Clausing et al. 1987). One of the reports suggested that an average child ingests only about 25 to 40 mg of soil daily (Gough 1991). However, about 1 to 2% of children are geophagic ("pica children") and ingest from 5 to 10 g of soil daily (USEPA 1989a). Known child-specific exposure factors have been reviewed by the USEPA (2002a).

1.5.1.3.4 *Adults*

Occupational exposures play an important role during adulthood. Often the exposures last for many years and exposure levels are much higher than those of the general population. Some of the common complex mixtures regulated in the workplace include coal tar pitch volatiles, mineral oil mist, petroleum distillates, and Stoddard solvent¹ (Hearl 2005). There are numerous possible combinations of chemicals found at the workplace; that is, most exposures are to chemical mixtures.

¹ Stoddard solvent is a colorless, flammable liquid that smells and tastes like kerosene. It will turn into a vapor at temperatures of 150 to 200 °C. Stoddard solvent is a petroleum mixture that is also known as dry cleaning safety solvent, petroleum solvent, and varnoline; its registered trade names are Texsolve S® and Varsol 1®. It is a chemical mixture that is similar to white spirits. Stoddard solvent is used as paint thinner; in some types of photocopier toners, printing inks, and adhesives; as a dry cleaning solvent; and as a general cleaner and degreaser.

Additive and synergistic effects of chemicals with common targets of toxicity are of great concern in occupational settings. For example, coexposure to acetone, sec-butyl acetate, and methyl ethyl ketone causes increased skin irritation; coexposure to heptane, methyl chloroform, and perchloroethylene increases central nervous system effects (Hearl 2005). In addition, personal behavior can affect the outcome of occupational exposures. For example, smoking increases the development of cancer in occupational exposures to asbestos (Selikoff et al. 1980) and to radon (Lundin et al. 1969; Archer 1985); alcohol increases liver effects of occupational exposure to hepatotoxicants such as carbon tetrachloride (Manno et al. 1996). Inadequate dietary protein enhances the toxicity of pesticides such as heptachlor (Boyd 1969; Shakman 1974). Household members, including children, can be exposed to workplace chemicals by coming into contact with contaminated work cloths (e.g., off-gassing of tetrachloroethylene).

1.5.1.3.5 *Seniors*

Elderly people are obviously not the only age category exposed to pharmaceuticals, but they are by far the most exposed. It is beyond the scope of this chapter to describe all the exposure scenarios and possible interactions. Interested readers are encouraged to consult other detailed literature on the topic (Calabrese 1991).

1.5.1.4 **Modeling and Measuring Human Exposure**

Several exposure models are available for human health risk assessment, some of which are summarized in a review by Fryer et al. (2006). They categorize exposure models according to the route of exposure:

- 1) environmental, distinguishing environmental concentration models (see Section 1.4) and human intake models;
- 2) dietary;
- 3) consumer product;
- 4) occupational;
- 5) aggregate, including multiple exposure pathways; and
- 6) cumulative, including multiple chemical exposure.

Fryer et al. (2006) conclude that the use of human exposure models still is fragmentary, with different organizations using different models for very similar exposure assessment situations. The main problem in the use of models is the lack of input data and the lack of validation of both the input data and the output of the model. They therefore recommend the development of an overall framework for human exposure (and risk) assessment.

One problem encountered when assessing exposure of human populations to contaminated land is spatial heterogeneity of pollution. To overcome this problem, Gay and Korre (2006) propose the combinations of spatial statistical methods for mapping soil concentrations, and probabilistic human health risk assessment methods. They applied geostatistical methods to map As concentrations in soil. Subsequently, an age-stratified human population was mapped across the contaminated area, and the intake of As by individuals was calculated using a modified version of the Contaminated Land Exposure Assessment (CLEA) model. This approach allowed a

determination of sites with clearly elevated human exposure, and may also be used to determine exposure to mixtures of chemicals.

Weis et al. (2005) reported the results of the ad hoc Committee on Environmental Exposure Technology Development. They identified a toolbox of methods for measuring external (environmental) and internal (biological) exposure and assessing human behaviors that influence the likelihood of exposure. The toolbox of environmental exposure methods includes environmental sensors, such as in vitro sensors, like personal dosimeters, to detect and quantify priority environmental exposure, and Geographical Information System (GIS) technology to map and link environmental and personal exposure. The latter technique may also be used to identify populations at risk. The internal exposure methods comprise biological sensors, toxicogenomic measurements, and body burden analyses. The latter also include determination of biomarkers of exposure, such as DNA adducts. All these methods may be used to determine exposure to single chemicals and mixtures.

1.5.1.5 Human Biobanks and Human Volunteer Monitoring of Exposure

Monitoring body burdens of chemicals in human populations may contribute to a better understanding of what chemicals and at what concentrations they get into the body. For chemicals with known toxicity levels, it may be possible to learn the prevalence of people with concentrations exceeding those toxicity levels. However, it should be noted that most biomonitoring studies in the general population are designed as survey studies. Many of the studies do not take into account exposure history. Therefore, the results often represent a snapshot at 1 point in time.

In the United States, the Centers for Disease Control and Prevention (CDC) released the third National Report on Human Exposure to Environmental Chemicals (CDC 2005). The report presents blood and urine levels for 148 environmental chemicals (or their metabolites) found in the general civilian US population over the 2-year period 2001–2002. The study is part of the National Health and Nutrition Examination Survey (NHANES), designed to provide an insight into the health and nutritional status of the US population. Future follow-ups are planned to encompass 2-year periods with information on trends of exposure in population groups defined by age, sex, and race. The latest report monitored 32 more chemicals than the second report, which encompassed the 2-year period from 1999 to 2000 (CDC 2003). The major chemical groups monitored included metals, phytoestrogens, PAHs, PCDDs, PCDFs, coplanar and mono-ortho-substituted biphenyls, non-dioxin-like PCBs, carbamate pesticides, organochlorine pesticides, pyrethroid pesticides (added in the third report), phthalates, organophosphate insecticides, and herbicides. Cotinine, a major metabolite of nicotine, was measured as an indicator of smoking. Urine creatinine was analyzed as a continuous variable for chemicals measured in urine to adjust for urinary dilution. Sample sizes varied under each of the categories from several hundreds to several thousands.

The large-scale monitoring studies enable researchers to track, over established time periods, trends in levels of exposure in human populations and to assess the effectiveness of public health efforts to reduce exposure to specific harmful chemicals. Aylward and Hays (2002) summarized recent trends in dioxins intake in the United States and in Western Europe. The intake estimates show clear decreases of

dioxin exposures in these countries. For example, USEPA's 2000 estimate of 0.6 pg TEQ/kg/day is 66% lower than the 1994 estimate of 1.7 pg/kg/day for PCDDs/Fs. In the United Kingdom, the intake levels for PCDDs/Fs were 4.6, 1.6, and 0.9 pg TEQ/kg/day in 1982, 1992, and 1997, respectively. Similarly, PCDDs/Fs intakes were estimated as 4.2, 1.8, and 0.5 pg TEQ/kg/day in 1978, 1984, and 1994, respectively, in the Netherlands. Similar trends were reported for PCB intakes (Aylward et al. 2002). These decreases in intake are reflected in decreases in human body burdens. A large number of studies in the general population in the United States, Canada, Germany, and France during 1972–1999 show a trend of substantial (almost 10-fold) decreases in human TCDD-only body burden over that time period (Aylward and Hays 2002). Considering the long half-life of TCDD, a 1-compartment pharmacokinetic model estimated that the decrease in intake must have been more than 95%. A recent retrospective time-trend study that analyzed levels of major halogenated aromatic hydrocarbons in human serum concluded that PCB and polybrominated biphenyl (PBB) levels are also decreasing since their phaseout in the 1970s (Sjodin et al. 2004). In contrast, concentrations of polybrominated diphenyl ethers (PBDEs) have been increasing in recent years in the United States, because of their use as flame retardants. Such a very substantial increase has not been observed in Europe, where PBDE levels in human serum are about 10 times lower than in the United States (Thomas et al. 2006).

Large-scale monitoring studies are also under way in Europe (Pohl et al. 2005). In 2003, the EU commission launched an initiative called SCALE, which stands for Science, Children, Raising Awareness, Legal Instruments, and Evaluation. The objectives are to reduce disease burden caused by environmental factors, to identify and prevent new environmental threats, and to strengthen the EU policy-making capacity. The EU commission asked for the cooperation of all stakeholders in identifying and addressing the most relevant children's environmental health issues. Biomonitoring was specifically addressed in a document that called for unified testing approaches all over Europe so that the study results are comparable (Pohl et al. 2005). From 2003 to 2005, the Robert Koch Institute monitored 18,000 children aged from <1 to 18 years at 150 different places in Germany over a 3-year period. Detailed interviews on pregnant women, parents, and children—together with medical examinations—focused on health risks in modern life, relevant environmental pollution, psychological health, and motoric development in childhood. Similarly, the Erasmus Medical Centre in Rotterdam monitors about 10,000 subjects over a 20-year period: from early fetal life until young adulthood. The project started in 2002. Physical examinations, questionnaires, interviews, and ultrasound examinations are performed; biological samples are collected as well. The focus is on pediatric growth, neurobehavioral development, pediatric diseases, and preventive health care for mother and child.

1.5.2 EXPOSURE IN ECOSYSTEMS

Estimating exposure through measurement of chemical concentrations in tissues or body fluids (e.g., blood, urine) of environmental species (or a suitable surrogate) is a long-standing concept going back many years. In recent times, interest in the

“biomonitoring” approach has grown to the point where the subject was the focus of a *Nature* editorial that looked explicitly at application in developed and developing regions (Whitfield 2001). Some current monitoring programs have been measuring residue levels of multiple chemicals in biological samples over extended periods, while others are targeted studies addressing particular sites or issues. The principal driver behind each of the studies is not usually to quantify the exact nature of multiple chemicals exposure, but instead to assess spatial and temporal changes in exposure level for particular contaminant groups of concern (e.g., metals, PCBs, organochlorine insecticides). Despite this focus, many of these monitoring schemes can provide valuable information on the nature of complex exposure scenarios, and a few even explicitly report on the potential mixture effects that may result from a multiple chemical exposure.

The nature of the different environmental compartments means that the species selected for exposure monitoring may vary between different environmental types (e.g., air, soils, sediment, freshwaters, and marine waters). Similarly, the characteristics of the chemicals being investigated (often persistent pollutants) can also influence species choice. For some environmental compartments, such as soils and sediments, it is possible to select species for monitoring that are exposed through different routes, such as via pore water or by ingestion of contaminated particulate matter. Because of the potential for pulsed exposure in mobile media such as air and running waters, sampling for exposure biomonitoring in these ecosystems may need to be repeated to avoid missing potentially significant exposures to mobile or ephemeral pollutants. In soil and sediment, which act as contaminant sinks, such temporal repetition may be less of an issue, although this depends on the persistence of the compounds being assessed. Further in these environments, spatial heterogeneity can be more of an issue. Approaches for different environmental compartments and details of the finding and outcomes of particular schemes associated with each are outlined in detail below.

1.5.2.1 Air

Exposure through air is particularly important for gases, volatile compounds and chemicals associated with the surfaces of small airborne particulates. Since plants are in intimate contact with the air through leaf cuticles and stomatal pores, this group has been most widely used for airborne contaminant monitoring. The first widespread application of plants for biomonitoring was to support policy implementation on issues relating to acidification and eutrophication (Cape et al. 1990; Bobbink et al. 1992). Building on this work on sulfur and nutrients, metals have also been the subject of numerous studies measuring both spatial and temporal aspects of accumulation in plant leaves. Burton et al. (1986) reported that many surveys involving metal analysis of lichen thalli reflected the spatial variability of metal deposition. Bargagli et al. (2002) compared metal concentrations in 2 common biomonitor species *Hypnum cupressiforme* (a moss) and *Parmelia caperata* (lichen) around an intensive mining area. While both moss and lichen were able to indicate the nature of the complex atmospheric emission occurring in the region, each accumulated different concentrations of different metals. This emphasizes the difficulty in reading across exposure scenarios between species.

This problem is, however, not unique to plants, but is in fact applicable to many taxa (Hopkin et al. 1993; Morgan and Morgan 1993; Newton et al. 1993). One area where the analysis of mosses and lichens has made a significant contribution to understanding of present and past pollution trends has been through the analysis of samples stored in herbaria. These samples provide useful information on temporal trends in deposition that can be used in the development of dynamic models to describe pollutant loading and fate in terrestrial environments (Hassanin et al. 2005). This issue may be particularly relevant in describing past exposure that may lead to current environmental and human health concerns (e.g., past carcinogen exposure).

An obvious issue regarding the use of plants for airborne pollutant biomonitoring is that they are also potentially exposed through the soil. This confounding effect is one reason why many monitoring studies of wet and dry deposition have focused on mosses and lichens, which lack roots and so rely on atmospheric deposition to surface cuticles to obtain adequate nutrients. For PAHs, and probably also for other lipophilic organic compounds, uptake from soil may also be limited by the strong adsorption to soil and lipid materials associated with root cells (Watts et al. 2006). For such compounds, therefore, air accounts for most of the burden on or in plant leaves and other aboveground tissues. This makes mosses and lichens excellent potential monitors for organic pollutants, with the physiology of the leaf, such as the presence and nature of leaf hairs and the form of any extracuticular wax present, being an important influence on the rate of uptake (Bakker et al. 1999; Jouraeva et al. 2002).

1.5.2.2 Water

The potential for sustained and pulsed exposure in riparian, lake, estuarine, and coastal systems has established biological monitoring as a potentially useful tool for characterizing the chemical status of these habitats. For monitoring contaminants in the water column, filter feeders and species with an extensive gill system are frequently favored. This is because the extensive contact between biological membranes and the water column in these species ensures that there is significant exposure. An example of the kind of coordinated approach that can be used for monitoring waterborne pollutants using a filter feeder species is the mussel watch program run by the National Center for Coastal Ocean Science in the United States. This scheme measures concentrations of over 100 contaminants in the tissues of marine mussels from almost 300 US sites covering the Atlantic, Pacific, and Gulf coasts, and also the American shores of 3 of the Great Lakes (Michigan, Huron, and Erie). Chemical groups measured include 18 elements (17 metals plus Si), over 50 PAHs, 31 PCBs, 31 organochlorinated compounds, organometals (such as butyl-tin compounds), PCDDs, and PCDFs. Major conclusions from the survey to date include evidence of a widespread, but declining exposure to multiple organochlorines; a widespread exposure to multiple PCBs that is reducing over time (but not as quickly as for the organochlorines), a widespread exposure to PAHs that has remained largely unchanged, except where there has been a specific accidental exposure (Page et al. 2005), a decreasing exposure to organo-tin compounds, and a very prominent exposure to metals that has reduced for some (e.g., lead) but remained unchanged for others (e.g., copper and zinc) (National Oceanic and Atmospheric Administration 2002).

The large-scale nature of the US mussel watch program has, of course, led to the development of many smaller-scale schemes in other countries and regions that have mirrored the use of mussel species as subjects for biological monitoring (Sole et al. 2000; Kim et al. 2002; Rainbow et al. 2004; Mendoza et al. 2006). Other schemes also used other bivalve species, such as oyster and scallop, as alternatives to mussels (Daskalakis 1996; Silva et al. 2003; Norum et al. 2005). These biomonitoring studies range from small-scale surveys of country or regional coastlines conducted for particular contaminant groups (e.g., metals, PAHs, PCBs) to major regional surveys that rival the US program (e.g., Tanabe 2000). Being also subject to the vagaries of the research funding system, it is often difficult to obtain sufficient funds to repeat surveys, meaning that in some cases, the temporal element that is so important in the mussel watch scheme is lost. The large-scale surveys set up for coastal waters in Asia are, however, beginning to generate data suitable for the derivation of temporal trends (Sudaryanto et al. 2002; Monirith et al. 2003).

A second major group of invertebrate species recommended for marine biomonitoring is crustaceans. The barnacle *Balanus improvisus* has been used as a biomonitor of the metals Cu, Zn, Cd, Fe, Pb, Mn, and Ni in the Gulf of Gdansk (Baumard et al. 1998). Insects have also been used in freshwater (Fialkowski et al. 2003). As well as invertebrates, fish have also been recommended as potential monitors of exposure to multiple contaminants. Despite particular issues with the use of fish for monitoring organic pollutants (see below), biomonitoring studies have been able to separate individuals from sampling regions (coastal sea vs. oceanic) with different levels of prevalent pollution (Stefanelli et al. 2004). Since fish are both abundant and important components of aquatic ecosystems (at intermediate to high levels within aquatic food webs), and also because fish are an important food source, monitoring of contaminant levels in fish can provide an important link between environmental exposure to multiple compounds and human exposure to the same compounds through diet (Meili et al. 2003).

While biomonitoring studies undoubtedly provide useful information of mixture exposure assessment, the presence or, more importantly, absence of a compound in a particular species does not provide the full picture. Physiology may have a major influence on the tendency of an organism to accumulate certain chemicals. For example, fish have a potential to metabolize organic molecules that is higher than in many of the invertebrate taxa that are the most common subject of invertebrate monitoring. This means that more recalcitrant persistent organic pollutants, such as PCBs and organochlorine insecticides, are accumulated at low or very low concentrations in fish, and easily biodegradable compounds, such as PAHs and chlorinated phenols, do not tend to accumulate (van der Oost et al. 2003). A further issue in water is the pulsed nature of potential exposure, particularly in rivers. In cases where conditions, such as surface flow, cause pollutant runoff (e.g., pesticides from an arable field), this exposure can cause detrimental effects to receiving stream ecosystems. Later chemical biomonitoring studies may, however, fail to identify this exposure if the compound is both degraded from stream sediment and rapidly metabolized by the chosen monitoring species. This problem can be seen as part of a wider issue with the use of biomonitoring to detect complex exposure to easily metabolized compounds, which is described later.

1.5.2.3 Sediment

As sediments act as pollutant sinks in aquatic systems, they can be important sources of exposure, and so of the entry of chemicals into aquatic food chains. Sediments are the ultimate residence location for many pollutants released to water. The widespread presence of complex mixtures of contaminants in sediment is thus likely to occur in any location where multiple localized and diffuse contaminant sources contribute to the overall chemical load within natural waters. The role of sediment in the receipt and resupply of the chemical to the water phase means that there is interest in monitoring sediment chemical pollutant load over both different spatial and temporal scales. Because the process of sediment deposition and chemical adsorption on the one hand and solubilization and resuspension on the other link the pollutant loads of the sediment and water column, many of the species that can be used to sample the environment for waterborne pollutants (e.g., filter feeders such as mussels) can also describe the pollutant load present in sediments (Baumard et al. 1998).

An alternative to the use of filter feeders as proxies for sediment exposure is to utilize fully sediment-dwelling species. Two groups have been suggested. The first are the annelids. In freshwaters, the most commonly studied is the sludge worm, *Tubifex tubifex* (Egeler et al. 1999), while in marine sediments 2 polychaetes, *Nereis diversicolor* and *Arenicola marina*, have been used as biomonitors of single and multiple chemical exposures with varying degrees of success (Kaag et al. 1998; Poirier et al. 2006). The second sediment-dwelling group used for biomonitoring is insects, and in particular chironomids (Bervoets et al. 2004; Martín-Díaz et al. 2005a). Overall, the use of sediment-dwelling species is less well established than for bivalves such as freshwater and marine mussel species and, as a result, this later group remains the preferred taxa for multiple chemical exposure assessment in aquatic systems.

1.5.2.4 Soil

As soil is an important sink for pollutants in terrestrial ecosystems, there is interest in assessing the exposure of humans and ecological species through measurement of concentrations in organisms exposed through this medium. Such measurements can reflect the direct application of chemicals to soils as pesticides or solid waste or by aerial deposition of contaminants to the soil surface (Van Brummelen et al. 1996b; Filzek et al. 2004). Unlike air and surface waters, soil is immobile and hard to dilute, so there is a potential for temporally more stable long-term exposure than in mobile media, although it is also more likely that exposure is more heterogeneous, even at relatively small spatial scales. Since true soil organisms, such as earthworms, nematodes, and springtails, have low mobility, this means that tissue residue concentrations in these species can provide a more reliable estimate of the local exposure than more active surface-dwelling species, such as beetles, centipedes, spiders, and small mammals, for which tissue residue concentrations provide a picture of the average of contaminant concentrations over the full home range. Thus, depending on the question being asked by selecting different soil species for monitoring, it is possible for direct analysis to both include and partially ignore local scale spatial heterogeneity. For example, by using earthworms as the subject species, Marinussen and Van der Zee (1996) were able to model the spatial patterns of exposure in heterogeneously

contaminated soils at an industrial facility. Such an analysis would, however, have been even more complex for very mobile species or animals with a complex life history for a single compound, let alone if multiple chemicals and their combined effects were to be considered.

Despite issues relating to chemical heterogeneity and the subsequent choice of monitoring species, the success of the mussel watch program for marine waters has prompted the initiation of schemes in soils that mirrored the marine approach. As in marine work, it is important to consider the physiology of the species when designing and interpreting the outputs of any monitoring scheme. For example, in laboratory experiments, slugs have been shown to be sensitive to metal pollution and were thus suggested to be useful for biological assessment of soil exposure (Marigomez et al. 1998). Work to assess the potential of slugs as biomonitors was, therefore, conducted at a copper-contaminated site, concluding that exposure and effect biomarkers recorded in sentinel slugs could be sensitive, quick, and cheap indices of metal pollution in soils (Marigomez et al. 1998). Similarly using woodlice, Hopkin et al. (1986) were able to conduct reliable exposure maps for the presence of multiple metals from diffuse and localized industrial sources over a medium-sized town (Reading) of 250,000 people in the United Kingdom. Jones and Hopkin (1991) also looked at the biomonitoring potential of woodlice and mollusks, concluding that both groups had potential and that each sampled exposure in a similar manner. This was indicated by the high correlation between tissue concentrations for each metal found between the different measured species. The studies outlined above have demonstrated the feasibility of using soil species to provide a picture to environmental exposure. A review of the published literature, however, shows that while this line of research was certainly the fashion 10 years ago, it is losing favor, with few recent papers focusing on this kind of spatially based tissue concentration assessments.

1.5.2.5 Monitoring of Food Chain Transfer

The potential for some chemicals to transfer through food chains has resulted in the development of a set of long-term, large-scale monitoring studies that measure exposure of top predators (predatory birds and mammals) through quantification of chemical concentrations in tissues (e.g., liver) and eggs. Monitoring of top predators arose in response, first, to recognition of the effects of organochlorine insecticides on bird populations (Newton and Wyllie 1992; Walker et al. 2001) and, second, to the realization that these pollutants have the potential to circulate across regions and ultimately around the globe (Wania and Mackay 1996; Gouin et al. 2004). The focus on top predators, which can range across large territories, means that the residue levels measured represent a cumulative exposure across the landscape, rather than the specific regions in which they were sampled. For some bird species in particular, exposure can also occur in different regions as a result of migration, and this can result in different patterns of tissue concentration depending on specific regional chemical usage patterns (Minh et al. 2002).

Although there are numerous fairly small-scale academic studies of the concentration of persistent pollutants in avian or mammalian predators (Kenntner et al. 2003a, 2003b; Berger et al. 2004; Jaspers et al. 2005; Hela et al. 2006), the 2 most

important long-term large-scale schemes for quantifying wildlife exposure through the food chain are the UK Predatory Bird Monitoring Scheme (UK-PBMS) and the Arctic Monitoring Program of the USEPA (US-AMP). To briefly summarize both schemes, the UK-PBMS has run since the mid-1960s and measured chemical concentrations in the tissues of birds collected as dead carcasses by volunteers and non-hatching (sterile) eggs collected under license from nest sites. Like many monitoring schemes, the focus has been on quantifying exposure, including spatial (Alcock et al. 2002; Broughton et al. 2003) and temporal (Newton et al. 1991, 1993) trends. Current analytes measured include organochlorines and their metabolites, PCBs, second-generation rodenticides, mercury, and in a more limited set of samples, polybrominated compounds and PAHs.

Analyses in the UK-PBMS concentrate on the use of single methods to measuring concentrations of particular subgroups of chemicals rather than on the application of diverse methods to detect the full range of residues that may be present. Reporting also focuses on trends for single chemicals in isolation. The exception to this is for PCBs, where the scheme expressly considered the combined chemical dose that is present in the tissues of the birds using the TEQ method of Ahlborg et al. (1994) and Van den Berg et al. (1998). Sum totals of concentrations may also have been reported for other chemical subgroups, such as the organochlorines and PAHs. While relatively coarse, these summed values provide a relatively simple means of defining the changes in concentrations of the measured entirety of these contaminant groups in tissue.

The US-AMP is probably the largest-scale pollutants monitoring initiative currently undertaken. The scheme focuses on measuring chemical concentrations in soils, water, and biota at higher latitudes. The focus on polar latitudes can be linked to concerns regarding the potential for movement of persistent organic pollutants (POPs) over time to higher regions of the globe (Wania and Mackay 1996). This process occurs because of the fact that warmer temperature at lower latitudes favors volatilization of POPs, while the colder temperature at higher latitudes favors deposition. Such cycles of volatilization at lower latitudes and deposition at higher latitudes (often termed global distillation) may, therefore, lead to the accumulation of higher concentrations and an increasing contribution from the more volatile compounds in polar regions.

Biota samples measured in US-AMP include arctic plants such as mosses, large terrestrial herbivores such as caribou, waterfowl, and top predators including birds of prey and mustelids (mink and marten). Aquatic invertebrates and some fish species are also measured. Like UK-PBMS, the focus in US-AMP is on the spatial and temporal trends for individual contaminant groups, rather than holistic assessments of multiple chemical exposures. Overall, schemes such as the US-AMP and UK-PBMS and similar small-scale national programs (Sørensen et al. 2004) can provide an excellent summary of mixture exposure scenarios for top predators exposed through the food chain. The databases generated by these programs could become essential data resources for further data mining for historic and current mixture exposure assessment for wildlife and possibly even humans.

While monitoring of chemical residue levels can provide a useful snapshot of the range and extent of current exposure to some compound groups, the approach does

have some limitations. Birds and mammals, like fish, have a high metabolic potential for some organic contaminants. This means that some chemicals that are subject to rapid degradation and metabolism are not easy to reliably detect. This favors measurement of more recalcitrant contaminants, such as PCBs and organochlorinated insecticides, and also some metals such as mercury and cadmium, for which modeling (Romijn et al. 1993a, 1993b, Spurgeon and Hopkin 1996) and measurements (Hunter et al. 1987, 1989; Read and Martin 1993; Kooistra et al. 2005) have indicated the potential for concentrations to reach potentially harmful levels in tissues (Nicholson et al. 1983). For organic compounds that rapidly decay or for metals that are subject to strong homeostatic regulation, analyses for top predators may provide only a limited summary of current exposure, even though such compounds may make a substantive contribution to toxic effects.

1.5.2.6 Multimedia Exposure Scenarios

The exposure scenarios detailed above focus on separate assessment using biomonitoring organisms applicable to the particular environment under consideration. This separation of exposure by environmental compartment does not reflect the true nature of exposure for many species. For example, higher plants are likely to be exposed to airborne contaminants through the leaf surface and through the soil solution; sediment-dwelling filter feeders can be exposed through the interstitial water and also through the main water column; and top predators can be exposed through air, water, and their food supply. Elucidating the principal exposure routes for different species is an extremely active area of research in both aquatic and terrestrial environments (Vink et al. 1995; Irving et al. 2003; Jager et al. 2003), and there are obvious implications for the assessment of exposure in different exposure routes, and as a result, careful consideration needs to be given to the assessment. Such assessments are likely to be specific for each taxon and need to consider the major life history and behavioral characteristics of the organism.

Even more complex than the situations of multiple exposure routes are situations where species move between different environments during different stages of the life cycles. At their simplest, such scenarios can simply be due to feeding in different places at varying times of the year. More complex scenarios occur in species that spend separate parts of their life cycle in different environments. A nice example is given by Linkov et al. (2002). In their model, PCB exposure and bioaccumulation in winter flounder is described, taking into account spatial and temporal exposure characteristics. Other examples include many species of insect (e.g., Ephemeroptera, Tricoptera, Chironomidae) that spend their larval stages fairly sedentary in water and their adult stages as mobile wide-ranging species in terrestrial ecosystems. For such species, it is difficult to establish the true complex nature of exposure, and so measurement may be the only appropriate method to evaluate the exact scenario. A promising attempt to model spatially explicit ecological exposure of terrestrial organisms, taking into account spatial, temporal, and multiple stressor interactions, and addressing landscape heterogeneity, has been described by Hope (2001, 2005). Hope (2005) identified physical (loss of habitat) and biological (lack of adequate food) stressors, in addition to chemical stressors. Other examples of studies that took into account spatial exposure

patterns by using geostatistical or GIS-based methods include Clifford et al. (1995) for dieldrin and Kooistra et al. (2005) for Cd accumulation in terrestrial food chains.

1.5.2.7 Critique on Biomonitoring Studies for Complex Exposure Assessment

Biological monitoring is one of the best ways to provide a picture of current exposure to environmental mixtures. However, monitoring programs have to be carefully designed and results reviewed with caution. Even the largest and most comprehensive studies are limited in their scope to assess the true nature of complex environmental exposure simply by the fact that it is not feasible to measure the full range of potential pollutants. In most cases schemes are designed to meet particular policy objectives (i.e., characterized predator exposure to POPs) or for particular site-specific scenarios (i.e., metal levels in mosses around a metal smelter). Even when chemicals can conceivably be measured, the potential for metabolism can present a significant problem. This relates not only to compounds that are rapidly metabolized, and so difficult to detect even after a significant exposure event, but also to interindividual variability, due, for example, to enzyme polymorphism, that may introduce variability into the system that may mask time-dependent or spatial exposure trends.

As outlined in the discussion for particular sections above, an important fact to consider when reviewing biological monitoring data is the behavioral characteristics and lifestyles of the subject species. In the simplest case, monitoring of sessile species may be useful to provide a local scale view of exposure, while measurement of highly mobile species may provide a region scale exposure summary. Even when species have broadly similar lifestyles (e.g., birds of prey), behavioral and prey differences mean that biomonitoring can show different results for different species. While direct measurement in monitoring studies can account for these differences, this presents a particular problem for reading across exposure scenarios for species with different habitats, food sources, and behavior. Only in cases where the variables and differences that have the greatest influence on the nature of exposure are known is it possible to make interspecies predictions regarding a particular exposure scenario.

1.5.2.8 Effect-Directed Assessment

A final means of assessing combined exposure is through the direct application of biological testing for effects-based assessment of complexly polluted media (e.g., effluents, soils, sediments). For the use of bioassays for direct assessment of complex mixtures, the reader is referred to Chapter 4.

Matching the development of bioassays for complex exposure assessment has been the development of the use of biomarkers. This area has been the subject of a number of detailed reviews, some wholeheartedly recommending the approach and others being more critical (Decaprio 1997; Kammenga et al. 2000; Gagne and Blaise 2004; Forbes et al. 2006). Whatever the pros and cons of biomarkers, the use of effects-based analysis clearly has an appeal in assessing exposure to mixtures and its consequences. Such approaches have already been used in human exposure monitoring. For example, metabolite monitoring is regularly used for occupational human

exposure in regulatory settings, and it is easy to envisage such approaches being used for exposure assessment in environmental species.

A final form of direct effect assessment that can be used for exposure monitoring is through monitoring the community composition for microorganisms, meso- and macrofauna, and plants. This approach is based on the fact that the exposure to specific pollution mixtures may be expected to result in the elimination of sensitive taxa and species. Physiological and community diversity-based profiling methods for microorganisms are becoming increasingly routine, and for microinvertebrates, the development of community-based monitoring systems, such as the River Invertebrate Prediction and Classification System (RIVPACS) scheme in the United Kingdom (Hawkins et al. 2000; Wright et al. 2000) and analogous schemes in other countries, is becoming increasingly widely used for assessing the ecological condition in regulatory regimes such as the EU Water Framework Directive. In these cases, the challenge lies practically in providing the required level of taxonomic resolution in “difficult taxa” and interpretationally in making causal links between the observed community change and the complex nature of the combined exposure to multiple stressors.

1.6 SUMMARY AND CONCLUSIONS

Estimation of mixture exposure requires an assessment of all steps from emission of chemicals, fate in the environment, bioavailability in different environmental compartments, and interactions at the uptake level. In addition, behavioral aspects and life-stage-specific exposures have to be considered.

Emission estimation methods are available for several sources and chemicals (or groups of chemicals), but focus is generally on single chemicals rather than on mixtures. Multimedia fate models may be used to predict or estimate the fate and distribution of chemicals in the environment. Such models are available for different scales, but the precision of the prediction usually increases with increasing level of details required. Physical–chemical properties of the chemicals and characteristics of the environment determine the composition of a mixture ending up at a certain site or in a certain place. Exposure is determined by factors affecting bioavailability of the chemical, such as binding strength to soil or sediment particles. Chemical–chemical interactions may also affect availability. In addition, individual behavior of organisms, including man, may influence exposure. Methods to determine exposure mainly include residue analysis, either in exposure media like air, water, soil, or food, or in tissues of organisms being exposed. Also, these monitoring methods usually are focusing on single chemicals rather than on mixtures. These long-term and large-scale data sets can provide essential information that can be used to validate the outcome of emission scenario and environmental fate models.

1.7 RECOMMENDATIONS

Although several emission registrations exist, these mainly focus on single (groups of) chemicals rather than on assessing mixture exposure. In addition, emission estimations or registrations seem mainly to take place on an ad hoc basis, with little international coordination.

- 1) Generate emission data that may help better estimate mixture exposure.
 - a) Collection of data on existing chemicals should be evaluated to determine if they are fit for the purpose of mixture scenario prediction.
 - b) International collaboration for exchanging emission data.
 - c) There is a special need to consider emission of new and emerging chemicals, for example, nanoparticles.

One of the reasons why current emission estimation methods do not focus on mixtures might be that there are so many possible combinations of chemicals. With some guidance on the combinations of chemicals most relevant or most likely to cause problems, it would become easier to focus.

- 2) Prioritize approaches to emission estimation to focus on most common and most relevant mixture emission scenarios.
 - a) Further development of desk-based methods for identifying most probable mixture scenarios and widespread release of modeling outcomes.
 - b) Refine large-scale monitoring programs, for example, global programs, such as the Arctic Monitoring and Assessment Program (AMAP) and the European Monitoring and Evaluation Program (EMEP) to focus on inclusion of the most relevant old and new chemicals alone and in mixtures.

Although several recent studies demonstrate awareness of the fact that chemical fate processes may have a large influence on the composition of mixtures, models usually only focus on single chemicals. Nevertheless, such models may be very useful to predict composition of mixtures, but validation is needed.

- 3) Studies on fate in the environment should include aspects that cause a change in mixture composition from emission until exposure.
 - a) Models that describe distribution of mixtures in the environment need to be validated.

Several factors and processes may lead to interactions between chemicals in the environment. Such interactions not only determine fate and transport in the environment, but may also play a role in determining uptake. More insight into such interactions is highly needed in order to enable a more accurate exposure assessment of mixtures.

- 4) Research on potential interactions between individual chemicals that might affect the exposure, availability, or toxicity of the mixture and inclusion of outcomes into developing multimedia fate models for mixtures.

Monitoring programs mainly focus on measuring (single) chemicals in the environment or in organisms as an indication of exposure. In many cases no additional information is provided, hampering a proper interpretation of such data.

- 5) Monitoring programs measuring total concentrations should include measures of parameters (environmental characteristics, for example, clay, organic carbon content, pH, dissolved organic carbon (DOC)) and physical chemical properties of compounds that help to evaluate bioavailability.

Several models have been developed to link bioavailability or uptake of chemicals in organisms to their speciation in the environment. Again, focus generally is on single chemicals, while extension for use on mixtures is desired.

- 6) Research is needed to assess if it is possible to extend integrated models that link exposure-toxicity (like the BLM) for use with mixtures.

Like many data, emission and exposure data are presented as constant values, often a mean with standard deviation. In environmental risk assessment, however, awareness is growing that a stochastic or probabilistic approach is more suitable to obtain insight in the possible risk of chemicals. This also requires expressing exposure data as statistical, probabilistic distributions. Also in this case, the focus should be extended to mixtures.

- 7) Improve methods for identification of the probabilistic distribution of short- and long-term exposure of possible chemical mixtures for ecosystems and humans.

An adequate assessment of exposure to mixtures may require development of improved tools for measurement or detection of chemicals, but also for assessing temporal aspects of exposure.

- 8) Generate data that may help better estimate mixture exposure.
 - a) Analytical methods should be available for new high-production-volume chemicals coming onto the market (e.g., as for pesticides).
 - b) Differentiate between simultaneous and subsequent exposure
 - i) Long-term trends at the same locations
 - ii) Spatial sampling at the same time
 - c) Better understanding of routes of exposure, potential entry and exposure of humans to new chemicals, and their contribution to mixtures.

Assessing human exposure is quite complex, because exposure is dependent on life stage and may be influenced by behavioral patterns. Prediction of exposure may be improved by accounting for these aspects.

- 9) Improve prediction of exposure for different life stages of humans for chemical mixtures accounting for behavior patterns.

ACKNOWLEDGMENTS

Thanks are due to Fred Heimbach for his valuable contribution to the discussions at the International SETAC/NoMiracle Workshop on Mixture Toxicity in Krakow that led to this chapter. Ad Ragas is acknowledged for his critical and valuable review of this chapter.

References

- Abadin H, Hibbs B, Pohl H. 1997. Breast-feeding exposure of infants to environmental contaminants—a public health risk assessment viewpoint. II. Cadmium, lead, mercury. *Toxicol Ind Health* 13:495–517.
- Abou-Donia MB, Makkawy HM, Campbell GM. 1985. Pattern of neurotoxicity of n-hexane, methyl-n-butyl ketone, 2,5-hexanediol, and 2,5-hexanedione alone and in combination with o-4-nitrophenyl phenylphosphonothionate in hens. *J Toxicol Environ Health* 16:85–100.
- ACGIH. 1984. Threshold limit values—discussion and thirty-five year index with recommendations. Cincinnati (OH): American Conference of Governmental Industrial Hygienists.
- ACGIH. 2000. 2000 TLVs and BEIs. Threshold limit values for chemical substances and physical agents and biological exposure indices. Cincinnati (OH): American Conference of Governmental Industrial Hygienists.
- Ahlborg UG, Becking GC, Birnbaum LS, Brouwer A, Derks H, Feeley M, Golor G, Hanberg A, Larsen JC, Liem AKD, Safe SH, Schlatter C, Waern F, Younes M, Yrjanheikki E. 1994. Toxic equivalency factors for dioxin-like PCBs. *Chemosphere* 28:1049–1067.
- Albers EP, Dixon KR. 2002. A conceptual approach to multiple-model integration in whole site risk assessment. In: Rizzoli AE, Jakeman AJ, editors, *Integrated assessment and decision support. Proceedings of the First Biennial Meeting of the International Environmental Modelling and Software Society. Part 1*. Manno (CH): iEMSs. p 293–298.
- Alcock RE, Boumphrey R, Malcolm HM, Osborn D, Jones KC. 2002. Temporal and spatial trends of PCB congeners in UK Gannet eggs. *Ambio* 31:202–206.
- Alda Álvarez O, Jager T, Kooijman SALM, Kammenga JE. 2005. Responses to stress of *Caenorhabditis elegans* populations with different reproductive strategies. *Funct Ecol* 19:656–664.
- Alda Álvarez O, Jager T, Marco Redondo E, Kammenga JE. 2006b. Assessing physiological modes of action of toxic stressors with the nematode *Acrobeloides nanus*. *Environ Toxicol Chem* 25:3230–3237.
- Alda Álvarez O, Jager T, Nuñez Coloa B, Kammenga JE. 2006a. Temporal dynamics of effect concentrations. *Environ Sci Technol* 40:2478–2484.
- Aldenberg T, Jaworska JS, Traas TP. 2002. Normal species sensitivity distributions and probabilistic ecological risk assessment. In: Posthuma L, Suter GW, II, Traas TP, editors, *Species sensitivity distributions in ecotoxicology*. Boca Raton (FL): Lewis Publishers. p 49–102.
- Alexander M. 1995. How toxic are toxic chemicals in soil? *Environ Sci Technol* 29:2713–2717.
- Ali N, Tardif R. 1999. Toxicokinetic modeling of the combined exposure to toluene and n-hexane in rats and humans. *J Occup Health* 41:95–103.
- Allen BC, Kavlock RJ, Kimmel CA, Faustman EM. 1994. Dose–response assessment for developmental toxicity II. Comparison of generic benchmark dose estimates with no observed adverse effect levels. *Fund Appl Toxicol* 23:487–495.
- Allen HE, editor. 2002. *Bioavailability of metals in terrestrial ecosystems: importance of partitioning for bioavailability to invertebrates, microbes, and plants. Metals and the Environment Series*. New York: SETAC.
- Altenburger R, Nendza M, Schüürmann G. 2003. Mixture toxicity and its modeling by quantitative structure–activity relationships. *Environ Toxicol Chem* 22:1900–1915.
- Altenburger R, Schmitt H, Schüürmann G. 2005. Algal toxicity of nitrobenzenes: combined effect analysis as a pharmacological probe for similar modes of interaction. *Environ Toxicol Chem* 24:324–333.

- Altenburger R, Walter H, Grote M. 2004. What contributes to the combined effect of a complex mixture? *Environ Sci Technol* 38:6353–6362.
- Amdur MO, Doull J, Klaassen CD, editors. 1991. *Casarett & Doull's toxicology: the basic science of poisons*. 4th ed. New York: McGraw-Hill.
- Amweg EL. 2006. Effect of piperonyl butoxide on permethrin toxicity in the amphipod *Hyalella azteca*. *Environ Toxicol Chem* 25:1817–1825.
- Andersen ME. 1995. Development of physiologically based pharmacokinetic and physiologically based pharmacodynamic models for applications in toxicology and risk assessment. *Toxicol Lett* 79:35–44.
- Andersen ME, Birnbaum LS, Barton HA, Eklund CR. 1997. Regional hepatic CYP1A1 and CYP1A2 induction with 2,3,7,8-tetrachlorodibenzo-p-dioxin evaluated with a multicompartiment geometric model of hepatic zonation. *Toxicol Appl Pharmacol* 144:145–155.
- Andersen ME, Clewell HJ III. 1983. Pharmacokinetic interaction of mixtures. In: *Proceedings of the 14th Annual Conference on Environmental Toxicology*. AFAMRL-TR-83-099. Dayton (OH): AFAMRL p 226–238.
- Andersen ME, Dennison JE. 2004. Mechanistic approaches for mixture risk assessments—present capabilities with simple mixtures and future directions. *Environ Toxicol Pharmacol* 16:1–11.
- Andersen ME, Gargas ML, Clewell HJ III, Severyn KM. 1987. Quantitative evaluation of the metabolic interactions between trichloroethylene and 1,1-dichloroethylene *in vivo* using gas uptake methods. *Toxicol Appl Pharmacol* 89:149–157.
- Ankley GT, Dierkes JR, Jensen DA, Peterson GS. 1991. Piperonyl butoxide as a tool in aquatic toxicological research with organophosphate insecticides. *Ecotoxicol Environ Safety* 21:266–274.
- Ankley GT, Schubauer-Berigan MK, Hoke RA. 2006. Use of toxicity identification techniques to identify dredged material disposal options: a proposed approach. *Environ Manage* 16:1–6.
- Archer V. 1985. Enhancement of lung cancer by cigarette smoking in uranium and other miners. *Carcinogenesis* 8:23–37.
- Arrhenius Å, Grönvall F, Scholze M, Backhaus T, Blanck H. 2004. Predictability of the mixture toxicity of 12 similarly acting congeneric inhibitors of photosystem II in marine periphyton and epipsammon communities. *Aquat Toxicol* 68:351–367.
- Arts GHP, Buijse-Bogdan LL, Belgers JDM, Van Rhenen-Kersten CH, Van Wijngaarden RPA, Roessing I, Maund SJ, Van den Brink PJ, Brock TCM. 2006. Ecological impact in ditch mesocosms of simulated spray drift from a crop protection program for potatoes. *IEAM* 2:105–125.
- Ashauer R, Boxall A, Brown C. 2006. Predicting effects on aquatic organisms from fluctuating or pulsed exposure to pesticides. *Environ Toxicol Chem* 25:1899–1912.
- Ashauer R, Boxall ABA, Brown CD. 2007a. Modeling combined effects of pulsed exposure to carbaryl and chlorpyrifos on *Gammarus pulex*. *Environ Sci Technol* 41:5535–5541.
- Ashauer R, Boxall ABA, Brown CD. 2007b. New ecotoxicological model to simulate survival of aquatic invertebrates after exposure to fluctuating and sequential pulses of pesticides. *Environ Sci Technol* 41:1480–1486.
- Ashford JR. 1981. General models for the joint action of mixtures of drugs. *Biometrics* 37:457–474.
- ATSDR. 1997. Toxicological profile for chlorpyrifos. Agency for Toxic Substances and Disease Registry. Atlanta (GA): Department of Health and Human Services, Public Health Service. Available from: <http://www.atsdr.cdc.gov/toxpro2.html>
- ATSDR. 1998a. Toxicological profile for chlorinated dibenzo-p-dioxins (CDDs). Agency for Toxic Substances and Disease Registry. Atlanta (GA): US Department of Health and Human Services, Public Health Service. Available from: <http://www.atsdr.cdc.gov/toxpro2.html>

- ATSDR. 1998b. Toxicological profile for radon 1990. Agency for Toxic Substances and Disease Registry. Atlanta (GA): US Department of Health and Human Services, Public Health Service. Available from: <http://www.atsdr.cdc.gov/toxpro2.html>
- ATSDR. 1999. Toxicological profile for lead. Agency for Toxic Substances and Disease Registry. Atlanta (GA): US Department of Health and Human Services, Public Health Service. Available from: <http://www.atsdr.cdc.gov/toxpro2.html>
- ATSDR. 2004a. Guidance manual for the assessment of joint toxic action of chemical mixtures. Agency for Toxic Substances and Disease Registry. Atlanta (GA): US Department of Health and Human Services. Available from: <http://www.atsdr.cdc.gov/interaction-profiles/ipga.html>
- ATSDR. 2004b. Interaction profile for persistent chemicals found in breast milk (chlorinated dibenzo-p-dioxins, hexachlorobenzene, p,p'-DDE, methylmercury and polychlorinated biphenyls). Agency for Toxic Substances and Disease Registry. Atlanta (GA): US Department of Health and Human Services, Public Health Service.
- ATSDR. 2004c. Interaction profile for arsenic, cadmium, chromium and lead. Agency for Toxic Substances and Disease Registry. Atlanta (GA): US Department of Health and Human Services, Public Health Service.
- ATSDR. 2006. Interaction profile for chlorpyrifos, lead, mercury, and methylmercury. Agency for Toxic Substances and Disease Registry. Atlanta (GA): US Department of Health and Human Services, Public Health Service.
- Aylward LL, Hays S, Finley B. 2002. Temporal trends in intake of dioxins from foods in the U.S. and Western Europe: issues with intake estimates and parallel trends in human body burdens. 22nd Int Symp Halogenated Environ Organic Pollutants POPs 55:235–238.
- Aylward LL, Hays SM. 2002. Temporal trends in human TCDD body burden: decreases over three decades and implications for exposure levels. J Expo Anal Environ Epidemiol 12:319–328.
- Baas J, Jager T, Kooijman SALM. 2009. A model to analyse effects of complex mixtures on survival. Ecotoxicol Environ Safety 72:669–676.
- Baas J, Van Houte BPP, Van Gestel CAM, Kooijman SALM 2007. Modelling the effects of binary mixtures on survival in time. Environ Toxicol Chem 26:1320–1327.
- Bachmann KA, Ghosh R. 2001. The use of *in vitro* methods to predict *in vivo* pharmacokinetics and drug interactions. Curr Drug Metab 2:299–314.
- Backhaus T, Altenburger R, Boedeker W, Faust M, Scholze M, Grimme LH. 2000a. Predictability of the toxicity of a multiple mixture of dissimilarly acting chemicals to *Vibrio fischeri*. Environ Toxicol Chem 19:2348–2356.
- Backhaus T, Arrhenius A, Blanck H. 2004. Toxicity of a mixture of dissimilarly acting substances to natural algal communities: predictive power and limitations of independent action and concentration addition. Environ Sci Technol 38:6363–6370.
- Backhaus T, Scholze M, Grimme LH. 2000b. The single substance and mixture toxicity of quinolones to the bioluminescent bacterium *Vibrio fischeri*. Aquat Toxicol 49:49–61.
- Bakker MI, Vorenhout M, Sijm DTHM, Kolloffel C. 1999. Dry deposition of atmospheric polycyclic aromatic hydrocarbons in three *Plantago* species. Environ Toxicol Chem 18:2289–2294.
- Balakin KV, Ekins S, Bugrim A, Ivanenkov YA, Korolev D, Nikolsky YV, Skorenko AV, Ivashchenko AA, Savchuk NP, Nikolskaya T. 2004. Kohonen maps for prediction of binding to human cytochrome P450 3A4. Drug Metab Dispos 32:1183–1189.
- Banks KE, Wood SH, Matthews C, Thuesen KA. 2003. Joint acute toxicity of diazinon and copper to *Ceriodaphnia dubia*. Environ Toxicol Chem 22:1562–1567.
- Barahona LM, Loyo L, Guerrero M, Ramírez S, Romero I, Jarquin CV, Albores A. 2005. Ecotoxicological evaluation of diesel-contaminated soil before and after a bioremediation process. Environ Toxicol 20:100–109.

- Barata C, Markich SJ, Baird DJ, Taylor G, Soares AMVM. 2002. Genetic variability in sub-lethal tolerance to mixtures of cadmium and zinc in clones of *Daphnia magna* Straus. *Aquat Toxicol* 60:85–99.
- Barber MC. 2003. A review and comparison of models for predicting dynamic chemical bio-concentration in fish. *Environ Toxicol Chem* 22:1963–1992.
- Bargagli R, Monaci F, Borghini F, Bravi F, Agnorelli C. 2002. Mosses and lichens as biomonitors of trace metals: a comparison study on *Hypneum cupressiforme* and *Parmelia caperata* in a former mining district in Italy. *Environ Pollut* 116:279–287.
- Barton CN. 1993. Nonlinear statistical models for the joint action of toxins. *Biometrics* 49:95–105.
- Barton HA, Creech JR, Godin CS, Randall GM, Seckel CS. 1995. Chloroethylene mixtures: pharmacokinetic modeling and *in vitro* metabolism of vinyl chloride, trichloroethylene, and trans-1,2-dichloroethylene in rat. *Toxicol Appl Pharmacol* 130:237–247.
- Baumard P, Budzinski H, Garrigues P, Sorbe JC, Burgeot T, Bellocq J. 1998. Concentrations of PAHs (polycyclic aromatic hydrocarbons) in various marine organisms in relation to those in sediments and to trophic level. *Mar Pollut Bull* 36:951–960.
- Bedaux JJM, Kooijman SALM. 1994. Statistical analysis of bioassays based on hazard modeling. *Environ Ecol Stat* 1:303–314.
- Belden JB, Gilliom RJ, Lydy MJ. 2007. How well can we predict the toxicity of pesticide mixtures to aquatic life. *Integrated Environ Assess Manage* 3:362–372.
- Belden JB, Lydy MJ. 2006. Joint toxicity of chlorpyrifos and esfenvalerate to fathead minnows and midge larvae. *Environ Toxicol Chem* 25:623–629.
- Belfroid A, Sikkenk M, Seinen W, Van Gestel K, Hermens J. 1994. The toxicokinetic behavior of chlorobenzenes in earthworm (*Eisenia andrei*) experiments in soil. *Environ Toxicol Chem* 13:93–99.
- Beliveau M, Krishnan K. 2005. A spreadsheet program for modeling quantitative structure-pharmacokinetic relationships for inhaled volatile organics in humans. *SAR QSAR Environ Res* 16:63–77.
- Beliveau M, Lipscomb J, Tardif R, Krishnan K. 2005. Quantitative structure-property relationships for interspecies extrapolation of the inhalation pharmacokinetics of organic chemicals. *Chem Res Toxicol* 18:475–485.
- Beliveau M, Tardif R, Krishnan K. 2003. Quantitative structure-property relationships for physiologically based pharmacokinetic modeling of volatile organic chemicals in rats. *Toxicol Appl Pharmacol* 189:221–232.
- Belz RG, Cedergreen N, Sørensen H. 2008. Hormesis in mixtures—can it be predicted? *Sci Total Environ* 404:77–87.
- Berenbaum MC. 1985. The expected effect of a combination of agents: the general solution. *J Theor Biol* 114:413–431.
- Berenbaum MC. 1989. What is synergy? *Pharmacol Rev* 1989:93–141.
- Berger U, Herzke D, Sandanger TM. 2004. Two trace analytical methods for determination of hydroxylated PCBs and other halogenated phenolic compounds in eggs from Norwegian birds of prey. *Anal Chem* 76:441–452.
- Berkowitz GS, Obel J, Deych E, Lapinski R, Godbold J, Liu Z, Landrigan PJ, Wolff MS. 2003. Exposure to indoor pesticides during pregnancy in a multiethnic, urban cohort. *Environ Health Perspect* 111:79–84.
- Bernillon P, Bois FY. 2000. Statistical issues in toxicokinetic modeling: a bayesian perspective. *Environ Health Perspect* 108(Suppl 5):883–893.
- Bervoets L, Meregalli G, De Cooman W, Goddeeris B, Blust R. 2004. Caged midge larvae (*Chironomus riparius*) for the assessment of metal bioaccumulation from sediments *in situ*. *Environ Toxicol Chem* 23:443–454.
- BFR (German Federal Institute for Risk Assessment). 2005. Documents of the BfR Forum on multiple residues of pesticide residues in food. Available from: <http://www.bfr.bund.de/cd/7078>

- Binder S, Sokal D, Maughn D. 1986. The use of tracer elements in estimating the amount of soil ingested by young children. *Arch Environ Health* 41:341–345.
- Binderup ML, Dalgaard M, Dragsted LO, Hossaini A, Ladefoged O, Lam HR, Larsen JC, Madsen C, Meyer O, Rasmussen ES, Reffstrup TK, Soborg I, Vinggaard AM, Ostergard G. 2003. Combined actions and interactions of chemicals in mixtures: the toxicological effects to exposures of mixtures of industrial and environmental chemicals. Report 2003:12. Soborg (DK): Danish Veterinary and Food Administration.
- Bisnoti MC, Jardim WF. 2003. Production of organic mercury from Hg⁰: experiments using microcosms. *J Brazil Chem Soc* 14:244–248.
- Bjorkman S. 2005. Prediction of drug disposition in infants and children by means of physiologically based pharmacokinetic (PBPK) modelling: theophylline and midazolam as model drugs. *Br J Clin Pharmacol* 59:691–704.
- Blanck H. 2002. A critical review of procedures and approaches used for assessing pollution-induced community tolerance (PICT) in biotic communities. *Human Ecol Risk Assess* 8:1003–1034.
- Bliss CI. 1939. The toxicity of poisons applied jointly. *Ann J Appl Biol* 26:585–615.
- Bobbink R, Heil GW, Raessen M. 1992. Atmospheric deposition and canopy exchange processes in heathland ecosystems. *Environ Pollut* 75:29–37.
- Bocquene G, Bellanger C, Cadiou Y, Galgani F. 1995. Joint action of combinations of pollutants on the acetylcholinesterase activity of several marine species. *Ecotoxicology* 4:266–279.
- Boedeker W, Altenburger R, Faust M, Grimme LH. 1990. Methods for the assessment of mixtures of plant protection substances (pesticides): mathematical analysis of combination effects in phytopharmacology and ecotoxicology. *Nachrichtenblatt Deutschen Pflanzensch* 42:70–78.
- Boedeker W, Altenburger R, Faust M, Grimme LH. 1992. Synopsis of concepts and models for the quantitative analysis of combination effects: from biometrics to ecotoxicology. *ACES* 4:45–53.
- Bond JA, Csanady GA, Gargas ML, Guengerich FP, Leavens T, Medinsky MA, Recio L. 1994. 1,3-Butadiene: linking metabolism, dosimetry, and mutation induction. *Environ Health Perspect* 102(Suppl 9):87–94.
- Boom SP, Meyer I, Wouterse AC, Russel FG. 1998. A physiologically based kidney model for the renal clearance of ranitidine and the interaction with cimetidine and probenecid in the dog. *Biopharm Drug Dispos* 19:199–208.
- Borgert CJ. 2007. Predicting interactions from mechanistic information: can omic data validate theories? *Toxicol Appl Pharmacol* 223:114–120.
- Boxall ABA, Maltby L. 1997. The effects of motorway runoff on freshwater ecosystems. 3. Toxicant confirmation. *Arch Environ Contam Toxicol* 33:9–16.
- Boyd EM. 1969. Dietary protein and pesticide toxicity in male weaning rats. *Bull WHO* 40:801–805.
- Brack W, Altenburger R, Ensenbach U, Moder M, Segner H, Schüürmann G. 1999. Bioassay-directed identification of organic toxicants in river sediment in the industrial region of Bitterfeld (Germany)—a contribution to hazard assessment. *Arch Environ Contam Toxicol* 37:164–174.
- Brian JV, Harris CA, Scholze M, Backhaus T, Booy P, Lamoree M, Pojana G, Jonkers N, Runnalls T, Bonfa A, Marcomini A, Sumpter JP. 2005. Accurate prediction of the response of freshwater fish to a mixture of estrogenic chemicals. *Environ Health Perspect* 113:721–728.
- Brocklebank JR, Namdari R, Law FC. 1997. An oxytetracycline residue depletion study to assess the physiologically based pharmacokinetic (PBPK) model in farmed Atlantic salmon. *Can Vet J* 38:645–646.
- Broderius S, Kahl M. 1985. Acute toxicity of organic chemical mixtures to the fathead minnow. *Aquat Toxicol* 6:307–322.

- Broughton RK, Osborn D, Shore RF, Wienburg CL, Wadsworth RA. 2003. Identifying pollution hot spots from polychlorinated biphenyl residues in birds of prey. *Environ Toxicol Chem* 22:2519–2524.
- Brown RP, Delp MD, Lindstedt SL, Rhomberg LR, Beliles RP. 1997. Physiological parameter values for physiologically based pharmacokinetic models. *Toxicol Ind Health* 13:407–484.
- Bulletti C, Flamigni C, Giacomucci E. 1996. Reproductive failure due to spontaneous abortion and recurrent miscarriage. *Hum Reprod Update* 2:118–136.
- Bundy JG, Sidhu JK, Rana F, Spurgeon DJ, Svendsen C, Wren JF, Stürzenbaum SR, Morgan AJ, Kille P. 2008. “Systems toxicology” approach identifies coordinated metabolic responses to copper in a terrestrial non-model invertebrate, the earthworm *Lumbricus rubellus*. *BMC Biol* 6:25 (doi:10.1186/1741-7007-6-25).
- Burkhard LP, Durhan EJ. 1991. Identification of nonpolar toxicants in effluents using toxicity-based fractionation with gas chromatography/mass spectrometry. *Anal Chem* 63:277–283.
- Burton KW, Morgan E, Roig A. 1986. Interactive effects of cadmium, copper and nickel on the growth of Sitka Spruce and studies of metal uptake from nutrient solutions. *New Phytol* 103:549–557.
- Cahill TM, Cousins I, Mackay D. 2003. General fugacity-based model to predict the environmental fate of multiple chemical species. *Environ Toxicol Chem* 22:483–493.
- Calabrese EJ. 1991. Multiple chemical interactions. Part 4: Drugs; Part 5: The drug-pollutant interface. Chelsea (MI): Lewis Publishers, p 389–578.
- Calabrese EJ. 2005. Paradigm lost, paradigm found: the re-emergence of hormesis as a fundamental dose response model in the toxicological sciences. *Environ Pollut* 138:8–411.
- Callaghan A, Hirthe G, Fisher T, Crane M. 2001. Effect of short-term exposure to chlorpyrifos on developmental parameters and biochemical biomarkers in *Chironomus riparius* Meigen. *Ecotoxicol Environ Safety* 50:19–24.
- Cape JN, Freersmith PH, Paterson IS, Parkinson JA, Wolfenden J. 1990. The nutritional status of *Picea abies* (L) Karst across Europe, and implications for forest decline. *Trees Struct Funct* 4:211–224.
- Carlile DJ, Zomorodi K, Houston JB. 1997. Scaling factors to relate drug metabolic clearance in hepatic microsomes, isolated hepatocytes, and the intact liver: studies with induced livers involving diazepam. *Drug Metab Dispos* 25:903–911.
- Casey M, Gennings C, Carter WH Jr, Moser VC, Simmons JE. 2005. Ds-optimal designs for studying combinations of chemicals using multiple fixed-ratio ray experiments. *Environmetrics* 16:129–147.
- Cassee FR, Groten JP, Van Bladeren PJ, Feron VJ. 1998. Toxicological evaluation and risk assessment of chemical mixtures. *Crit Rev Toxicol* 28:73–101.
- CDC. 2003. Second national report on human exposure to environmental chemicals. CDC/NCEH Pub. 02-0716. Atlanta (GA): Department of Health and Human Services, Centers for Disease Control and Prevention.
- CDC. 2005. Third national report on human exposure to environmental chemicals. CDC/NCEH Pub. 05-0570. Atlanta (GA): Department of Health and Human Services, Centers for Disease Control and Prevention.
- Cedergreen N, Kudsk P, Mathiassen SK, Sørensen H, Streibig JC. 2007. Reproducibility of binary-mixture toxicity studies. *Environ Toxicol Chem* 26:149–156.
- Cedergreen N, Streibig JC. 2005. Can the choice of endpoint lead to contradictory results of mixture-toxicity experiments? *Environ Toxicol Chem* 24:1676–1683.
- Cerklewski FL, Forbes RM. 1976. Influence of dietary zinc on lead toxicity in the rat. *J Nutr* 106:689–696.
- Chapman PM. 1986. Sediment quality criteria from the sediment quality TRIAD—an example. *Environ Toxicol Chem* 5:957–964.

- Chapman PM, McDonald BG, Lawrence GS. 2002. Weight-of-evidence issues and frameworks for sediment quality (and other) assessments. *Human Ecol Risk Assess* 8:1489–1515.
- Chapman PM, Power EA, Burton GA Jr. 1992. Integrative assessments in aquatic ecosystems. In: Burton GA Jr, editor, *Sediment toxicity assessment*. Chelsea (MI): Lewis Publishers. p 313–340.
- Chèvre N, Loeffe C, Singer H, Stamm C, Fenner K, Escher BI. 2006. Including mixtures in the determination of water quality criteria for herbicides in surface water. *Environ Sci Technol* 40:426–435.
- Chien JY, Thumme KE, Slattery JT. 1997. Pharmacokinetic consequences of induction of CYP2E1 by ligand stabilization. *Drug Metab Dispos* 25:1165–1175.
- Cho EA, Bailer J, Oris JT. 2003. Effect of methyl tert-butyl ether on the bioconcentration and photoinduced toxicity of fluoranthene in fathead minnow larvae (*Pimephales promelas*). *Environ Sci Technol* 37:1306–1310.
- Chou TC, Talalay P. 1983. Analysis of combined drug effects—a new look at a very old problem. *Trends Pharmacol Sci* 4:450–454.
- Ciucu A. 2002. Progress and perspectives in biosensors for environmental monitoring. *Roum Biotechnol Lett* 7:537–546.
- Clark S, Bornschein RL, Pan W, Menrath W, Roda S, Grote J. 1996. The relationship between surface dust lead loadings on carpets and the blood lead of young children. *Environ Geochem Health* 18:143–146.
- Clausing P, Brunekeef B, Van Wijnen JH. 1987. A method for estimating soil ingestion by children. *Int Arch Occup Environ Health* 59:73–82.
- Clewell HJ III, Andersen ME. 1985. Risk assessment extrapolations and physiological modeling. *Toxicol Ind Health* 1:111–131.
- Clewell RA, Merrill EA, Yu KO, Mahle DA, Sterner TR, Mattie DR, Robinson PJ, Fisher JW, Gearhart JM. 2003. Predicting fetal perchlorate dose and inhibition of iodide kinetics during gestation: a physiologically-based pharmacokinetic analysis of perchlorate and iodide kinetics in the rat. *Toxicol Sci* 73:235–255.
- Clifford PA, Barchers DE, Ludwig DF, Sielken RL, Klingensmith JS, Graham RV, Banton MI. 1995. An approach to quantifying spatial components of exposure for ecological risk assessment. *Environ Toxicol Chem* 14:895–906.
- Coffey T, Gennings C, Simmons JE, Herr WD. 2005. D-Optimal experimental designs to test for departure from additivity in a fixed-ratio mixture ray. *Toxicol Sci* 88:467–476.
- Cogliano VJ. 1997. Plausible upper bounds: are their sums plausible? *Risk Anal* 17:77–84.
- Cogliano VJ. 1998. Assessing the cancer risk from environmental PCBs. *Environ Health Perspect* 106:317–323.
- Coombe VT, Moore KW, Hutchings MJ. 1999. TIE and TRE: an abbreviated guide to dealing with toxicity. *Water Sci Technol* 39:91–97.
- COT. 2002. Risk assessment of mixtures of pesticides and similar substances. London: Committee on Toxicity of Chemicals in Food, Consumer Products and the Environment, UK Food Standards Agency. Available from: <http://cot.food.gov.uk/pdfs/cotwig2000-2.pdf>
- Cotter MA, Policz DL, Pösch G, Dawson DA. 2000. Analysis of the combined osteolathyritic effects of beta-aminopropionitrile and diethyldithiocarbamate on *Xenopus* development. *Toxicol Sci* 58:144–152.
- Cova D, Nebuloni C, Arnoldi A, Bassoli A, Trevisan M, DelRe AAM. 1996. N-nitrosation of triazines in human gastric juice. *J Agric Food Chem* 44:2852–2855.
- Covington TR, Gentry PR, Van Landingham CB, Andersen ME, Kester JE, Clewell HJ. 2007. The use of Markov chain Monte Carlo uncertainty analysis to support a public health goal for perchloroethylene. *Regul Toxicol Pharmacol* 47:1–18.
- Cowan CE, Versteeg DJ, Larson RJ, Kloeppersams PJ. 1995. Integrated approach for environmental assessment of new and existing substances. *Regul Toxicol Pharmacol* 21:3–31.

- Crofton K, Craft ES, Hedge JM, Gennings C, Simmons JE, Carchman RA, Carter WH Jr, deVito JM. 2005. Thyroid-hormone-disrupting chemicals: evidence for dose-dependent additivity or synergism. *Environ Health Perspect* 113:1549–1554.
- CSTEE. 2000. The available scientific approaches to assess the potential effects and risk of chemicals on terrestrial ecosystems. C2/JCD/csteeop/Ter91100/D(0). Brussels (BE): European Commission. p 178.
- Cui Y, Zhu Y-G, Zhai R, Huang Y, Qiu Y, Liang J. 2005. Exposure to metal mixtures and human health impacts in a contaminated area in Nanning, China. *Environ Intern* 31:784–790.
- Cuppen JGM, Crum SJH, Van den Heuvel HH, Smidt RA, Van den Brink PJ. 2002. The effects of a mixture of two insecticides on freshwater microcosms. I. Fate of insecticides and responses of macroinvertebrates. *Ecotoxicology* 11:19–34.
- Damgaard IN, Skakkebaek NE, Toppari J, Virtanen HE, Shen H, Schramm KW, Petersen JH, Jensen TK, Main KM. 2006. Persistent pesticides in human breast milk and cryptorchidism. *Environ Health Perspect* 114:1133–1138.
- Daskalakis KD. 1996. Variability of metal concentrations in oyster tissue and implications to biomonitoring. *Mar Pollut Bull* 32:794–801.
- David RM, Clewell HJ, Gentry PR, Covington TR, Morgott DA, Marino DJ. 2006. Revised assessment of cancer risk to dichloromethane. II. Application of probabilistic methods to cancer risk determinations. *Regul Toxicol Pharmacol* 45:55–65.
- Decaprio AP. 1997. Biomarkers: coming of age for environmental health and risk assessment. *Environ Sci Technol* 31:1837–1848.
- De Graaf C, Vermeulen NP, Feenstra KA. 2005. Cytochrome P450 in silico: an integrative modeling approach. *J Med Chem* 48:2725–2755.
- De Groot MH. 1986. Probability and statistics. 2nd ed. Reading (MA): Addison-Wesley Pub.
- De Groot MJ, Ekins S. 2002. Pharmacophore modeling of cytochromes P450. *Adv Drug Deliv Rev* 54:367–383.
- De Groot MJ, Kirton SB, Sutcliffe MJ. 2004. *In silico* methods for predicting ligand binding determinants of cytochromes P450. *Curr Top Med Chem* 4:1803–1824.
- DeKoning EP, Karmaus W. 2000. PCB exposure *in utero* and via breast milk: a review. *J Expo Anal Environ Epidemiol* 10:285–293.
- De Maagd PGJ, Van de Klundert ICM, Van Wezel AP, Opperhuizen A, Sijm DTHM. 1997. Lipid content and time-to-death-dependent lethal body burdens of naphthalene and 1,2,4-trichlorobenzene in fathead minnow (*Pimephales promelas*). *Ecotoxicol Environ Safety* 38:232–237.
- De March BGE. 1987. Simple similar action and independent joint action—two similar models for the joint effects of toxicants applied as mixtures. *Aquat Toxicol* 9:291–304.
- Deneer JW. 2000. Toxicity of mixtures of pesticides in aquatic systems. *Pest Manage Sci* 56:516–520.
- Dennison JE, Andersen ME, Clewell HJ, Yang RSH. 2004a. Development of a physiologically based pharmacokinetic model for volatile fractions of gasoline using chemical lumping analyses. *Environ Sci Technol* 38:5674–5681.
- Dennison JE, Andersen ME, Dobrev ID, Mumtaz MM, Yang RSH. 2004b. PBPK modeling of complex hydrocarbon mixtures: gasoline. *Environ Toxicol Pharmacol* 16:107–119.
- Dennison JE, Andersen ME, Yang RSH. 2003. Characterization of the pharmacokinetics of gasoline using PBPK modeling with a complex mixture chemical lumping approach. *Inhalation Toxicol* 15:961–968.
- De Rosa CT, El-Masri HE, Pohl H, Cibus W, Mumtaz MM. 2004. Implications of chemical mixtures for public health practice. *J Toxicol Environ Health* 7:339–350.
- De Rosa CT, Hansen H, Wilbur S, Pohl HR, El-Masri HA, Mumtaz MM. 2001. Interactions. In: Bingham E, Cohrssen B, Powell C, editors, *Patty's toxicology*. Vol. 1. New York: John Wiley & Sons, p 233–284.

- De Zwart D. 2005. Ecological effects of pesticide use in the Netherlands: modeled and observed effects in the field ditch. *IEAM* 1:123–134.
- De Zwart D, Dyer SD, Posthuma L, Hawkins CP. 2006. Use of predictive models to attribute potential effects of mixture toxicity and habitat alteration on the biological condition of fish assemblages. *Ecol Appl* 16:1295–1310.
- De Zwart D, Posthuma L. 2005. Complex mixture toxicity for single and multiple species: proposed methodologies. *Environ Toxicol Chem* 24:2665–2676.
- De Zwart D, Rutgers M, Notenboom J. 1998. Assessment of site-specific ecological risks of soil contamination: a design of an assessment methodology. Report nr 711701011. Bilthoven (NL): National Institute for Public Health and the Environment (RIVM).
- De Zwart D, Sterkenburg A. 2002. Toxicity-based assessment of water quality. In: Posthuma L, Suter GW II, Traas TP, editors, *Species sensitivity distributions in ecotoxicology*. Boca Raton (FL): Lewis Publishers, p 383–402.
- Dobrev I, Andersen ME, Yang RSH. 2001. Assessing interaction thresholds for trichloroethylene, tetrachloroethylene, and 1,1,1-trichloroethane using gas uptake studies and PBPK modeling. *Arch Toxicol* 75:134–144.
- Dobrev I, Andersen ME, Yang RSH. 2002. *In silico* toxicology: simulating interaction thresholds for human exposure to mixtures of trichloroethylene, tetrachloroethylene, and 1,1,1-trichloroethane. *Environ Health Perspect* 110:1031–1039.
- Dobson PD, Kell DB. 2008. Carrier-mediated cellular uptake of pharmaceutical drugs: an exception or the rule? *Nature Rev Drug Discovery* 7:205–220.
- Dorne JLCM. 2007. Human variability in hepatic and renal elimination: implications for risk assessment. *J Appl Toxicol* 27:411–420.
- Dorne JLCM, Papadopoulos A. 2008. Do uncertainty factors take into account toxicokinetic interactions? Conclusions and recommendations from the sixth framework project NOMIRACLE. *Toxicol Lett* 180:S90–S90.
- Dorne JLCM, Ragas AMJ, Frampton GK, Spurgeon DS, Lewis DF. 2007b. Trends in human risk assessment of pharmaceuticals. *Anal Bioanal Chem* 387:1167–1172.
- Dorne JLCM, Ragas AMJ, Lokke H. 2006. Harmonisation of uncertainty factors in human and ecological risk assessment. *Toxicology* 226:77–78.
- Dorne JLCM, Skinner L, Frampton GK, Spurgeon DJ, Ragas AMJ. 2007a. Human and environmental risk assessment of pharmaceuticals: differences, similarities, lessons from toxicology. *Anal Bioanal Chem* 387:1259–1268.
- Dorne JLCM, Walton K, Renwick AG. 2003. Human variability in CYP3A4 metabolism and CYP3A4-related uncertainty, factors for risk assessment. *Food Chem Toxicol* 41:201–224.
- Dorne JLCM, Walton K, Renwick AG. 2005. Human variability in xenobiotic metabolism and pathway-related uncertainty factors for chemical risk assessment: a review. *Food Chem Toxicol* 43:203–216.
- Drescher K, Bödeker W. 1995. Assessment of the combined effects of substances—the relationship between concentration addition and independent action. *Biometrics* 51:716–730.
- D'Souza RW, Francis WR, Andersen ME. 1988. Physiological model for tissue glutathione depletion and increased resynthesis after ethylene dichloride exposure. *J Pharmacol Exp Ther* 245:563–568.
- Durhan EJ, Norberg-King TJ, Burkhard LP. 1993. Methods for aquatic toxicity identification evaluations. Phase II toxicity identification evaluation procedures for samples exhibiting acute and chronic toxicity. EPA/600/R-92/080. Duluth (MN): Environmental Research Laboratory, Office of Research and Development, US Environmental Protection Agency.
- EC. 2001. White paper: strategy for a future Chemicals Policy Commission of the European Communities. COM(2001) 88 final. Brussels (BE).

- EC. 2003. Technical guidance document on risk assessment. Ispra (IT): European Chemicals Bureau (ECB), Institute for Health and Consumer Protection, European Commission, Joint Research Centre.
- ECB. 2003a. Technical guidance document on risk assessment in support of Commission Directive 93/67/EEC on risk assessment for new notified substances; Commission Regulation (EC) No. 1488/94 on risk assessment for existing substances; Directive 98/8/EC of the European Parliament and of the Council concerning the placing of biocidal products on the market. Part II. Environmental risk assessment. Ispra (IT): European Commission–Joint Research Centre, Institute for Health and Consumer Protection, European Chemicals Bureau (ECB), Chap 3, Appendix 1.
- ECB. 2003b. Technical guidance document on risk assessment in support of Commission Directive 93/67/EEC on risk assessment for new notified substances; Commission Regulation (EC) No. 1488/94 on risk assessment for existing substances; Directive 98/8/EC of the European Parliament and of the Council concerning the placing of biocidal products on the market. Part IV. Emission scenario documents. Ispra (IT): European Commission–Joint Research Centre, Institute for Health and Consumer Protection, European Chemicals Bureau (ECB), Chap 7.
- Egeler P, Römbke J, Meller M, Knacker T, Nagel R. 1999. Bioaccumulation test with tubificid sludgeworms in artificial media—development of a standardisable method. *Hydrobiologia* 406:271–280.
- Egnell AC, Houston JB, Boyer CS. 2005. Predictive models of CYP3A4 heteroactivation: *in vitro-in vivo* scaling and pharmacophore modeling. *J Pharmacol Exp Ther* 312:926–937.
- Eide I, Neverdal G, Thorvaldsen B, Grung B, Kvalheim OM. 2002. Toxicological evaluation of complex mixtures by pattern recognition: correlating chemical fingerprints to mutagenicity. *Environ Health Perspect* 110(Suppl 6):985–988.
- Ekins S. 2003. *In silico* approaches to predicting drug metabolism, toxicology and beyond. *Biochem Soc Trans* 31(Pt 3):611–614.
- Ekins S, Andreyev S, Ryabov A, Kirillov E, Rakhmatulin EA, Sorokina S, Bugrim A, Nikolskaya T. 2006. A combined approach to drug metabolism and toxicity assessment. *Drug Metab Dispos* 34:495–503.
- El-Masri HA, Constan AA, Ramsdell HS, Yang RSH. 1996b. Physiologically based pharmacodynamic modeling of an interaction threshold between trichloroethylene and 1,1-dichloroethylene in Fischer 344 rats. *Toxicol Appl Pharmacol* 141:124–132.
- El-Masri HA, Tessari JD, Yang RSH. 1996c. Exploration of an interaction threshold for the joint toxicity of trichloroethylene and 1,1-dichloroethylene: utilization of a PBPK model. *Arch Toxicol* 70:527–539.
- El-Masri HA, Thomas RS, Sabados GR, Phillips JK, Constan AA, Benjamin SA, Andersen ME, Mehendale HM, Yang RSH. 1996a. Physiologically based pharmacokinetic/pharmacodynamic modeling of the toxicologic interaction between carbon tetrachloride and kepone. *Arch Toxicol* 70:704–713.
- Emond C, Charbonneau M, Krishnan K. 2005. Physiologically based modeling of the accumulation in plasma and tissue lipids of a mixture of PCB congeners in female Sprague-Dawley rats. *J Toxicol Environ Health A* 68:1393–1412.
- Escher BI, Hunziker RW, Schwarzenbach RP. 2001. Interaction of phenolic uncouplers in binary mixtures: concentration-additive and synergistic effects. *Environ Sci Technol* 35:3905–3914.
- Escher BI, Sigg L. 2004. Chemical speciation of organics and of metals at biological interfaces. In: Van Leeuwen HP, Köster W, editors, *Physicochemical kinetics and transport at biointerfaces*. Vol. 9. Chichester (UK): John Wiley. p 205–271.

- Evans JS, Gray GM, Sielken RLJ, Smith AE, Valdez FC, Graham JD, 1994. Use of probabilistic expert judgment in uncertainty analysis of carcinogenic potency. *Reg Toxicol Pharmacol* 20:15–36.
- Faessel HM, Slocum HK, Rustum YM, Greco WR. 1999. Folic acid-enhanced synergy for the combination of trimetrexate plus the glycinamide ribonucleotide formyltransferase inhibitor 4-[2-(2-amino-4-oxo-4,6,7,8-tetrahydro-3H-pyrimidino[5,4,6][1,4]thiazin-6-yl)-(S)-ethyl]-2,5-thienoyl amino-L-glutamic acid (AG2034)—comparison across sensitive and resistant human tumor cell lines. *Biochem Pharmacol* 57:567–577.
- Fairman R, Mead CD, Williams WP. 1998. Environmental risk assessment: approaches, experiences and information sources. Environmental Issue Report 4. Copenhagen (DK): European Environmental Agency.
- Faust M, Altenburger R, Backhaus T, Blanck H, Boedeker W, Gramatica P, Hamer V, Scholze M, Vighi M, Grimme LH. 2003. Joint algal toxicity of 16 dissimilarly acting chemicals is predictable by the concept of independent action. *Aquat Toxicol* 63:43–63.
- Faust M, Altenburger R, Backhaus T, Boedeker W, Gramatica P, Hamer V, Scholze M, Vighi M, Grimme LH. 2001. Predicting the joint algal toxicity of multi-component s-triazine mixtures at low-effect concentrations of individual toxicants. *Aquat Toxicol* 56:13–32.
- Fay M. 2005. Exposure to contaminant mixtures at US hazardous waste sites. In: Aral MM, Brebbia CA, Maslia ML, Sinks T, editors, *Environmental exposure and health*. WIT transactions on ecology and the environment. Vol. 85. Southhampton (UK): WIT Press. p 227–232.
- Fay RM, Mumtaz MM. 1996. Development of a priority list of chemical mixtures, occurring at 1188 hazardous waste sites, using HazDat database. *Food Chem Toxicol* 34:1163–1165.
- Fenner K, Scheringer M, MacLeod M, Matthies M, McKone T, Stroebe M, Beyer A, Bonnell M, Le Gall AC, Klasmeier J, Mackay D, Van De Meent D, Pennington D, Scharenberg B, Suzuki N, Wania F. 2005. Comparing estimates of persistence and long-range transport potential among multimedia models. *Environ Sci Technol* 39:1932–1942.
- Fernandes AR, Rose M, White S, Mortimer DN, Gem M. 2006. Dioxins and polychlorinated biphenyls (PCBs) in fish oil dietary supplements: occurrence and human exposure in the UK. *Food Add Contam* 23:939–947.
- Fernandez MF, Araque P, Kiviranta H, Molina-Molina JM, Rantakokko P, Laine O, Vartiainen T, Olea N. 2007a. PBDEs and PBBs in the adipose tissue of women from Spain. *Chemosphere* 66:377–383.
- Fernandez MF, Olmos B, Granada A, López-Espinosa MJ, Molina-Molina JM, Fernandez JM, Cruz M, Olea-Serrano F, Olea N. 2007b. Human exposure to endocrine disrupting chemicals and prenatal risk factors for cryptorchidism and hypospadias: a nested case-control study. *Environ Health Perspect* 115(Suppl 1):8–14.
- Feron VJ, Groten JP. 2002. Toxicological evaluation of chemical mixtures. *Food Chem Toxicol* 40:825–839.
- Feron VJ, Groten JP, van Zorge JA, Cassee FR, Jonker D, van Bladeren PJ. 1995. Toxicity studies in rats of simple mixtures of chemicals with the same or different target organs. *Toxicol Lett* 82/83:505–512.
- Ferrario J, Byrne C, Lorber M, Saunders P, Leese W, Dupuy A, Winters D, Cleverly D, Schaum J, Pinsky P, Deyrup C, Ellis R, Walcott J. 1997. A statistical survey of dioxin-like compounds in the United States poultry fat. *Organohalogen Compounds* 32:245–251.
- Ferrario J, Byrne C, McDaniel D, Dupuy A, Harless R. 1996. Determination of 2,3,7,8-chlorine substituted dibenzo-p-dioxins and furans at the part per trillion level in United States beef fat using high resolution gas chromatography/high resolution mass spectrometry. *Anal Chem* 68:647–652.
- Ferreira KL, Burton JD, Coble HD. 1995. Physiological basis for antagonism of fluzafop-p by DPX-PE350. *Weed Sci* 43:184–191.

- Fialkowski W, Klonowska-Olejnik M, Smith BD, Rainbow PS. 2003. Mayfly larvae (*Baetis rhodani* and *B. vernus*) as biomonitors of trace metal pollution in streams of a catchment draining a zinc and lead mining area of Upper Silesia, Poland. *Environ Pollut* 121:253–267.
- Filser JG, Johanson G, Kessler W, Kreuzer PE, Stei P, Baur C, Csanady GA. 1993. A pharmacokinetic model to describe toxicokinetic interactions between 1,3-butadiene and styrene in rats: predictions for human exposure. *IARC Sci Publ* 127:65–78.
- Filzek PDB, Spurgeon DJ, Broll G, Svendsen C, Hankard PK, Kammenga JE, Donker MH, Weeks JM. 2004. Pedological characterisation of sites along a transect from a primary cadmium/lead/zinc smelting works. *Ecotoxicology* 13:725–737.
- Finney DJ. 1942. The analysis of toxicity tests on mixtures of poisons. *Ann Appl Biol* 29:82–94.
- Fiore BJ, Anderson HA, Hanrahan MS, Olson LJ, Sonzogni WC. 1989. Sport fish consumption and body burden levels of chlorinated hydrocarbons: a study of Wisconsin anglers. *Arch Environ Health* 44:82–88.
- Forbes VE, Palmqvist A, Bach L. 2006. The use and misuse of biomarkers in ecotoxicology. *Environ Toxicol Chem* 25:272–280.
- Foster KL, MacKay D, Parkerton TF, Webster E, Milford L. 2005. Five-stage environmental exposure assessment strategy for mixtures: gasoline as a case study. *Environ Sci Technol* 39:2711–2718.
- Fouchecourt MO, Beliveau M, Krishnan K. 2001. Quantitative structure-pharmacokinetic relationship modelling. *Sci Total Environ* 274:125–135.
- Franchi M, Carrer P, Kotzias D, Rameckers E, Seppanen O, van Bronswijk JEMH, Viegi G, Gilder JA, Valovirta E. 2006. Working towards healthy air in dwellings in Europe. *Allergy* 61:864–868.
- Frayse B, Baudin J-P, Garnier-Laplace J, Adam C, Boudou A. 2002. Effects of Cd and Zn waterborne exposure on the uptake and depuration of ⁵⁷Co, ¹¹⁰Ag and ¹³⁴Cs by the Asiatic clam (*Corbicula fluminea*) and the zebra mussel (*Dreissena polymorpha*) — whole organism study. *Environ Pollut* 118:297–306.
- Frederick CB, Potter DW, Chang-Mateu MI, Andersen ME. 1992. A physiologically based pharmacokinetic and pharmacodynamic model to describe the oral dosing of rats with ethyl acrylate and its implications for risk assessment. *Toxicol Appl Pharmacol* 114:246–260.
- Fries GF, Marrow, GS. 1992. Influence of soil properties on the uptake of hexachlorobiphenyls by rats. *Chemosphere* 24:109–113.
- Fryer M, Collins CD, Ferrier H, Colville RN, Nieuwenhuijsen MJ. 2006. Human exposure modelling for chemical risk assessment: a review of current approaches and research and policy implications. *Environ Sci Policy* 9:261–274.
- Fulton MH, Key PB. 2001. Acetylcholinesterase inhibition in estuarine fish and invertebrates as an indicator of organophosphorus insecticide exposure and effects. *Environ Toxicol Chem* 20:37–45.
- Gabrielsson J, Weiner D. 2000. Pharmacokinetic and pharmacodynamic data analysis, concepts and applications. 3rd ed. Stockholm (SE): Swedish Pharmaceutical Press.
- Gagne F, Blaise C. 2004. Review of biomarkers and new techniques for *in-situ* aquatic studies with bivalves. In: Thompson KC, Wadhia, K, Loibner A, editors, *Environmental toxicity testing*. Sheffield Analytical Chemistry Series. Oxford (UK): Blackwell Publishing, Chap 7.
- Garcia-Ortega S, Holliman PJ, Jones DL. 2006. Toxicology and fate of Pestanal® and commercial propetamphos formulations in river and estuarine sediment. *Sci Total Environ* 366:826–836.
- Gaumont Y, Kisliuk RL, Parsons JC, Greco WR. 1992. Quantitation of folic-acid enhancement of antifolate synergism. *Cancer Res* 52:2228–2235.

- Gay JR, Korre A. 2006. A spatially-evaluated methodology for assessing risk to a population from contaminated land. *Environ Pollut* 142:227–234.
- Gelman A, Bois FY, Jiang J. 1996. Physiological pharmacokinetic analysis using population modeling and informative prior distributions. *J Am Stat Assoc* 91:1400–1412.
- Gennings C. 1995. An efficient experimental design for detecting departure from additivity in mixtures of many chemicals. *Toxicology* 105:189–197.
- Gennings C. 1996. Economical designs for detecting and characterizing departure from additivity in mixtures of many chemicals. *Food Chem Toxicol* 34:1053–1058.
- Gennings C, Carter WH. 1995. Utilizing concentration–response data from individual components to detect statistically significant departures from additivity in chemical mixtures. *Biometrics* 51:1264–1277.
- Gennings C, Carter WH, Campain JA, Bae DS, Yang RSH. 2002. Statistical analysis of interactive cytotoxicity in human epidermal keratinocytes following exposure to a mixture of four metals. *J Agric Biol Environ Stat* 7:58–73.
- Gennings C, Carter WH Jr, Carchman RA, Teuschler LK, Simmons JE, Carney EW. 2005. A unifying concept for assessing toxicological interactions: changes in slope. *Toxicol Sci* 88:287–297.
- Gennings C, Carter WH, Caseya M, Moser V, Carchman R, Simmons JE. 2004. Analysis of functional effects of a mixture of five pesticides using a ray design. *Environ Toxicol Pharmacol* 18:115–125.
- Gentry PR, Covington TR, Clewell HJ. 2003. Evaluation of the potential impact of pharmacokinetic differences on tissue dosimetry in offspring during pregnancy and lactation. *Regul Toxicol Pharmacol* 38:1–16.
- Gerhardt A, Janssens de Bisthoven L, Guhr K, Soares AMVM, Pereira MJ. 2008. Phytotoxicity assessment of acid mine drainage: *Lemna gibba* bioassay and diatom community structure. *Ecotoxicology* 17:47–58.
- Gerhardt A, Janssens de Bisthoven L, Soares AMVM. 2004. Macroinvertebrate response to acid mine drainage: community metrics and on-line behavioural toxicity bioassay. *Environ Pollut* 130:263–274.
- Gerhardt A, Janssens de Bisthoven L, Soares AMVM. 2005. Effects of acid mine drainage and acidity on the activity of *Choroterpes picteti* (Ephemeroptera). *Arch Environ Contam Toxicol* 48:450–459.
- Ginsberg G, Hattis D, Sonawane B. 2004. Incorporating pharmacokinetic differences between children and adults in assessing children's risks to environmental toxicants. *Toxicol Appl Pharmacol* 198:164–183.
- Gobas FAPC, McCorquodale JR, Haffner GD. 1993. Intestinal absorption and biomagnification of organochlorines. *Environ Toxicol Chem* 12:567–576.
- Goktepe I, Plhak LC. 2002. Comparative toxicity of two azadirachtin-based neem pesticides to *Daphnia pulex*. *Environ Toxicol Chem* 21:31–36.
- Gough M. 1991. Human exposures from dioxin in soil—a meeting report. *J Toxicol Environ Health* 32:205–245.
- Gouin T, Mackay D, Jones KC, Harner T, Meijer SN. 2004. Evidence for the “grasshopper” effect and fractionation during long-range atmospheric transport of organic contaminants. *Environ Pollut* 128:139–148.
- Greco WR, Bravo G, Parsons JC. 1995. The search for synergy: a critical review from a response surface perspective. *Pharmacol Rev* 47:331–385.
- Greco WR, Park HS, Rustum YM. 1990. An application of a new approach for the quantitation of drug synergism to the combination of cis-diamminedichloroplatinum and 1-b-D-arabinosefuranosylcytosine. *Cancer Res* 50:5318–5327.
- Greco, WR, Unkelbach HD, Pösch G, Sühnel J, Kundi M, Boedeker W. 1992. Consensus on concepts and terminology for interaction assessment: the Saarselskä agreement. *Arch Complex Environ Stud* 4:65–69.

- Grimme LH, Altenburger R, Boedeker W, Faust M. 1994. Kombinationswirkungen von Schadstoffen—Toxizität binärer Kombinationen von Pestiziden und Tensiden im Algenbiotest. Forschungsbericht Nr. 94-10207205 im Auftrag des Umweltbundesamtes.
- Grote M, Brack W, Walter HA, Altenburger R. 2005. Confirmation of cause-effect relationships using effect-directed analysis for complex environmental samples. *Environ Toxicol Chem* 24:1420–1427.
- Groten JP, Feron VJ, Suhnel J. 2001. Toxicology of simple and complex mixtures. *Trends Pharm Sci* 22:316–322.
- Groten JP, Schoen ED, Van Bladeren PJ, Kuper CF, Van Zorge JA, Feron VJ. 1997. Subacute toxicity of a mixture of nine chemicals in rats: detecting interactive effects with a fractionated two-level factorial design. *Fund Appl Toxicol* 36:15–29.
- Guha S, Jaffe PR, Peters CA. 1998. Bioavailability of mixtures of PAHs partitioned into the micellar phase of a nonionic surfactant. *Environ Sci Technol* 32:2317–2324.
- Guha S, Peters CA, Jaffe PR. 1999. Multisubstrate biodegradation kinetics of naphthalene, phenanthrene, and pyrene mixtures. *Biotechn Bioeng* 65:491–499.
- Gust KA, Fleegeer JW. 2005. Exposure-related effects on Cd bioaccumulation explain toxicity of Cd-phenanthrene mixtures in *Hyalella azteca*. *Environ Toxicol Chem* 24:2918–2926.
- Haanstra L, Doelman P, Oude Voshaar JH. 1985. The use of sigmoidal dose response curves in soil ecotoxicological research. *Plant Soil* 84:293–297.
- Haas CN, Cidambi K, Kersten S, Wright K. 1996. Quantitative description of mixture toxicity: effect of level of response on interactions. *Environ Toxicol Chem* 15:1429–1437.
- Haas CN, Kersten SP, Wright K, Frank MJ, Cidambi K. 1997. Generalization of independent response model for toxic mixtures. *Chemosphere* 34:699–710.
- Hack CE. 2006. Bayesian analysis of physiologically based toxicokinetic and toxicodynamic models. *Toxicology* 221:241–248.
- Haddad S, Charest-Tardif G, Krishnan K. 2000c. Physiologically based modeling of the maximal effect of metabolic interactions on the kinetics of components of complex chemical mixtures. *J Toxicol Environ Health A* 61:209–223.
- Haddad S, Charest-Tardif G, Tardif R, Krishnan K. 2000b. Validation of a physiological modeling framework for simulating the toxicokinetics of chemicals in mixtures. *Toxicol Appl Pharmacol* 167:199–209.
- Haddad S, Krishnan K. 1998. Physiological modeling of toxicokinetic interactions: implications for mixture risk assessment. *Environ Health Perspect* 106(Suppl 6):1377–1384.
- Haddad S, Poulin P, Krishnan K. 2000a. Relative lipid content as the sole mechanistic determinant of the adipose tissue:blood partition coefficients of highly lipophilic organic chemicals. *Chemosphere* 40:839–843.
- Haddad S, Tardif R, Charest-Tardif G, Krishnan K. 1999. Physiological modeling of the toxicokinetic interactions in a quaternary mixture of aromatic hydrocarbons. *Toxicol Appl Pharmacol* 161:249–257.
- Haddad S, Withey J, Laparé S, Law FCP, Tardif R, Krishnan K. 1998. Physiologically-based pharmacokinetic modeling of pyrene in the rat. *Environ Toxicol Pharmacol* 5:245–255.
- Hakooz N, Ito K, Rawden H, Gill H, Lemmers L, Boobis AR, Edwards RJ, Carlile DJ, Lake BG, Houston JB. 2006. Determination of a human hepatic microsomal scaling factor for predicting *in vivo* drug clearance. *Pharm Res* 23:533–539.
- Harbers JV, Huijbregts MAJ, Posthuma L, Van de Meent D. 2006. Estimating the impact of high-production-volume chemicals on remote ecosystems by toxic pressure calculation. *Environ Sci Technol* 40:1573–1580.
- Hass U, Scholze M, Christiansen S, Dalggaard M, Vinggaard AM, Axelstad M, Metzendorff SB, Kortenkamp A. 2007. Combined exposure to anti-androgens exacerbates disruption of sexual differentiation in the rat. *Environ Health Perspect* 115(Suppl 1):122–128.
- Hassanin A, Johnston AE, Thomas GO, Jones KC. 2005. Time trends of atmospheric PBDEs inferred from archived UK herbage. *Environ Sci Technol* 39:2436–2441.

- Hatzinger PB, Alexander M. 1995. Effect of aging of chemicals in soil on their biodegradability and extractability. *Environ Sci Technol* 29:537–545.
- Hauser R, Chen Z, Pothier L, Ryan L, Altshul L. 2003a. The relationship between human semen parameters and environmental exposure to polychlorinated biphenyls and p,p'-DDE. *Environ Health Perspect* 111:1505–1511.
- Hauser R, Singh NP, Chen Z, Pothier L, Altshul L. 2003b. Lack of an association between environmental exposure to polychlorinated biphenyls and p,p'-DDE and DNA damage in human sperm measured using the neutral comet assay. *Hum Reprod* 18:2525–2533.
- Hawkins CP, Norris RH, Hogue JN, Feminella JW. 2000. Development and evaluation of predictive models for measuring the biological integrity of streams. *Ecol Appl* 10:1456–1477.
- Haws NW, Ball WP, Bouwer EJ. 2006. Modeling and interpreting bioavailability of organic contaminant mixtures in subsurface environments. *J Contam Hydrol* 82:255–292.
- Hearl FJ. 2005. Occupational exposure to chemical mixtures. Presented at the First International Conference on Environmental Exposure and Health, Atlanta (GA).
- Hela DG, Konstantinou IK, Sakellarides TM, Lambropoulou DA, Akriotis T, Albanis TA. 2006. Persistent organochlorine contaminants in liver and fat of birds of prey from Greece. *Arch Environ Contam Toxicol* 50:603–613.
- Hemond HF, Solo-Gabriele HM. 2004. Children's exposure to arsenic from CCA-treated wooden decks and playground structures. *Risk Anal* 24:51–64.
- Hendriks AJ, Heikens A. 2001. The power of size. 2. Rate constants and equilibrium ratios for accumulation of inorganic substances related to species weight. *Environ Toxicol Chem* 20:1421–1437.
- Hendriks AJ, Van der Linde A, Cornelissen G, Sijm D. 2001. The power of size. 1. Rate constants and equilibrium ratios for accumulation of organic substances related to octanol-water partition ratio and species weight. *Environ Toxicol Chem* 20:1399–1420.
- Henning-de Jong I, Van Zelm R, Huijbregts MAJ, De Zwart D, Van der Linden TMA, Wintersen A, Posthuma L, Van de Meent D. 2008. Ranking of agricultural pesticides in the Rhine-Meuse-Scheldt Basin based on toxic pressure in marine ecosystems. *Environ Toxicol Chem* 27:737–745.
- Hermens J, Busser F, Leeuwangh P, Musch A. 1985a. Quantitative structure–activity relationships and mixture toxicity of organic chemicals in *Photobacterium phosphoreum*: the Microtox test. *Ecotoxicol Environ Safety* 9:17–25.
- Hermens J, Busser F, Leeuwangh P, Musch A. 1985c. Quantitative correlation studies between acute lethal toxicity of 15 organic halides to the guppy (*Poecilia reticulata*) and chemical reactivity towards 4-nitrobenzylpyridine. *Toxicol Environ Chem* 9:219–223.
- Hermens J, Canton H, Janssen P, De Jong R. 1984. Quantitative structure–activity relationships and toxicity studies of mixtures of chemicals with anaesthetic potency: acute lethal and sublethal toxicity to *Daphnia magna*. *Aquat Toxicol* 5:143–154.
- Hermens J, Leeuwangh P, Musch A. 1985b. Joint toxicity of mixtures of groups of organic aquatic pollutants to the guppy (*Poecilia reticulata*). *Ecotoxicol Environ Safety* 9:321–326.
- Hertwich EG, McKone TE, Pease WS. 1999. Parameter uncertainty and variability in evaluative fate and exposure models. *Risk Anal* 19:1193–1204.
- Hertzberg RC, MacDonell MM. 2002. Synergy and other ineffective mixture risk definitions. *Sci Total Environ* 288:31–42.
- Hertzberg RC, Teuschler LK. 2002. Evaluating quantitative formulas for dose–response assessment of chemical mixtures. *Environ Health Perspect* 110:965–970.
- Heugens EHW. 2003. Predicting effects of multiple stressors on aquatic biota. PhD thesis, University of Amsterdam (NL).

- Heugens EHW, Hendriks AJ, Dekker T, van Straalen NM, Admiraal W. 2001. A review of the effects of multiple stressors on aquatic organisms and analysis of uncertainty factors for use in risk assessment. *Crit Rev Toxicol* 31:247–284.
- Hewlett PS, Plackett RL. 1959. A unified theory for quantal responses to mixtures of drugs: non-interactive action. *Biometrics* 15:591–610.
- Hickie BE, Mackay D, De Koning J. 1999. Lifetime pharmacokinetic model for hydrophobic contaminants in marine mammals. *Environ Toxicol Chem* 18:2622–2633.
- Hill AV. 1910. The possible effects of the aggregation of the molecules of haemoglobin on its dissociation curves. *J Physiol* 40:iv–vii.
- Hodgson E, Rose RL. 2005. Human metabolism and metabolic interactions of deployment-related chemicals. *Drug Metab Rev* 37:1–39.
- Holford NH, Sheiner LB. 1981. Pharmacokinetic and pharmacodynamic modeling *in vivo*. *Crit Rev Bioeng* 5:273–322.
- Hope BK. 2001. A case study comparing static and spatially explicit ecological exposure analysis methods. *Risk Anal* 21:1001–1010.
- Hope BK. 2005. Performing spatially and temporally explicit ecological exposure assessments involving multiple stressors. *Human Ecol Risk Assess* 11:539–565.
- Hopkin SP, Hardisty GN, Martin MH. 1986. The woodlouse *Porcellio scaber* as a “biological indicator” of zinc, cadmium, lead and copper pollution. *Environ Pollut* 11B:271–290.
- Hopkin SP, Jones DT, Dietrich D. 1993. The terrestrial isopod *Porcellio scaber* as a monitor of the bioavailability of metals: towards a global “woodlouse watch” scheme. *Sci Total Environ Suppl*:357–365.
- Houba VGJ, Lexmond TM, Novozamsky I, Van der Lee JJ. 1996. State of the art and future developments in soil analysis for bioavailability assessment. *Sci Total Environ* 178:21–28.
- Houston JB, Galetin A. 2003. Progress towards prediction of human pharmacokinetic parameters from *in vitro* technologies. *Drug Metab Rev* 35:393–415.
- Houston JB, Kenworthy KE. 2000. *In vitro-in vivo* scaling of CYP kinetic data not consistent with the classical Michaelis-Menten model. *Drug Metab Dispos* 28:246–254.
- Houtman CJ, Cenijn PH, Hamers T, Lamoree MH, Legler J, Murk AJ, Brouwer A. 2004. Toxicological profiling of sediments using *in vitro* bioassays, with emphasis on endocrine disruption. *Environ Toxicol Chem* 23:32–40.
- Huang XH, Qiu FR, Xie HT, Li J. 2005. Pharmacokinetic and pharmacodynamic interaction between irbesartan and hydrochlorothiazide in renal hypertensive dogs. *J Cardiovasc Pharmacol* 46:863–869.
- Humphrey HEB. 1983. Evaluation of humans exposed to waterborne chemicals in the Great Lakes. Final report to the Environmental Protection Agency. Lansing (MI): Department of Public Health.
- Hunter BA, Johnson MS, Thompson DJ. 1987. Ecotoxicology of copper and cadmium in a contaminated grassland ecosystem. 3. Small mammals. *J Appl Ecol* 24:601–614.
- Hunter BA, Johnson MS, Thompson DJ. 1989. Ecotoxicology of copper and cadmium in a contaminated grassland ecosystem. 4. Tissue distribution and age accumulation in small mammals. *J Appl Ecol* 29:89–99.
- Hutcheson MS, Pedersen D, Anastasa ND, Fitzgerald J, Silverman D. 1996. Beyond TPH: health-based evaluation of petroleum hydrocarbon exposures. *Reg Toxicol Pharmacol* 24:85–101.
- Ibarluzea JJ, Fernandez MF, Santa-Marina L, Olea-Serrano MF, Rivas AM, Aurrekoetxea JJ, Exposito J, Lorenzo M, Torne P, Villalobos M, Pedraza V, Sasco AJ, Olea N. 2004. Breast cancer risk and the combined effect of environmental oestrogens. *Cancer Causes Control* 15:591–600.
- IJC. 1983. An inventory of chemical substances identified in the Great Lakes ecosystem. Vols. 1–6. Windsor, Ontario (CA): International Joint Commission.

- Irving EC, Baird DJ, Culp JM. 2003. Ecotoxicological responses of the mayfly *Baetis tricaudatus* to dietary and waterborne cadmium: implications for toxicity testing. *Environ Toxicol Chem* 22:1058–1064.
- Isaacs KK, Evans MV, Harris TR. 2004. Visualization-based analysis for a mixed-inhibition binary PBPK model: determination of inhibition mechanism. *J Pharmacokinet Pharmacodyn* 31:215–242.
- Ishigam M, Uchiyama M, Kondo T, Iwabuchi H, Inoue S, Takasaki W, Ikeda T, Komai T, Ito K, Sugiyama Y. 2001. Inhibition of *in vitro* metabolism of simvastatin by itraconazole in humans and prediction of *in vivo* drug-drug interactions. *Pharm Res* 18:622–631.
- Ito K, Houston JB. 2004. Comparison of the use of liver models for predicting drug clearance using *in vitro* kinetic data from hepatic microsomes and isolated hepatocytes. *Pharm Res* 21:785–792.
- Ito K, Houston JB. 2005. Prediction of human drug clearance from *in vitro* and preclinical data using physiologically based and empirical approaches. *Pharm Res* 22:103–112.
- Ito N, Imaida K, Hasegawa R, Tsuda H. 1989a. Rapid bioassay methods for carcinogens and modifiers of hepatocarcinogenesis. *Crit Rev Toxicol* 19:385–415.
- Ito N, Tatematsu M, Hasegawa R, Tsuda H. 1989b. Medium-term bioassay system for detection of carcinogens and modifiers of hepatocarcinogenesis utilizing the GST-P positive liver cell focus as an endpoint marker. *Toxicol Pathol* 17:630–641.
- IUPAC 1997. IUPAC compendium of chemical terminology. 2nd ed. Triangle Park (NC): International Union of Pure and Applied Chemistry.
- Jacobson JL, Jacobson SW. 1996. Intellectual impairment in children exposed to polychlorinated biphenyls *in utero*. *N Engl J Med* 335:783–789.
- Jager T, Crommentuijn T, Van Gestel CAM, Kooijman SALM. 2004. Simultaneous modeling of multiple endpoints in life-cycle toxicity tests. *Environ Sci Technol* 38:2894–2900.
- Jager T, Crommentuijn T, Van Gestel CAM, Kooijman SALM. 2007. Chronic exposure to chlorpyrifos reveals two modes of action in the springtail *Folsomia candida*. *Environ Pollut* 145:452–458.
- Jager T, Fleuren RHLJ, Hogendoorn EA, De Korte G. 2003. Elucidating the routes of exposure for organic chemicals in the earthworm, *Eisenia andrei* (Oligochaeta). *Environ Sci Technol* 37:3399–3404.
- Jager T, Heugens EHW, Kooijman SALM. 2006. Making sense of ecotoxicological test results: towards application of process-based models. *Ecotoxicology* 15:305–314.
- Jager T, Kooijman SALM. 2005. Modeling receptor kinetics in the analysis of survival data for organophosphorus pesticides. *Environ Sci Technol* 39:8307–8314.
- Jager T, Kooijman SALM. 2009. A biology-based approach for quantitative structure–activity relationships (QSARs) in ecotoxicity. *Ecotoxicology* 18:187–196.
- Jager T, Posthuma L, De Zwart D, Van de Meent D. 2007. Novel view on predicting acute toxicity: decomposing toxicity data in species vulnerability and chemical potency. *Ecotoxicol Environ Safety* 67:311–322.
- Jager T, Vandenbrouck T, Baas J, De Coen WM, Kooijman SALM. 2010. A biology-based approach for mixture toxicity of multiple endpoints over the life cycle. *Ecotoxicology* (doi:10.1007/s10646-009-0417-z).
- Jager T, Van der Wal L, Fleuren RHLJ, Barendregt A, Hermens JLM. 2005. Bioaccumulation of organic chemicals in contaminated soils: evaluation of bioassays with earthworms. *Environ Sci Technol* 39:293–298.
- Jang JY, Droz PO, Kim S. 2001. Biological monitoring of workers exposed to ethylbenzene and co-exposed to xylene. *Int Arch Occup Environ Health* 74:31–37.
- Janssen MPM, Bruins A, De Vries TH, Van Straalen NM. 1991. Comparison of cadmium kinetics in four soil arthropod species. *Arch Environ Contam Toxicol* 20:305–312.

- Janssen RPT, Posthuma L, Baerselman R, Den Hollander HA, Van Veen RPM, Peijnenburg WJGM. 1997. Equilibrium partitioning of heavy metals in Dutch field soils. II. Prediction of metal accumulation in earthworms. *Environ Toxicol Chem* 16:2479–2488.
- Janssens de Bisthoven L, Gerhardt A, Soares AMVM. 2004. Effects of acid mine drainage on *Chironomus* spp. (Diptera) in laboratory and *in situ* bioassays with the multispecies freshwater biomonitor. *Environ Toxicol Chem* 23:1123–1128.
- Janssens de Bisthoven L, Gerhardt A, Soares AMVM. 2005. Chironomidae as bioindicators of acid mine drainage stress. *Hydrobiologia* 532:181–191.
- Janssens de Bisthoven L, Gerhardt A, Soares AMVM. 2006. Behavioural changes and acute toxicity of the freshwater shrimp *Atyaephyra desmaresti* Millet (Decapoda: Natantia) from exposure to acid mine drainage. *Ecotoxicology* 15:215–227.
- Jaspers V, Covaci A, Maervoet J, Dauwe T, Voorspoels S, Schepens P, Eens M. 2005. Brominated flame retardants and organochlorine pollutants in eggs of little owls (*Athene noctua*) from Belgium. *Environ Pollut* 136:81–88.
- Jensen J, Mesman M. (Eds). 2006. Ecological risk assessment of contaminated land. Decision support for site specific investigations. Report 711701047. Bilthoven (NL): National Institute for Public Health and the Environment (RIVM).
- Johnston G, Walker CH, Dawson A. 1994. Interactive effects of prochloraz and malathion in pigeon, starling and hybrid red-legged partridge. *Environ Toxicol Chem* 13:115–120.
- Jones DT, Hopkin SP. 1991. Biological monitoring of metal pollution in terrestrial ecosystems. In: Ravera O, editor, *Terrestrial and aquatic ecosystems: perturbation and recovery*. Chichester (UK): Ellis Horwood. p 148–152.
- Jonker D, Woutersen RA, Feron VJ. 1996. Toxicity of mixtures of nephrotoxics with similar or dissimilar mode of action. *Food Chem Toxicol* 34:1075–1082.
- Jonker D, Woutersen RA, van Bladeren PJ, Til HP, Feron VJ. 1990. 4-Week oral toxicity study of a combination of eight chemicals in rats: comparison with the toxicity of the individual compounds. *Food Chem Toxicol* 28:623–631.
- Jonker D, Woutersen RA, van Bladeren PJ, Til HP, Feron VJ. 1993. Subacute (4-wk) oral toxicity of a combination of four nephrotoxins in rats: comparison with the toxicity of the individual compounds. *Food Chem Toxicol* 31:125–136.
- Jonker DM, Vermeij DA, Edelbroek PM, Voskuyl RA, Piotrovsky VK, Danhof M. 2003. Pharmacodynamic analysis of the interaction between tiagabine and midazolam with an allosteric model that incorporates signal transduction. *Epilepsia* 44:329–338.
- Jonker MJ. 2003. Joint toxic effects on *Caenorhabditis elegans*: on the analysis and interpretation of mixture toxicity data. PhD thesis, Wageningen University, Wageningen (NL).
- Jonker MJ, Svendsen C, Bedaux JJM, Bongers M, Kammenga JE. 2005. Significance testing of synergistic/antagonistic, dose level-dependent, or dose ratio-dependent effects in mixture dose–response analysis. *Environ Toxicol Chem* 24:2701–2713.
- Jonker MJ, Sweijen RAJC, Kammenga JE. 2004. Toxicity of simple mixtures to the nematode *Caenorhabditis elegans* in relation to soil sorption. *Environ Toxicol Chem* 23:480–488.
- Jonkers RE, Koopmans RP, Portier EJ, van Bostel CJ. 1991. Debrisoquine phenotype and the pharmacokinetics and beta-2 receptor pharmacodynamics of metoprolol and its enantiomers. *J Pharmacol Exp Ther* 256:959–966.
- Jonsson F, Johanson G. 2003. The Bayesian population approach to physiological toxicokinetic-toxicodynamic models—an example using the MCSim software. *Toxicol Lett* 138:143–150.
- Jouraeva VA, Johnson DL, Hassett JP, Nowak DJ. 2002. Differences in accumulation of PAHs and metals on the leaves of *Tilia × euchlora* and *Pyrus calleryana*. *Environ Pollut* 120:331–338.
- Kaag NHBM, Scholten MCT, Van Straalen NM. 1998. Factors affecting PAH residues in the lugworm *Arenicola marina*, a sediment feeding polychaete. *J Sea Res* 40:251–261.

- Kammenga J, Dallinger R, Donker MH, Köhler HR, Simonsen V, Triebskorn R, Weeks JM. 2000. Biomarkers in terrestrial invertebrates: potential and limitations for ecotoxicological soil risk assessment. *Rev Environ Contam Toxicol* 164:93–147.
- Kanamitsu S, Ito K, Green CE, Tyson CA, Shimada N, Sugiyama Y. 2000a. Prediction of *in vivo* interaction between triazolam and erythromycin based on *in vitro* studies using human liver microsomes and recombinant human CYP3A4. *Pharm Res* 17:419–426.
- Kanamitsu SI, Ito K, Okuda H, Ogura K, Watabe T, Muro K, Sugiyama Y. 2000b. Prediction of *in vivo* drug-drug interactions based on mechanism-based inhibition from *in vitro* data: inhibition of 5-fluorouracil metabolism by (E)-5-(2-bromovinyl)uracil. *Drug Metab Dispos* 28:467–474.
- Kapo KE, Burton GA Jr. 2006. A geographic information systems-based, weights of evidence approach for diagnosing aquatic ecosystem impairment. *Environ Toxicol Chem* 25:2237–2249.
- Kavlock RJ, Daston GP, De Rosa C, Fenner-Crisp P, Gray LE, Kaattari S, Lucier G, Luster M, Mae MJ, Maczka C, Miller R, Moore J, Rolland R, Scott G, Sheehan DM, Sinks T, Tilson HA. 1996. Research needs for the risk assessment of health and environmental effects of endocrine disruptors: a report of the USEPA-sponsored workshop. *Environ Health Perspect* 104(Suppl 4):715–740.
- Kedderis GL, Mason AD, Niang LL, Wilkes CR. 2006. Exposures and internal doses of trihalomethanes in humans: multi-route contributions from drinking water [Final]. EPA/600/R-06/087. Washington (DC): US Environmental Protection Agency.
- Kelsey JW, Alexander M. 1997. Declining bioavailability and inappropriate estimation of risk of persistent compounds. *Environ Toxicol Chem* 16:582–585.
- Kenntner N, Krone O, Altenkamp R, Tataruch F. 2003a. Environmental contaminants in liver and kidney of free-ranging northern goshawks (*Accipiter gentilis*) from three regions of Germany. *Arch Environ Contam Toxicol* 45:128–135.
- Kenntner N, Krone O, Oehme G, Heidecke D, Tataruch F. 2003b. Organochlorine contaminants in body tissue of free-ranging white-tailed eagles from northern regions of Germany. *Environ Toxicol Chem* 22:1457–1464.
- Keys DA, Schultz IR, Mahle DA, Fisher JW. 2004. A quantitative description of suicide inhibition of dichloroacetic acid in rats and mice. *Toxicol Sci* 82:381–393.
- Kim SK, Oh JR, Shim WJ, Lee DH, Yim UH, Hong SH, Shin YB, Lee DS. 2002. Geographical distribution and accumulation features of organochlorine residues in bivalves from coastal areas of South Korea. *Mar Pollut Bull* 45:268–279.
- King DJ, Lyne RL, Girling A, Peterson DR, Stephenson R, Short D. 1996. Environmental risk assessment of petroleum substances: the hydrocarbon block method. Report 96/52. Brussels: Concawe, Petroleum Products Ecology Group.
- King JK, Harmon SM, Fu TT, Gladden JB. 2002. Mercury removal, methylmercury formation, and sulfate-reducing bacteria profiles in wetland mesocosms. *Chemosphere* 46:859–870.
- Klaassen CD. 1996. Casarett and Doull's toxicology: the basic science of poisons. New York: McGraw-Hill.
- Klein MT, Hou G, Quann R, Wei W, Liao KH, Yang RSH, Campaign JA, Mazurek M, Broadbelt LJ. 2002. BioMOL: a computer-assisted biological modeling tool for complex chemical mixtures and biological processes at the molecular level. *Environ Health Perspect* 110(Suppl 6):1025–1029.
- Kodell RL, Chen JJ. 1994 Reducing conservatism in risk estimation for mixtures of carcinogens. *Risk Anal* 14:327–332.
- Kodell RL, Pounds JG. 1991. Assessing the toxicity of mixtures of chemicals. In: Krewski D, Franklin C, editors, *Statistics in toxicology*. New York: Gordon and Breach, p 559–591.

- Könemann H. 1980. Structure–activity relationships and additivity in fish toxicities of environmental pollutants. *Ecotoxicol Environ Safety* 4:415–421.
- Könemann H. 1981. Fish toxicity tests with mixtures of more than two chemicals: a proposal for a quantitative approach and experimental results. *Toxicology* 19:229–238.
- Kooijman SALM. 1981. Parametric analyses of mortality rates in bioassays. *Water Res* 15:107–119.
- Kooijman SALM. 1996. An alternative for NOEC exists, but the standard model has to be abandoned first. *Oikos* 75:310–316.
- Kooijman SALM. 2000. *Dynamic energy and mass budgets in biological systems*. Cambridge (UK): Cambridge University Press.
- Kooijman SALM. 2001. Quantitative aspects of metabolic organization: a discussion of concepts. *Phil Trans R Soc London B* 356:331–349.
- Kooijman SALM, Bedaux JJM. 1996. Analysis of toxicity tests on *Daphnia* survival and reproduction. *Water Res* 30:1711–1723.
- Kooistra L, Huijbregts MAJ, Ragas AMJ, Wehrens R, Leuven RSEW. 2005. Spatial variability and uncertainty in ecological risk assessment: a case study on the potential risk of cadmium for the little owl in a Dutch river flood plain. *Environ Sci Technol* 39:2177–2187.
- Koppe JG. 1995. Nutrition and breast-feeding. *Eur J Obstet Gynecol Reprod* 61:73–78.
- Kortenkamp A. 2007. Ten years of mixing cocktails—a review of combination effects of endocrine disrupting chemicals. *Environ Health Perspect* 115(Suppl 1):98–105.
- Kortenkamp A, Altenburger R. 1998. Synergisms with mixtures of xenoestrogens: a reevaluation using the method of isoboles. *Sci Total Environ* 221:59–73.
- Kortenkamp A, Faust M, Scholze M, Backhaus T. 2007. Low-level exposure to multiple chemicals: reason for human health concerns? *Environ Health Perspect* 115(Suppl 1):106–114.
- Kosian PA, Makynen EA, Monson PD, Mount DR, Spacie A, Mekenyan OG, Ankley GT. 1998. Application of toxicity-based fractionation techniques and structure–activity relationship models for the identification of phototoxic polycyclic aromatic hydrocarbons in sediment pore water. *Environ Toxicol Chem* 17:1021–1033.
- Kramarz P. 1999a. The dynamics of accumulation and decontamination of cadmium and zinc in carnivorous invertebrates. 2. The centipede *Lithobius mutabilis* Koch. *Bull Environ Contam Toxicol* 63:538–545.
- Kramarz P. 1999b. The dynamics of accumulation and decontamination of cadmium and zinc in carnivorous invertebrates. 1. The ground beetle, *Poecilus cupreus* L. *Bull Environ Contam Toxicol* 63:531–537.
- Krishnan K, Andersen ME, Clewell HJ, Yang RSH. 1994. Physiologically based pharmacokinetic modeling of chemical mixtures. In: Yang RSH, editor, *Toxicology of chemical mixtures: case studies, mechanisms and novel approaches*. New York: Academic Press. p 399–437.
- Krishnan K, Andersen ME, Hayes AW. 2001. *Physiologically based pharmacokinetic modeling in toxicology*. 4th ed. Philadelphia (PA): Taylor and Francis. p 193–241.
- Krishnan K, Brodeur J. 1991. Toxicological consequences of combined exposure to environmental pollutants. *Arch Complex Environ Studies* 3:1–106.
- Krishnan K, Haddad S, Beliveau M, Tardif R. 2002. Physiological modeling and extrapolation of pharmacokinetic interactions from binary to more complex chemical mixtures. *Environ Health Perspect* 110(Suppl 6):989–994.
- Krishnan K, Pelekis M. 1995. Hematotoxic interactions: occurrence, mechanisms and predictability. *Toxicology* 105:355–364.
- Krull IS, Mills K, Hoffman G, Fine DH. 1980. The analysis of N-nitrosoatrazine and N-nitrosocarbaryl in whole mice. *J Anal Toxicol* 4:260–262.

- Küster E, Dorusch F, Vogt C, Weiss H, Altenburger R. 2004. On line biomonitors used as a tool for toxicity reduction evaluation of *in situ* groundwater remediation techniques. *Biosensors Bioelectronics* 19:1711–1722.
- Kwon CS, Penner D. 1995. The interaction of insecticides with herbicide activity. *Weed Technol* 9:119–124.
- Landrum PF, Steevens JA, Gossiaux DC, McElroy M, Robinson S, Begnoche L, Chernyak S, Hickey J. 2004. Time-dependent lethal body residues for the toxicity of pentachlorobenzene to *Hyalella azteca*. *Environ Toxicol Chem* 23:1335–1343.
- Lau CE, Wang Y, Falk JL. 1997. Differential reinforcement of low rate performance, pharmacokinetics and pharmacokinetic-pharmacodynamic modeling: independent interaction of alprazolam and caffeine. *J Pharmacol Exp Ther* 281:1013–1029.
- Law FC, Abedini S, Kennedy CJ. 1991. A biologically based toxicokinetic model for pyrene in rainbow trout. *Toxicol Appl Pharmacol* 110:390–402.
- Leavens TL, Bond JA. 1996. Pharmacokinetic model describing the disposition of butadiene and styrene in mice. *Toxicology* 113:310–313.
- Lee JH, Landrum PF. 2006a. Application of multi-component damage assessment model (MDAM) for the toxicity of metabolized PAH in *Hyalella azteca*. *Environ Sci Technol* 40:1350–1357.
- Lee JH, Landrum PF. 2006b. Development of a multi-component damage assessment model (MDAM) for time-dependent mixture toxicity with toxicokinetic interactions. *Environ Sci Technol* 40:1341–1349.
- Lee JH, Landrum PF, Koh CH. 2002a. Prediction of time-dependent PAH toxicity in *Hyalella azteca* using a damage assessment model. *Environ Sci Technol* 36:3131–3138.
- Lee JH, Landrum PF, Koh CH. 2002b. Toxicokinetics and time-dependent PAH toxicity in the amphipod *Hyalella azteca*. *Environ Sci Technol* 36:3124–3130.
- Lee JS, Lee JH. 2005. Influence of acid volatile sulfides and simultaneously extracted metals on the bioavailability and toxicity of a mixture of sediment-associated Cd, Ni, and Zn to polychaetes *Neanthes arenaceodentata*. *Sci Total Environ* 338:229–241.
- Legierse KCHM, Verhaar HJM, Vaes WHJ, De Bruijn JHM, Hermens JLM. 1999. Analysis of the time-dependent acute aquatic toxicity of organophosphorus pesticides: the critical target occupation model. *Environ Sci Technol* 33:917–925.
- Lepper, P. 2005. Manual on the methodological framework to derive environmental quality standards for priority substances in accordance with Article 16 of the Water Framework Directive (2000/60/EC). Schmallingenberg (DE): Fraunhofer-Institute, Molecular Biology and Applied Ecology.
- Leslie HA, Hermens JLM, Kraak MHS. 2004. Baseline toxicity of a chlorobenzene mixture and total body residues measured and estimated with solid-phase microextraction. *Environ Toxicol Chem* 23:2017–2021.
- Levsen K, Preiss A, Spraul M. 2003. Structure elucidation of unknown pollutants of environmental samples by coupling HPLC to NMR and MS. In: Namiesnik J, Chrzanowski W, Zmijewska P, editors, New horizons and challenges in environmental analysis and monitoring, Workshop, Gdansk (PL), August 18–29. p 150–180.
- Lewtas J. 1985. Development of a comparative potency method for cancer risk assessment of complex mixtures using short-term *in vivo* and *in vitro* bioassays. *Toxicol Ind Health* 1:193–203.
- Lewtas J. 1988. Genotoxicity of complex mixtures: strategies for the identification and comparative assessment of airborne mutagens and carcinogens from combustion sources. *Fund Appl Toxicol* 10:571–589.
- Liao KH. 2004. Development and validation of a hybrid reaction network/physiologically based pharmacokinetic model of benzo[a]pyrene and its metabolites. PhD dissertation, Department of Chemical and Biological Engineering, Colorado State University, Fort Collins (CO).

- Liao KH, Dobrev I, Dennison JE, Andersen ME, Reisfeld B, Reardon KF, Campain JA, Wei W, Klein MT, Quann RJ, Yang RSH. 2002. Application of biologically based computer modeling to simple or complex mixtures. *Environ Health Perspect* 110(Suppl 6):957–963.
- Lichtenstein EP, Liang TT, Anderegge BN. 1973. Synergism of insecticides by herbicides. *Science* 181:847–849.
- Linkov I, Burmistrov D, Cura J, Bridges TS. 2002. Risk-based management of contaminated sediments: consideration of spatial and temporal patterns in exposure modeling. *Environ Sci Technol* 36:238–246.
- Lock K, Janssen CR. 2001. Zinc and cadmium body burdens in terrestrial oligochaetes: use and significance in environmental risk assessment. *Environ Toxicol Chem* 20:2067–2072.
- Lock K, Janssen CR. 2003. Influence of ageing on zinc bioavailability in soils. *Environ Pollut* 126:371–374.
- Loewe S, Muischneck H. 1926. Effect of combinations: mathematical basis of problem. *Arch Exp Pathol Pharmacol* 114:313–326.
- Lohitnavy M, Lu Y, Lohitnavy O, Chubb LS, Hirono S, Yang RSH. 2008. A possible role of multidrug resistance-associated protein 2 (Mrp2) in hepatic excretion of PCB126, an environmental contaminant: PBPK/PD modeling. *Toxicol Sci* 104:27–39.
- Loonen H, Muir DCG, Parsons JR, Govers HAJ. 1997. Bioaccumulation of polychlorinated dibenzo-p-dioxins in sediments by oligochaetes: influence of exposure pathway and contact time. *Environ Toxicol Chem* 16:1518–1525.
- Loos M, Schipper AM, Schlink U, Strebel K, Ragas AMJ. 2010. Receptor-oriented approaches in wildlife and human exposure modelling: a comparative study. *Environ Model Software* 25:369–382.
- Lorber M, Cleverly D, Schaum J, Phillips L, Schweer G, Leighton T. 1994. Development and validation of an air-to-beef food chain model for dioxin-like compounds. *Sci Total Environ* 156:39–65.
- Lorber M, Saunders P, Ferrario J, Leese W, Winters D, Cleverly D, Schaum J, Deyrup C, Ellis R, Walcott J, Dupuy A, Byrne C, McDaniel D. 1997. A statistical survey of dioxin-like compounds in United States pork fat. *Organohalogen Compounds* 32:80–86.
- Lorber MN, Winters DL, Griggs J, Cook R, Baker S, Ferrario J, Byrne C, Dupuy A, Schaum J. 1998. A national survey of dioxin-like compounds in the United States milk supply. *Organohalogen Compounds* 38:125–129.
- Loureiro S, Soares AMVM, Nogueira AJA. 2005. Terrestrial avoidance behaviour tests as screening tool to assess soil contamination. *Environ Pollut* 138:121–131.
- Lu Y, Lohitnavy M, Reddy M, Lohitnavy O, Eickman E, Ashley A, Gerjevic L, Xu Y, Conolly RB, Yang RSH. 2008. Quantitative analysis of liver GST-P foci promoted by a chemical mixture of hexachlorobenzene and PCB 126: implication of size-dependent cellular growth kinetics. *Arch Toxicol* 82:103–116.
- Lucier GW, Rumbaugh RC, McCoy Z, Hass R, Harvan D, Albro P. 1986. Ingestion of soil contaminated with 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) alters hepatic enzyme activities in rats. *Fund Appl Toxicol* 6:364–371.
- Luecke RH, Wosilait WD. 1979. Drug elimination interactions: analysis using a mathematical model. *J Pharmacokinet Biopharm* 7:629–641.
- Lundin F, Lloyd J, Smith E. 1969. Mortality of uranium miners in relation to radiation exposure, hard-rock mining and cigarette smoking—1950 through 1967. *Health Phys* 16:571–578.
- Luoma SN, Rainbow PS. 2005. Why is metal bioaccumulation so variable? Biodynamics as a unifying concept. *Environ Sci Technol* 39:1921–1931.
- Lutz WK, Lutz RW, Andersen ME. 2006. Dose-incidence relationships derived from superposition of distributions of individual susceptibility on mechanism-based dose responses for biological effects. *Toxicol Sci* 90:33–38.

- Lydy MJ, Linck SL. 2003. Assessing the impact of triazine herbicides on organophosphate insecticide toxicity to the earthworm *Eisenia fetida*. Arch Environ Contam Toxicol 45:343–349.
- Lyons M, Yang RSH, Mayeno AN, Reisfeld B. 2008. Computational toxicology of chloroform: reverse dosimetry using Bayesian inference, Markov chain Monte Carlo simulation, and human biomonitoring data. Environ Health Perspect 116:1040–1046.
- Mackay D, Fraser A. 2000. Bioaccumulation of persistent organic chemicals: mechanisms and models. Environ Pollut 110:375–391.
- Mackay D, Paterson S, Shiu WY. 1992a. Generic models for evaluating the regional fate of chemicals. Chemosphere 24:695–717.
- Mackay D, Puig H, McCarty LS. 1992b. An equation describing the time course and variability in uptake and toxicity of narcotic chemicals to fish. Environ Toxicol Chem 11:941–951.
- MacLeod M, Fraser AJ, Mackay D. 2002. Evaluating and expressing the propagation of uncertainty in chemical fate and bioaccumulation models. Environ Toxicol Chem 21:700–709.
- MADEP. 2002. Characterizing risks posed by petroleum contaminated sites: implementation of the MADEP VPH/EPH approach. Boston (MA): Massachusetts Department of Environmental Protection. Available from: <http://www.mass.gov/dep/cleanup/laws/policies.htm#02-411>
- MADEP. 2003. Updated petroleum hydrocarbon fraction toxicity values for the VPH/EPH/APH methodology. Boston (MA): Massachusetts Department of Environmental Protection. Available from: <http://www.mass.gov/dep/water/drinking/standards/pethydro.htm>
- Mahmood I. 2002. Prediction of clearance in humans from *in vitro* human liver microsomes and allometric scaling: a comparative study of the two approaches. Drug Metabol Drug Interact 19:49–64.
- Main KM, Kiviranta H, Virtanen HE, Sundquist E, Tuomisto JT, Tuomisto J, Vartiainen T, Skakkebaek NE, Toppari J. 2007. Flame retardants in placenta and breast milk and cryptorchidism in newborn boys. Environ Health Perspect 115:1519–1526.
- Manceau A, Tamura N, Celestre RS, MacDowell AA, Geoffroy N, Sposito G, Padmore HA. 2003. Molecular-scale speciation of Zn and Ni in soil ferromanganese nodules from loess soils of the Mississippi Basin. Environ Sci Technol 37:75–80.
- Mandema JW, Heijligers-Feijen CD, Tukker E, De Boer AG, Danhof M. 1992a. Modeling of the effect site equilibration kinetics and pharmacodynamics of racemic baclofen and its enantiomers using quantitative EEG effect measures. J Pharmacol Exp Ther 261:88–95.
- Mandema JW, Kuck MT, Danhof M. 1992b. *In vivo* modeling of the pharmacodynamic interaction between benzodiazepines which differ in intrinsic efficacy. J Pharmacol Exp Ther 261:56–61.
- Manno M, Rezzadore M, Grossi M, Sbrana C. 1996. Potentiation of occupational carbon tetrachloride toxicity by ethanol abuse. Hum Exp Toxicol 15:294–300.
- Marigomez I, Kortabitarte M, Dussart GBJ. 1998. Tissue-level biomarkers in sentinel slugs as cost-effective tools to assess metal pollution in soils. Arch Environ Contam Toxicol 34:167–176.
- Marino DJ, Clewell HJ, Gentry PR, Covington TR, Hack CE, David RM, Morgott DA. 2006. Revised assessment of cancer risk to dichloromethane. Part I. Bayesian PBPK and dose-response modeling in mice. Regul Toxicol Pharmacol 45:44–54.
- Marinussen MPJC, Van der Zee SEATM. 1996. Conceptual approach to estimating the effects of home-range size on the exposure of organisms to spatially variable soil contamination. Ecol Model 87:83–89.

- Martín-Díaz ML, Blasco J, de Canales MG, Sales D, DelValls TA. 2005a. Bioaccumulation and toxicity of dissolved heavy metals from the Guadalquivir Estuary after the Aznalcollar mining spill using *Ruditapes philippinarum*. Arch Environ Contam Toxicol 48:233–241.
- Martín-Díaz ML, Villena-Lincoln A, Bamber S, Blasco J, DelValls TA. 2005b. An integrated approach using bioaccumulation and biomarker measurements in female shore crab, *Carcinus maenas*. Chemosphere 58:615–626.
- Matscheko N, Lundstedt S, Svensson L, Harju M, Tysklind M. 2002. Accumulation and elimination of 16 polycyclic aromatic compounds in the earthworm (*Eisenia fetida*). Environ Toxicol Chem 21:1724–1729.
- Mattsson JL. 2007. Mixtures in the real world: the importance of plant self-defense toxicants, mycotoxins, and the human diet. Toxicol Appl Pharmacol 223:125–132.
- Mayeno AN, Yang RSH, Reisfeld B. 2005. Biochemical reaction network modeling: a new tool for predicting metabolism of chemical mixtures. Environ Sci Technol 39:5363–5371.
- Mayer P, Holmstrup M. 2008. Passive dosing of soil invertebrates with polycyclic aromatic hydrocarbons: limited chemical activity explains toxicity cutoff. Environ Sci Technol 42:7516–7521.
- Mayer P, Tolls J, Hermens L, Mackay D. 2003. Equilibrium sampling devices. Environ Sci Technol 37:184A–191A.
- McCarty LS, Borgert CJ. 2006. Review of the toxicity of chemical mixtures: theory, policy and regulatory practice. Reg Toxicol Pharmacol 45:119–143.
- McCarty LS, Mackay D. 1993. Enhancing ecotoxicological modelling and assessment: body residues and modes of toxic action. Environ Sci Technol 27:1719–1728.
- McCarty LS, Ozburn GW, Smith AD, Dixon DG. 1992. Toxicokinetic modeling of mixtures of organic chemicals. Environ Toxicol Chem 11:1037–1047.
- Mehendale HM. 1984. Potentiation of halomethane hepatotoxicity: chlordecone and carbon tetrachloride. Fund Appl Toxicol 4:295–308.
- Mehendale HM. 1991. Role of hepatocellular regeneration and hepatobular healing in the final outcome of liver injury: a two-stage model of toxicity. Biochem Pharmacol 42:1155–1162.
- Mehendale HM. 1994. Mechanism of the interactive amplification of halomethane hepatotoxicity and lethality by other chemicals. In: Yang RSH, editor, Toxicology of chemical mixtures: case studies, mechanisms, and novel approaches. San Diego (CA): Academic Press. p 299–334.
- Meili M, Bishop K, Bringmark L, Johansson K, Munthe J, Sverdrup H, De Vries W. 2003. Critical levels of atmospheric pollution: criteria and concepts for operational modelling of mercury in forest and lake ecosystems. Sci Total Environ 304:83–106.
- Mendoza G, Gutierrez L, Pozo-Gallardo K, Fuentes-Rios D, Montory M, Urrutia R, Barra R. 2006. Polychlorinated biphenyls (PCBs) in mussels along the Chilean Coast. Environ Sci Pollut Res 13:67–74.
- Mesman M, Rutgers M, Peijnenburg WJGM, Bogte JJ, Dirven-Van Breemen ME, De Zwart D, Posthuma L, Schouten AJ. 2003. Site-specific ecological risk assessment: the Triad approach in practice. In: Conference proceedings of CONSOIL: 8th International FKZ/TNO Conference on Contaminated Soil, Ghent (BE), May 12–16, 2003. p 649–656.
- Metzdorff SB, Dalgaard M, Christiansen S, Axelstad M, Hass U, Kiersgaard MK, Scholze M, Kortenkamp A, Vinggaard AM. 2007. Dysgenesis and histological changes of genitals and perturbations of gene expression in male rats after *in utero* exposure to antiandrogens. Toxicol Sci 98:87–98.

- Milesen BE, Chambers JE, Chen WL, Dettbarn W, Ehrich M, Eldefrawi AT, Gaylor DW, Hamernik K, Hodgson E, Karczmar AG, Padilla S, Pope CN, Richardson RJ, Saunders DR, Sheets LP, Sultatos LG, Wallace KB. 1998. Common mechanism of toxicity: a case study of organophosphorus pesticides. *Toxicol Sci* 41:8–20.
- Miners JO, Knights KM, Houston JB, Mackenzie PI. 2006. *In vitro-in vivo* correlation for drugs and other compounds eliminated by glucuronidation in humans: pitfalls and promises. *Biochem Pharmacol* 71:1531–1539.
- Minh TB, Kunisue T, Yen NTH, Watanabe M, Tanabe S, Hue ND, Qui V. 2002. Persistent organochlorine residues and their bioaccumulation profiles in resident and migratory birds from North Vietnam. *Environ Toxicol Chem* 21:2108–2118.
- Monirith I, Ueno D, Takahashi S, Nakata H, Sudaryanto A, Subramanian A, Karuppiiah S, Ismail A, Muchtar M, Zheng JS, Richardson BJ, Prudente M, Hue ND, Tana TS, Tkalin AV, Tanabe S. 2003. Asia-Pacific mussel watch: monitoring contamination of persistent organochlorine compounds in coastal waters of Asian countries. *Mar Pollut Bull* 46:281–300.
- Monosson E. 2005. Chemical mixtures: considering the evolution of toxicology and chemical assessment. *Environ Health Perspect* 113:383–390.
- Mood AM, Graybill FA, Boes DC. 1974. Introduction to the theory of statistics. 3rd ed. Auckland (NZ): McGraw-Hill Book Company.
- Moolgavkar SH, Luebeck G. 1990. Two-event model for carcinogenesis: biological, mathematical, and statistical considerations. *Risk Anal* 10:323–341.
- Moolgavkar SH, Venzon DJ. 2000. Two-event model for carcinogenesis. *Math Biosci* 47:55–77.
- Morgan JE, Morgan AJ. 1993. Seasonal changes in the tissue-metal (Cd, Zn and Pb) concentrations in two ecophysiologically dissimilar earthworm species: pollution-monitoring implications. *Environ Pollut* 82:1–7.
- Mould DR, DeFeo TM, Reece S, Milla G, Limjuco R, Crews T, Choma N, Patel IH. 1995. Simultaneous modeling of the pharmacokinetics and pharmacodynamics of midazolam and diazepam. *Clin Pharmacol Ther* 58:35–43.
- Mount DI, Anderson-Carnahan DM. 1988. Methods for aquatic toxicity identification evaluations. Phase I. Toxicity characterization procedures. EPA/600/3-88/034. Duluth (MN): Environmental Research Laboratory, Office of Research and Development, US Environmental Protection Agency.
- Mount DI, Anderson-Carnahan L. 1989. Methods for aquatic toxicity identification evaluations. Phase II toxicity identification procedures. EPA/600/3-88/035. Washington (DC): US Environmental Protection Agency.
- Mount DI, Norberg-King TJ. 1993. Methods for aquatic toxicity identification evaluations. Phase II toxicity identification evaluation procedures for samples exhibiting acute and chronic toxicity. EPA/600/R-92/081. Duluth (MN): Environmental Research Laboratory, Office of Research and Development, US Environmental Protection Agency.
- Mu X, LeBlanc GA. 2004. Synergistic interaction of endocrine-disrupting chemicals: model development using an ecdysone receptor antagonist and a hormone synthesis inhibitor. *Environ Toxicol Chem* 23:1085–1091.
- Muenchow G. 1986. Ecological use of failure time analysis. *Ecology* 67:246–250.
- Mulder C, Aldenberg T, De Zwart D, Van Wijnen HJ, Breure AM. 2005. Evaluating the impact of pollution on plant-Lepidoptera relationships. *Environmetrics* 16:357–373.
- Mulder C, Breure A. 2006. Impact of heavy metal pollution on plants and leaf-miners. *Environ Chem Lett* 4:83–86.
- Munns WRJ, Kroes R, Veith G, Suter GWI, Damstra T, Water MD. 2003a. Approaches for integrated risk assessment. *Human Ecol Risk Assess* 9:267–272.
- Munns WRJ, Suter GWI, Damstra T, Kroes R, Reiter W, Marafante E. 2003b. Integrated risk assessment—results of an international workshop. *Human Ecol Risk Assess* 9:379–386.

- Murk AJ, Leonards PEG, Bulder AS, Jonas AS, Rozemeijer MJC, Denison MS, Koeman JH, Brouwer A. 1997. The CALUX (chemical-activated luciferase expression) assay adapted and validated for measuring TCDD equivalents in blood plasma. *Environ Toxicol Chem* 16:1583–1589.
- Nagel R, Loskill R, editors. 1991. Bioaccumulation in aquatic systems. Contributions to the assessment. Weinheim (DE): VCH.
- Namdari R. 1998. A physiologically based toxicokinetic model of pyrene and its major metabolites in starry flounder, *Platichthys stellatus*. Thesis dissertation, Burnaby, British Columbia (CA): Simon Fraser University.
- Narotsky MG, Weller EA, Chinchilli VM, Kavlock RJ. 1995. Nonadditive developmental toxicity in mixtures of trichloroethylene, di(2-ethylhexyl) phthalate, and heptachlor in a 5×5×5 design. *Fund Appl Toxicol* 27:203–216.
- National Health and Environmental Effect Research. 2005. Wildlife research strategy. EPA 600/R-04/050. Research Triangle Park (NC): Office of Research and Development, US Environmental Protection Agency.
- National Oceanic and Atmospheric Administration. 2002. Contaminant trends in US National Estuarine Research Reserves. Silver Springs (MD): NOAA.
- Nesnow S. 1990. Mouse skin tumours and human lung cancer: relationships with complex environmental emissions. In: Vainio H, Sorsa M, McMichael AJ, editors, *Complex mixtures and cancer risk*. Lyon (FR): IARC Scientific Publication, p 44–54.
- Nesnow S, Mass MJ, Ross JA, Galati AJ, Lambert GR, Gennings C, Carter WH, Stoner GD. 1998. Lung tumorigenic interactions in strain A/J mice of five environmental polycyclic aromatic hydrocarbons. *Environ Health Perspect* 106:1337–1346.
- Neter N, Kutner HK, Nachtsheim CJ, Wasserman W. 1996. *Applied linear statistical models*. 4th ed. Boston: WCB/McGraw-Hill.
- Newman MC, McCloskey JT. 1996. Time-to-event analyses of ecotoxicity data: ecotoxicology 5:187–196.
- Newman MC, McCloskey JT. 2000. The individual tolerance concept is not the sole explanation for the probit dose-effect model. *Environ Toxicol Chem* 19:520–526.
- Newton I, Wyllie I. 1992. Recovery of a Sparrowhawk population in relation to declining pesticide contamination. *J Appl Ecol* 29:476–484.
- Newton I, Wyllie I, Asher A. 1991. Mortality causes in British barn owls *Tyto alba*, with a discussion of aldrin dieldrin poisoning. *Ibis* 133:162–169.
- Newton I, Wyllie I, Asher A. 1993. Long-term trends in organochlorine and mercury residues in some predatory birds in Britain. *Environ Pollut* 79:143–151.
- Nichols JW, Fitzsimmons PN, Whiteman FW. 2004a. A physiologically based toxicokinetic model for dietary uptake of hydrophobic organic compounds by fish. II. Simulation of chronic exposure scenarios. *Toxicol Sci* 77:219–229.
- Nichols JW, Fitzsimmons PN, Whiteman FW, Dawson TD, Babeu L, Juenemann J. 2004b. A physiologically based toxicokinetic model for dietary uptake of hydrophobic organic compounds by fish. I. Feeding studies with 2,2',5,5'-tetrachlorobiphenyl. *Toxicol Sci* 77:206–218.
- Nichols JW, McKim JM, Andersen ME, Gargas ML, Clewell HJ, Erickson RJ. 1990. A physiologically based toxicokinetic model for the uptake and disposition of waterborne organic chemicals in fish. *Toxicol Appl Pharmacol* 106:433–447.
- Nicholson JK, Kendall MD, Osborn, D. 1983. Cadmium and nephrotoxicity. *Nature* 304:633–635.
- [NIOSH] National Institute for Occupational Safety and Health. 1976. Criteria for a recommended standard for occupational exposure to methylene chloride. Cincinnati (OH): National Institute for Occupational Safety and Health.
- Nisbet RM, Muller EB, Lika K, Kooijman SALM. 2000. From molecules to ecosystems through dynamic energy budget models. *J Anim Ecol* 69:913–926.

- Nong A, McCarver DG, Hines RN, Krishnan K. 2006. Modeling interchild differences in pharmacokinetics on the basis of subject-specific data on physiology and hepatic CYP2E1 levels: a case study with toluene. *Toxicol Appl Pharmacol* 214:78–87.
- Norberg-King TJ, Mount DI, Amato JR, Jensen DA, Thompson JA. 1992. Toxicity identification evaluation: characterization of chronically toxic effluents, phase I. USEPA/600/6-91/005F. Duluth (MN): Environmental Research Laboratory, Office of Research and Development, US Environmental Protection Agency.
- Norum U, Lai VWM, Cullen WR. 2005. Trace element distribution during the reproductive cycle of female and male spiny and Pacific scallops, with implications for biomonitoring. *Mar Pollut Bull* 50:175–184.
- [NRC] National Research Council. 1983. Risk assessment in the federal government: managing the process. Washington (DC): Committee on the Institutional Means for Assessment of Risks to Public Health, Commission on Life Sciences, National Research Council, National Academy Press.
- [NRC] National Research Council. 1989. Mixtures. In: Drinking water and health. Vol. 9. Washington (DC): Safe Drinking Water Committee, National Research Council, National Academy of Sciences, National Academy Press.
- Obach RS. 1997. Nonspecific binding to microsomes: impact on scale-up of *in vitro* intrinsic clearance to hepatic clearance as assessed through examination of warfarin, imipramine, and propranolol. *Drug Metab Dispos* 25:1359–1369.
- Obach RS, Walsky RL, Venkatakrishnan K, Gaman EA, Houston JB, Tremaine LM. 2006. The utility of *in vitro* cytochrome P450 inhibition data in the prediction of drug-drug interactions. *J Pharmacol Exp Ther* 316:336–348.
- [OECD] Organisation for Economic Co-operation and Development. 1999. Compendium of estimation methods to quantify releases to the environment for use in pollutant release and transfer registries. Paris: Organisation for Economic Cooperation and Development.
- [OECD] Organisation for Economic Co-operation and Development. 2002a. Resource compendium of PRTR release estimation techniques. Part 1. Summary of point source techniques. ENV/JM/MONO(2002)20. Paris: Organisation for Economic Cooperation and Development, Environment Directorate, Joint Meeting of the Chemicals Committee and the Working Party on Chemicals, Pesticides and Biotechnology.
- [OECD] Organisation for Economic Co-operation and Development. 2002b. Emission scenario document on textile finishing industry. Paris: Organisation for Economic Cooperation and Development, Environment Directorate.
- [OECD] Organisation for Economic Co-operation and Development. 2002c. Emission scenario document on industrial surfactants [Draft]. Paris: Organisation for Economic Cooperation and Development, Environment Directorate.
- [OECD] Organisation for Economic Co-operation and Development. 2004. Guidance document on the use of multimedia models for estimating overall environmental persistence and long-range transport. OECD Series on Testing and Assessment No. 45, ENV/JM/MONO(2004)5. Paris: Organisation for Economic Cooperation and Development, Environment Directorate, Joint Meeting of the Chemicals Committee and the Working Party on Chemicals, Pesticides and Biotechnology.
- [OECD] Organisation for Economic Co-operation and Development. 2006. Comparison of emission estimation methods used in pollutant release and transfer registers and emission scenario documents: case study of pulp and paper and textile sectors. OECD Series on Testing and Assessment No. 52, ENV/JM/MONO(2006)6. Paris: Organisation for Economic Cooperation and Development, Environment Directorate, Joint Meeting of the Chemicals Committee and the Working Party on Chemicals, Pesticides and Biotechnology.

- Office of Emergency and Remedial Response. 1991. Risk assessment guidance for Superfund: human health evaluation manual: risk evaluation of remedial alternatives. Vol. 1, Part C, Publication 9285.7-01C. Washington (DC): US Environmental Protection Agency.
- O'Halloran K. 2006. Toxicological considerations of contaminants in the terrestrial environment for ecological risk assessment. *Human Ecol Risk Assess* 12:74–83.
- Oomen AG, Sips A, Groten JP, Sijm D, Tolls J. 2000. Mobilization of PCBs and lindane from soil during *in vitro* digestion and their distribution among bile salt micelles and proteins of human digestive fluid and the soil. *Environ Sci Technol* 34:297–303.
- Oomen AG, Tolls J, Kruidenier M, Bosgra SSD, Sips A, Groten JP. 2001. Availability of polychlorinated biphenyls (PCBs) and lindane for uptake by intestinal Caco-2 cells. *Environ Health Perspect* 109:731–737.
- Oomen AG, Tolls J, Sips A, Groten JP. 2003. *In vitro* intestinal lead uptake and transport in relation to speciation. *Archiv Environ Contam Toxicol* 44:116–124.
- [OSHA] Occupational Safety and Health Administration. 1993. Air contaminants; rule. 29 CFR 1910.1000. Occupational Safety and Health Administration. Federal Register 58(124):35338–35351.
- [OSHA] Occupational Safety and Health Administration. 2001. OSHA regulations (standards—29 CFR): air contaminants (standard number: 1910.1000). Washington (DC): Occupational Safety and Health Administration, US Department of Labor.
- Ou YC, Conolly RB, Thomas R, Gustafson DL, Long ME, Dovrev ID, Chubb LS, Xu Y, Lapidot S, Andersen ME, Yang RSH. 2003. Stochastic simulation of hepatic preneoplastic foci development for four chlorobenzene congeners in a medium-term bioassay. *Toxicol Sci* 73:301–314.
- Ou YC, Conolly RB, Thomas RS, Xu Y, Andersen ME, Chubb LS, Pitot HC, Yang RSH. 2001. A clonal growth model: time-course simulations of liver foci growth following penta- or hexachlorobenzene treatment in a medium-term bioassay. *Cancer Res* 61:1879–1889.
- Page DS, Boehm PD, Brown JS, Neff JM, Burns WA, Bence AE. 2005. Mussels document loss of bioavailable polycyclic aromatic hydrocarbons and the return to baseline conditions for oiled shorelines in Prince William Sound, Alaska. *Mar Environ Res* 60:422–436.
- Paquin PR, Gorsuch JW, Apte S, Batley GE, Bowles KC, Campbell PGC, Delos CG, Di Toro DM, Dwyer RL, Galvez F, Gensemer RW, Goss GG, Hogstrand C, Janssen CR, McGeer JC, Naddy RB, Playle RC, Santore RC, Schneider U, Stubblefield WA, Wood CM, Wu KB. 2002a. The biotic ligand model, a historical overview. *Comp Biochem Physiol C* 133:3–35.
- Paquin PR, Zoltay V, Winfield RP, Wu KB, Mathew R, Santore RC, Di Toro DM. 2002b. Extension of the biotic ligand model of acute toxicity to a physiologically-based model of the survival time of rainbow trout (*Oncorhynchus mykiss*) exposed to silver. *Comp Biochem Physiol C* 133:305–343.
- Payne J, Scholze M, Kortenkamp A. 2001. Mixtures of four organochlorines enhance human breast cancer cell proliferation. *Environ Health Perspect* 109:391–397.
- Peijnenburg WJGM, Jager T. 2003. Monitoring approaches to assess bioaccessibility and bioavailability of metals: matrix issues. *Ecotoxicol Environ Safety* 56:63–77.
- Pelekis M, Krishnan K. 1997. Assessing the relevance of rodent data on chemical interactions for health risk assessment purposes: a case study with dichloromethane-toluene mixture. *Regul Toxicol Pharmacol* 25:79–86.
- Pereira R, Ribeiro R, Goncalves F. 2004. Scalp hair analysis as a tool in assessing human exposure to heavy metals (S. Domingos mine, Portugal). *Sci Total Environ* 327:81–92.
- Pierik FH, Burdorf A, Deddens JA, Juttmann RE, Weber RFA. 2004. Maternal and paternal risk factors for cryptorchidism and hypospadias: a case-control study in newborn boys. *Environ Health Perspect* 112:1570–1576.

- Pieters BJ, Jager T, Kraak MHS, Admiraal W. 2006. Modeling responses of *Daphnia magna* to pesticide pulse exposure under varying food conditions: intrinsic versus apparent sensitivity. *Ecotoxicology* 15:601–608.
- Pilling ED, Bromley-Challenor KA, Walker CH, Jepson PC. 1995. Mechanism of synergism between the pyrethroid insecticide lambda-cyhalothrin and the imidazole fungicide prochloraz, in the honeybee (*Apis mellifera* L.). *Pest Biochem Physiol* 51:1–11.
- Plackett RL, Hewlett, PS. 1952. Quantal responses to mixtures of poisons. *J Royal Stat Soc Ser B* 14:141–163.
- Plackett RL, Hewlett PS. 1963a. A unified theory for quantal responses to mixtures of drugs: the fitting to data of certain models for two non-interactive drugs with complete positive correlation of tolerances. *Biometrics* 19:517–531.
- Plackett RL, Hewlett PS. 1963b. Quantal response to mixtures of poisons. *J R Stat Soc B* 14:141–163.
- Playle RC. 2004. Using multiple metal-gill binding models and the toxic unit concept to help reconcile multiple-metal toxicity results. *Aquat Toxicol* 67:359–370.
- Pösch G. 1993. Combined effects of drugs and toxic agents. New York: Springer-Verlag.
- Poet TS, Kousba AA, Dennison SL, Timchalk C. 2004. Physiologically based pharmacokinetic/pharmacodynamic model for the organophosphorus pesticide diazinon. *Neurotoxicology* 25:1013–1030.
- Pohl H, Hibbs B. 1996. Breast-feeding exposure of infants to environmental contaminants—a public health risk assessment viewpoint: chlorinated dibenzo-p-dioxins and chlorinated dibenzofurans. *Toxicol Ind Health* 12:593–611.
- Pohl HR, McClure P, De Rosa CT. 2004. Persistent chemicals found in breast milk and their possible interactions. *Environ Toxicol Pharmacol* 18:259–266.
- Pohl HR, Roney N, Wilbur S, Hansen H, De Rosa CT, 2003. Six interaction profiles for simple mixtures. *Chemosphere* 53:183–197.
- Pohl HR, Tylanda CA. 2000. Breast-feeding exposure of infants to selected pesticides: a public health viewpoint. *Toxicol Ind Health* 16:65–77.
- Pohl HR, van Engelen J, Wilson J, Sips A. 2005. Risk assessment of chemicals and pharmaceuticals in the pediatric population: a workshop report. *Regul Toxicol Pharmacol* 42:83–95.
- Poiger H, Schlatter C. 1986. Pharmacokinetics of 2,3,7,8-TCDD in man. *Chemosphere* 15:1489–1494.
- Poirier L, Berthet B, Amiard JC, Jeantet AY, Amiard-Triquet C. 2006. A suitable model for the biomonitoring of trace metal bioavailabilities in estuarine sediments: the annelid polychaete *Nereis diversicolor*. *J Mar Biol Assoc UK* 86:71–82.
- Posthuma L, Baerselman R, Van Veen RPM, Dirven-van Breemen EM. 1997. Single and joint toxic effects of copper and zinc on reproduction of *Enchytraeus crypticus* in relation to sorption of metals in soils. *Ecotoxicol Environ Safety* 38:108–121.
- Posthuma L, De Zwart D. 2006. Predicted effects of toxicant mixtures are confirmed by changes in fish species assemblages in Ohio, USA, rivers. *Environ Toxicol Chem* 25:1094–1105.
- Posthuma L, De Zwart D, Wintersen A, Lijzen J, Swartjes F, Cuypers C, Van Noort P, Harmsen J, Groenenberg BJ. 2006. Beslissen over bagger op bodem. Deel 1. Systeembenadering, model en praktijkvoorbeelden. Report 711701044. Bilthoven (NL): National Institute for Public Health and the Environment (RIVM).
- Posthuma L, Richards S, De Zwart D, Dyer SD, Sibley P, Hickey C, Altenburger R. 2008. Mixture extrapolation approaches. In: Solomon KR, Brock TCM, De Zwart D, Dyer SD, Posthuma L, Richards S, Sanderson H, Sibley R, Van den Brink PJ, editors, Extrapolation practice for ecotoxicological effect characterization of chemicals. Results of the EXPECT workshop, February 2005, St. Petersburg, FL, USA. Pensacola (FL): SETAC Press.

- Posthuma L, Traas TP, Suter GW II, editors. 2002. Species sensitivity distributions in ecotoxicology. Boca Raton (FL): Lewis Publishers.
- Posthuma L, Van Straalen NM. 1993. Heavy-metal adaptation in terrestrial invertebrates: a review of occurrence, genetics, physiology and ecological consequences. *Comp Biochem Physiol C* 106:11–38.
- Posthumus R, Traas TP, Peijnenburg W, Hulzebos EM. 2005. External validation of EPIWIN biodegradation models. *SAR QSAR Environ Res* 16:135–148.
- Poulin P, Schoenlein K, Theil FP. 2001. Prediction of adipose tissue: plasma partition coefficients for structurally unrelated drugs. *J Pharm Sci* 90:436–447.
- Poulin P, Theil FP. 2002. Prediction of pharmacokinetics prior to *in vivo* studies. 1. Mechanism-based prediction of volume of distribution. *J Pharm Sci* 91:129–156.
- Price K, Haddad S, Krishnan K. 2003a. Physiological modeling of age-specific changes in the pharmacokinetics of organic chemicals in children. *J Toxicol Environ Health A* 66:417–433.
- Price K, Krishnan K. 2005. An integrated QSAR-PBPK model for simulating pharmacokinetics of chemicals in mixtures. 44th Annual Meeting of the Society of Toxicology, New Orleans (LA), March 6–10.
- Price PS, Conolly RB, Chaisson CF, Gross EA, Young JS, Mathis ET, Tedder DR. 2003b. Modeling interindividual variation in physiological factors used in PBPK models of humans. *Crit Rev Toxicol* 33:469–503.
- Psaty BM, Furberg CD, Ray WA, Weiss NS. 2004. Potential for conflict of interest in the evaluation of suspected adverse drug reactions. *J Am Med Assoc* 292:2622–2631.
- Purcell KJ, Cason GH, Gargas ML, Andersen ME, Travis CC. 1990. *In vivo* metabolic interactions of benzene and toluene. *Toxicol Lett* 52:141–152.
- Putzrath RM. 2000. Reducing uncertainty of risk estimates for mixtures of chemicals within regulatory constraints. *Regul Toxicol Pharmacol* 31:44–52.
- Ra JS, Lee BC, Chang NI, Kim SD. 2006. Estimating the combined toxicity by two-step prediction model on the complicated chemical mixtures from wastewater treatment plant effluents. *Environ Toxicol Chem* 25:2107–2113.
- Ragas AMJ, Etienne RS, Willemssen FH, Van de Meent D. 1999. Assessing model uncertainty for environmental decision making: a case study of the coherence of independently derived environmental quality objectives for air and water. *Environ Toxicol Chem* 18:1856–1867.
- Rainbow PS. 2002. Trace metal concentrations in aquatic invertebrates: why and so what? *Environ Pollut* 120:497–507.
- Rainbow PS, Fialkowski W, Sokolowski A, Smith BD, Wolowicz M. 2004. Geographical and seasonal variation of trace metal bioavailabilities in the Gulf of Gdansk, Baltic Sea using mussels (*Mytilus trossulus*) and barnacles (*Balanus improvisus*) as biomonitors. *Mar Biol* 144:271–286.
- Rajapakse N, Silva E, Kortenkamp A. 2002. Combining xenoestrogens at levels below individual no-observed effect concentrations dramatically enhances steroid hormone action. *Environ Health Perspect* 110:917–921.
- Raymond JW, Rogers TN, Shonnard DR, Kline AA. 2001. A review of structure-based biodegradation estimation methods. *J Hazardous Mater* 84:189–215.
- Read HJ, Martin MH. 1993. The effects of heavy metals on populations of small mammals from woodlands in Avon (England); with particular emphasis on metal concentrations in *Sorex araneus* L. and *Sorex minutus* L. *Chemosphere* 27:2197–2211.
- Redding LE, Sohn MD, McKone TE, Chen JW, Wang SL, Hsieh DPH, Yang RSH. 2008. Population physiologically-based pharmacokinetic modeling for the human lactational transfer of PCB 153 with consideration of worldwide human biomonitoring results. *Environ Health Perspect* 116:1629–1634.
- Reffstrup TK. 2002. Combined actions of pesticides in food. Report 2002:19. Soborg (DK): Danish Veterinary and Food Administration.

- Regnell O. 1994. The effect of pH and dissolved oxygen levels on methylation and partitioning of mercury in freshwater model systems. *Environ Pollut* 84:7–13.
- Reichenberg F, Mayer P. 2006 Two complementary sides of bioavailability: accessibility and chemical activity of organic contaminants in sediments and soils. *Environ Toxicol Chem* 25:1239–1245.
- Reisfeld B, Mayeno AN, Lyons MA, Yang RSH. 2007. Physiologically-based pharmacokinetic and pharmacodynamic modeling, in computational toxicology. In: Ekins S, editor, Risk assessment for pharmaceutical and environmental chemicals. Hoboken (NJ): John Wiley & Sons, p 33–69.
- Reisfeld B, Yang RSH. 2004. A reaction network model for CYP2E1-mediated metabolism of toxicant mixtures. *Environ Toxicol Pharmacol* 18:173–179.
- Renn O, Benighaus C. 2006. Framing the perception of cumulative stressors, especially chemical risks. Report on approaches to the characterization of knowledge of risks, uncertainties and ambiguity and their use and quality assurance in the IP domain. EU FP6 Project NOMIRACLE, Deliverable 4.3.2. Stuttgart (DE): Dialogik.
- Renwick AG, Hayes AW. 2001. Toxicokinetics: pharmacokinetics in toxicology. 4th ed. Philadelphia (PA): Taylor and Francis.
- Reynders H, Van Campenhout K, Bervoets L, De Coen WM, Blust R. 2006. Dynamics of cadmium accumulation and effects in common carp (*Cyprinus carpio*) during simultaneous exposure to water and food (*Tubifex tubifex*). *Environ Toxicol Chem* 25:1558–1567.
- Rice GE, Teuschler LK, Bull RJ, Feder PI, Simmons JE. 2009. Evaluating the similarity of complex drinking water disinfection by-product mixtures: overview of the issues. *J Toxicol Environ Health A* 72:429–436.
- Rider CV, Furr J, Wilson VS, Gray LE Jr. 2008. A mixture of seven antiandrogens induces reproductive malformations in rats. *Int J Androl* 31:249–262.
- Riley RJ, McGinnity DF, Austin RP. 2005. A unified model for predicting human hepatic, metabolic clearance from *in vitro* intrinsic clearance data in hepatocytes and microsomes. *Drug Metab Dispos* 33:1304–1311.
- Riviere JE, Brooks JD. 2007. Prediction of dermal absorption from complex chemical mixtures: incorporation of vehicle effects and interactions into a QSPR framework. *SAR QSAR Environ Res* 18:31–44.
- Rodgers T, Leahy D, Rowland M. 2005. Physiologically based pharmacokinetic modelling. 1. Predicting the tissue distribution of moderate-to-strong bases. *J Pharm Sci* 94:1259–1276.
- Rodgers T, Rowland M. 2006. Physiologically based pharmacokinetic modelling. 2. Predicting the tissue distribution of acids, very weak bases, neutrals and zwitterions. *J Pharm Sci* 95:1238–1257.
- Roelofs D, Mariën J, Van Straalen NM. 2007. Differential gene expression profiles associated with heavy metal tolerance in the soil insect *Orchesella cincta*. *Insect Biochem Mol Biol* 37:287–295.
- Rogan WJ, Gladen BC, McKinney JD, Carreras N, Hardy P, Thullen J, Tingelstad J, Tully M. 1986. Polychlorinated biphenyls (PCBs) and dichlorophenyl dichloroethene (DDE) in human milk: effects of maternal factors and previous lactation. *Am J Public Health* 76:172–177.
- Romijn CAFM, Luttik R, Canton J. 1993a. Presentation of a general algorithm to include effect assessment on secondary poisoning in the derivation of environmental quality criteria. 2. Terrestrial food chains. *Ecotoxicol Environ Safety* 26:61–83.
- Romijn CAFM, Luttik R, Van de Meent D, Sloof W, Canton J. 1993b. Presentation of a general algorithm to include effect assessment on secondary poisoning in the derivation of environmental quality criteria. 1. Aquatic food chains. *Ecotoxicol Environ Safety* 26:61–83.

- Ross HLB. 1996. The interaction of chemical mixtures and their implications on water quality guidelines. Hon thesis, University of Technology, Sydney, NSW (AU).
- Ross HLB, Warne MStJ. 1997. Most chemical mixtures have additive aquatic toxicity. In Proceedings of the Third Annual Conference of the Australasian Society for Ecotoxicology, Brisbane (AU), July 17–19, p 30.
- Rowland M, Tozer TN. 1995. Clinical pharmacokinetics concepts and applications. 3rd ed. Media (PA): Williams & Williams.
- Rozman KK, Doull J. 2000. Dose and time as variables of toxicity. *Toxicology* 144:169–178.
- Russel FG, Wouterse AC, Van Ginneken CA. 1987. Physiologically based pharmacokinetic model for the renal clearance of salicylic acid and the interaction with phenolsulfonphthalein in the dog. *Drug Metab Dispos* 15:695–701.
- Russel FG, Wouterse AC, Van Ginneken CA. 1989. Physiologically based pharmacokinetic model for the renal clearance of iodopyracet and the interaction with probenecid in the dog. *Biopharm Drug Dispos* 10:137–152.
- Sanchez-Dardon J, Voccia I, Hontela A, Chilmonczyk S, Dunier M, Boermans H, Blakley B, Fournier M. 1999. Immunomodulation by heavy metals tested individually or in mixtures in rainbow trout (*Oncorhynchus mykiss*) exposed *in vivo*. *Environ Toxicol Chem* 18:1492–1497.
- Sarangapani R, Teeguarden J, Plotzke KP, McKim JM Jr, Andersen ME. 2002. Dose–response modeling of cytochrome p450 induction in rats by octamethylcyclotetrasiloxane. *Toxicol Sci* 67:159–172.
- Schechter A, Gasiewicz TA. 1987a. Health hazard assessment of chlorinated dioxins and dibenzofurans contained in human milk. *Chemosphere* 16:2147–2154.
- Schechter A, Gasiewicz TA. 1987b. Human breast milk levels of dioxins and dibenzofurans: significance with respect to current risk assessments. *ACS Symp Ser* 338:162–173.
- Schechter A, Li L. 1997. Dioxins, dibenzofurans, dioxin-like PCBs, and DDE in US fast food, 1995. *Chemosphere* 34:1449–1457.
- Schechter A, Startin J, Wright C, Kelly M, Papke O, Lis A, Ball M, Olson JR. 1994. Congener-specific levels of dioxins and dibenzofurans in US food and estimated daily dioxin toxic equivalent intake. *Environ Health Perspect* 102:962–966.
- Schmider J, von Moltke LL, Shader RI, Harmatz JS, Greenblatt DJ. 1999. Extrapolating *in vitro* data on drug metabolism to *in vivo* pharmacokinetics: evaluation of the pharmacokinetic interaction between amitriptyline and fluoxetine. *Drug Metab Rev* 31:545–560.
- Scholz NL, Truelove NK, Labenia JS, Baldwin DH, Collier TK. 2006. Dose-additive inhibition of chinook salmon acetylcholinesterase activity by mixtures of organophosphate and carbamate insecticides. *Environ Toxicol Chem* 25:1200–1207.
- Scholze M, Boedeker W, Faust M, Backhaus T, Altenburger R, Grimme LH. 2001. A general best-fit method for concentration–response curves and the estimation of low-effect concentrations. *Environ Toxicol Chem* 20:448–457.
- Schramm KW. 1990. Exams 2—Exposure analysis modeling system. *Toxicol Environ Chem* 26:73–82.
- Schuler LJ, Landrum PF, Lydy MJ. 2006. Comparative toxicity of fluoranthene and pentachlorobenzene to three freshwater invertebrates. *Environ Toxicol Chem* 25:985–994.
- Scott Fordsmand JJ, Krogh PH, Weeks JM. 2000. Responses of *Folsomia fimetaria* (Collembola: Isotomidae) to copper under different soil copper contamination histories in relation to risk assessment. *Environ Toxicol Chem* 19:1297–1303.
- Segel IH. 1975. Enzyme kinetics: behavior and analysis of rapid equilibrium and steady-state enzyme systems. Toronto (CA): John Wiley & Sons.
- Selikoff IJ, Seidman H, Hammond C. 1980. Mortality effects of cigarette smoking among site asbestos factory workers. *J Natl Cancer Inst* 65:507–513.

- [SETAC] Society of Environmental Toxicology and Chemistry. 2004. Technical issue paper: whole effluent toxicity testing. Pensacola (FL): Society of Environmental Toxicology and Chemistry.
- Shakman RA. 1974. Nutritional influences on the toxicity of environmental pollutants. *Arch Environ Health* 28:105–113.
- Sharma-Shanti S, Schat H, Vooijs R, Van Heerwaarden LM. 1999. Combination toxicology of copper, zinc, and cadmium in binary mixtures: concentration-dependent antagonistic, nonadditive, and synergistic effects on root growth in *Silene vulgaris*. *Environ Toxicol Chem* 18:348–355.
- Shin KH, Ahn Y, Kim KW. 2005. Toxic effect of biosurfactant addition on the biodegradation of phenanthrene. *Environ Toxicol Chem* 24:2768–2774.
- Shiverick KT, Slikker W, Rogerson SJ, Miller RK. 2003. Drugs and the placenta—a workshop report. *Placenta* 24:S55–S59.
- Siegrist M, Cvetkovich G. 2001. Better negative than positive? Evidence of a bias for negative information about possible health dangers. *Risk Anal* 21:199–206.
- Sijm DTHM, Van der Linde A. 1995. Size-dependent bioconcentration kinetics of hydrophobic organic chemicals in fish based on diffusive mass transfer and allometric relationships. *Environ Sci Technol* 29:2769–2777.
- Silva CAR, Rainbow PS, Smith BD. 2003. Biomonitoring of trace metal contamination in mangrove-lined Brazilian coastal systems using the oyster *Crassostrea rhizophorae*: comparative study of regions affected by oil, salt pond and shrimp farming activities. *Hydrobiologia* 501:199–206.
- Silva E, Rajapakse N, Kortenkamp A. 2002. Something from “nothing”—eight weak estrogenic chemicals combined at concentrations below NOECs produce significant mixture effects. *Environ Sci Technol* 36:1751–1756.
- Simmons JE, Richardson SD, Speth TF, Miltner RJ, Rice G, Schenck K, Hunter III ES, Teuschler LK. 2002. Development of a research strategy for integrated technology-based toxicological and chemical evaluation of complex mixtures of drinking water disinfection byproducts. *Environ Health Perspect* 110:1013–1024.
- Sjodin A, Jones RS, Focant JF, Lapeza C, Wang RY, McGahee EE III, Zhang Y, Turner WE, Slazyk B, Needham LL, Patterson DG Jr. 2004. Retrospective time-trend study of polybrominated diphenyl ether and polybrominated and polychlorinated biphenyl levels in human serum from the United States. *Environ Health Perspect* 112:654–658.
- Skaggs SM, Foti RS, Fisher MB. 2006. A streamlined method to predict hepatic clearance using human liver microsomes in the presence of human plasma. *J Pharmacol Toxicol Methods* 53:284–290.
- Slaveykova VI, Wilkinson KJ. 2005. Predicting the bioavailability of metals and metal complexes: critical review of the biotic ligand model. *Environ Chem* 2:9–24.
- Slooff W, De Zwart D. 1991. The pT-value as environmental policy indicator for the exposure to toxic substances. Report nr. 719102 003. Bilthoven (NL): National Institute for Public Health and the Environment (RIVM).
- Smit CE, Van Gestel CAM. 1998. Effects of soil type, prepercolation, and ageing on bioaccumulation and toxicity of zinc for the springtail *Folsomia candida*. *Environ Toxicol Chem* 17:1132–1141.
- Sokal RR, Rohlf FJ. 1995. Biometry, the principles and practice of statistics in biological research. 3rd ed. San Francisco (CA): Freeman.
- Sole M, Porte C, Barcelo D, Albaiges J. 2000. Bivalves residue analysis for the assessment of coastal pollution in the Ebro Delta (NW Mediterranean). *Mar Pollut Bull* 40:746–753.
- Solomon KR, Brock TCM, De Zwart D, Dyer SD, Posthuma L, Richards S, Sanderson H, Sibley P, Van den Brink PJ. 2008. Extrapolation practice for ecotoxicological effect characterization of chemicals. Pensacola (FL): SETAC Press.

- Sonzogni W, Maack L, Degenhardt D, Anderson H, Fiore B. 1991. Polychlorinated biphenyl congeners in blood of Wisconsin sport fish consumers. *Arch Environ Contam Toxicol* 20:56–60.
- Sørensen PB, Vorkamp K, Thomsen M. 2004. Persistent organic pollutants (POPs) in the Greenland environment—long-term temporal changes and effects on eggs of a bird of prey. NERI Technical Report 509. Silkeborg (DK): National Environment Research Institute.
- Speijers GJA, Speijers MHM. 2004. Combined toxic effects of mycotoxins. *Toxicol Lett* 153:91–98.
- Sprague JB. 1970. Measurement of pollutant toxicity to fish. II. Utilizing and applying bioassay results. *Water Res* 4:3–32.
- Spurgeon DJ, Hopkin SP. 1996. Risk assessment of the threat of secondary poisoning by metals of predators of earthworms in the vicinity of a primary smelting works. *Sci Total Environ* 187:167–183.
- Squillace PJ, Scott JC, Moran MJ, Nolan T, Koplin DW. 2002. VOCs, pesticides, nitrate, and their mixtures in groundwater used for drinking water in the United States. *Environ Sci Technol* 36:1923–1930.
- Stark JD. 2006. Toxicity endpoints used in risk assessment: what do they really mean? *SETAC Globe* 7(2):29–30.
- Staunton S. 2004. Sensitivity analysis of the distribution coefficient, K_d , of nickel with changing soil chemical properties. *Geoderma* 122:281–290.
- Steen Redeker E, Bervoets L, Blust R. 2004. Dynamic model for the accumulation of cadmium and zinc from water and sediment by the aquatic oligochaete, *Tubifex tubifex*. *Environ Sci Technol* 38:6193–6200.
- Steen Redeker E, Blust R. 2004. Accumulation and toxicity of cadmium in the aquatic oligochaete *Tubifex tubifex*: a kinetic modeling approach. *Environ Sci Technol* 38:537–543.
- Steevens JA, Benson WH. 1999. Toxicological interactions of chlorpyrifos and methyl mercury in the amphipod, *Hyalella azteca*. *Toxicol Sci* 52:168–177.
- Stefanelli P, Ausili A, Di Muccio A, Fossi C, Di Muccio S, Rossi S, Colasanti A. 2004. Organochlorine compounds in tissues of swordfish (*Xiphias gladius*) from Mediterranean Sea and Azores islands. *Mar Pollut Bull* 49:938–950.
- Sterner TR, Robinson PJ, Mattie DR, Burton GA. 2005. The toxicology of chemical mixtures risk assessment for human and ecological receptors. AFRL-HE-WP-TR-2005-0173. Wright-Patterson AFB (OH): Air Force Research Laboratory, Human Effectiveness Directorate, Biosciences and Protection Division, Applied Biotechnology Branch.
- Stork LG, Gennings C, Carter WH Jr, Teuschler LK, Carney EW. 2008. Empirical evaluation of sufficient similarity in dose–response for environmental risk assessment of chemical mixtures. *J Agric Biol Environ Stat* 13:313–333.
- Strenkowski-Nix LC, Forrest A, Schentag JJ, Nix DE. 1998. Pharmacodynamic interactions of ciprofloxacin, piperacillin, and piperacillin/tazobactam in healthy volunteers. *J Clin Pharmacol* 38:1063–1071.
- Struijs J. 2003. Evaluatie van pT. De bepaling van toxische stress in Rijkswateren. Report nr 860703001. Bilthoven (NL): National Institute for Public Health and the Environment (RIVM).
- Sudaryanto A, Takahashi S, Monirith I, Ismail A, Muchtar M, Zheng J, Richardson BJ, Subramanian A, Prudente M, Hue ND, Tanabe S. 2002. Asia-Pacific mussel watch: monitoring of butyltin contamination in coastal waters of Asian developing countries. *Environ Toxicol Chem* 21:2119–2130.
- Sugita O, Sawada Y, Sugiyama Y, Iga T, Hanano M. 1982. Physiologically based pharmacokinetics of drug–drug interaction: a study of tolbutamide–sulfonamide interaction in rats. *J Pharmacokin Biopharm* 10:297–316.

- Sühnel J. 1992. Assessment of interaction of biologically active agents by means of the isobole approach: fundamental assumptions and recent developments. *ACES* 4:35–44.
- Suter II GW, Munns WR Jr, Sekizawa W. 2003. Types of integration in risk assessment and management, and why they are needed. *Human Ecol Risk Assess* 9:273–279.
- Suzuki H, Iwatsubo T, Sugiyama Y. 1995. Applications and prospects for physiologically based pharmacokinetic (PB-PK) models involving pharmaceutical agents. *Toxicol Lett* 82/83:349–355.
- Svenson A, Sanden B, Dalhammar G, Remberger M, Kaj L. 2000. Toxicity identification and evaluation of nitrification inhibitors in wastewaters. *Environ Toxicol* 15:527–532.
- Swain S, Wren J, Stürzenbaum SR, Kille P, Morgan AJ, Jager T, Jonker MJ, Hankard PK, Svendsen C, Owen J, Hedley BA, Blaxter M, Spurgeon DJ. 2010. Linking toxicants mechanism of action and physiological mode of action in *Caenorhabditis elegans*. *BMC Biol*.
- Swan SH, Kruse RL, Liu F, Barr DB, Drobnis EZ, Redmon JB, Wang C, Brazil C, Overstreet JW, the Study for Future Families Research Group. 2003. Semen quality in relation to biomarkers of pesticide exposure. *Environ Health Perspect* 111:1478–1484.
- Swan SH, Main KM, Liu F, Stewart SL, Kruse RL, Calafat AM, Mao CS, Redmon JB, Ternand CL, Sullivan S, Teague JL, the Study for Future Families Research Group. 2005. Decrease in anogenital distance among male infants with prenatal phthalate exposure. *Environ Health Perspect* 113:1056–1061.
- Tanabe S. 2000. Asia-Pacific mussel watch progress report. *Mar Pollut Bull* 40:651–651.
- Tardif R, Charest-Tardif G. 1999. The importance of measured end-points in demonstrating the occurrence of interactions: a case study with methylchloroform and m-xylene. *Toxicol Sci* 49:312–317.
- Tardif R, Charest-Tardif G, Brodeur J, Krishnan K. 1997. Physiologically based pharmacokinetic modeling of a ternary mixture of alkyl benzenes in rats and humans. *Toxicol Appl Pharmacol* 144:120–134.
- Tardif R, Lapare S, Charest-Tardif G, Brodeur J, Krishnan K. 1995. Physiologically-based pharmacokinetic modeling of a mixture of toluene and xylene in humans. *Risk Anal* 15:335–342.
- Tardif R, Lapare S, Krishnan K, Brodeur J. 1993. Physiologically based modeling of the toxicokinetic interaction between toluene and m-xylene in the rat. *Toxicol Appl Pharmacol* 120:266–273.
- Ter Laak TL, Agbo SO, Barendregt A, Hermens JLM. 2006. Freely dissolved concentrations of PAHs in soil pore water: measurements via solid-phase extraction and consequences for soil tests. *Environ Sci Technol* 40:1307–1313.
- Teuschler LK. 2007. Deciding which chemical mixtures risk assessment methods work best for what mixtures. *Toxicol Appl Pharmacol* 223:139–147.
- Teuschler LK, Gennings C, Stiteler WM, Hertzberg RC, Colman JT, Thiagarajah A, Lipscomb JC, Hartley WR, Simmons JE. 2000. A multiple-purpose design approach to the evaluation of risks from mixtures of disinfection by-products. *Drug Chem Toxicol* 23:307–321.
- Teuschler LK, Klaunig J, Carney E, Chambers J, Conolly R, Gennings C, Giesy J, Hertzberg R, Klaassen C, Kodell R, Paustenbach D, Yang R. 2002. Support of science-based decisions concerning the evaluation of the toxicology of mixtures: a new beginning. *Reg Toxicol Pharmacol* 36:34–39.
- Teuschler LK, Rice GE, Wilkes CR, Lipscomb JC, Power FW. 2004. A feasibility study of cumulative risk assessment methods for drinking water disinfection by-product mixtures. *J Toxicol Environ Health A* 67:755–777.
- Theil FP, Guentert TW, Haddad S, Poulin P. 2003. Utility of physiologically based pharmacokinetic models to drug development and rational drug discovery candidate selection. *Toxicol Lett* 138:29–49.

- Thomas GO, Wilkinson M, Hodson S, Jones KC. 2006. Organohalogen chemicals in human blood from the United Kingdom. *Environ Pollut* 141:30–41.
- Thomas RS, Conolly RB, Gustafson DL, Long ME, Benjamin SA, Yang RSH. 2000. A physiologically based pharmacodynamic analysis of hepatic foci within a medium-term liver bioassay using pentachlorobenzene as a promoter and diethylnitrosamine as an initiator. *Toxicol Appl Pharmacol* 166:128–137.
- Thomsen M, Sørensen PB, Fauser P, Ragas A, Peirano F. 2006. Prioritised listing of VOCs/semi-VOCs including test scenarios. Report D.1.2.2 from EC FP6-IP NoMiracle, restricted.
- Thorpe KL, Gross-Sorokin M, Johnson I, Brighty G, Tyler C. 2006. An assessment of the model of concentration addition for predicting the estrogenic activity of chemical mixtures in wastewater treatment works effluents. *Environ Health Perspect* 114(Suppl 1):90–97.
- Thrall KD, Poet TS. 2000. Determination of biokinetic interactions in chemical mixtures using real-time breath analysis and physiologically based pharmacokinetic modeling. *J Toxicol Environ Health A* 59:653–670.
- Timchalk C, Poet TS. 2008. Development of a physiologically based pharmacokinetic and pharmacodynamic model to determine dosimetry and cholinesterase inhibition for a binary mixture of chlorpyrifos and diazinon in the rat. *Neurotoxicology* 29:428–443.
- Tinwell H, Ashby J. 2004. Sensitivity of the immature rat uterotrophic assay to mixtures of estrogens. *Environ Health Perspect* 112:575–582.
- Toose L, Woodfine DG, MacLeod M, Mackay D, Gouin J. 2004. BETR-World: a geographically explicit model of chemical fate: application to transport of alpha-HCH to the Arctic. *Environ Pollut* 128:223–240.
- Tozer TN, Rowland M. 2006. Introduction to pharmacokinetics and pharmacodynamics: the quantitative basis of drug therapy. Baltimore (MD): Lippincott Williams & Wilkins.
- Trapp S, Matthies M. 1995. Generic one-compartment model for uptake of organic chemicals by foliar vegetation. *Environ Sci Technol* 29:2333–2338.
- Trapp S, McFarlane JC. 1995. Plant contamination: modeling and simulation of organic chemical processes. Boca Raton (FL): Lewis Publishers.
- Tuk B, van Gool T, Danhof M. 2002. Mechanism-based pharmacodynamic modeling of the interaction of midazolam, bretazenil, and zolpidem with ethanol. *J Pharmacokinet Pharmacodyn* 29:235–250.
- Umbreit TH, Hesse EJ, Gallo MA. 1986a. Differential bioavailability of TCDD from contaminated soils. *Abstracts Am Chem Soc* 191:47.
- Umbreit TH, Hesse EJ, Gallo MA. 1986b. Comparative toxicity of TCDD contaminated soil from Times Beach, Missouri, and Newark, New Jersey. *Chemosphere* 15:2121–2124.
- [USEPA] US Environmental Protection Agency. 1986. Guidelines for health risk assessment of chemical mixtures. US Environmental Protection Agency. *Federal Register* 51(185):34014–34025.
- [USEPA]. US Environmental Protection Agency. 1989a. Exposure factors handbook. USEPA/600/8-89/043. Washington (DC): Office of Health and Environmental Assessment: US Environmental Protection Agency.
- [USEPA] US Environmental Protection Agency. 1989b. Risk assessment guidance for superfund: human health evaluation manual. Vol. 1, Part A, EPA/540/1-89/002. Washington (DC): Office of Emergency and Remedial Response: US Environmental Protection Agency.
- [USEPA] US Environmental Protection Agency. 1991. Methods for aquatic toxicity identification evaluations. Phase I. Toxicity characterization procedures. 2nd ed., EPA/600/6-91/003. Washington (DC): Office of Research and Development: US Environmental Protection Agency.

- [USEPA] US Environmental Protection Agency. 1994. EPA's national water quality inventory: 1992. Report to Congress. Report 841-R-94-001. Washington (DC): US Environmental Protection Agency.
- [USEPA] US Environmental Protection Agency. 1995. Whole effluent toxicity: guidelines establishing test procedures for the analysis of pollutants. Office of Science and Technology, US Environmental Protection Agency. Federal Register 60(199):53529–53544.
- [USEPA] US Environmental Protection Agency. 1996. PCBs: cancer dose–response assessment and application to environmental mixtures. EPA/600/P-96/001F. Washington (DC): US Environmental Protection Agency, National Center for Environmental Assessment.
- [USEPA] US Environmental Protection Agency. 1997. Ecological risk assessment guidance for Superfund, process for designing and conducting ecological risk assessments. EPA 540-R-97-006. Washington (DC): Office of Solid Waste and Emergency Response: US Environmental Protection Agency.
- [USEPA] US Environmental Protection Agency. 1998. Guidelines for ecological risk assessment. EPA/630/R-95/002F. US Environmental Protection Agency, Risk Assessment Forum. Federal Register 63(93):26846–26924.
- [USEPA] US Environmental Protection Agency. 1999. Residual risk report to Congress. EPA-453/R-99-00. Triangle Park (NC): Office of Air Quality, Planning and Standards: US Environmental Protection Agency.
- [USEPA] US Environmental Protection Agency. 2000a. Exposure and human health reassessment of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) and related compounds [draft final]. Part II, EPA/600P-00/001(September). Washington (DC).
- [USEPA] US Environmental Protection Agency. 2000b. Supplementary guidance for conducting health risk assessment of chemical mixtures. EPA/630/R-00/002, ORD/NCEA. Cincinnati (OH): US Environmental Protection Agency.
- [USEPA] US Environmental Protection Agency. 2002a. Child-specific exposure factors handbook. EPA-600-P-00-002B, NTIS PB2003-101678. Washington (DC): National Center for Environmental Assessment, Office of Research and Development.
- [USEPA] US Environmental Protection Agency. 2002b. Guidance on cumulative risk assessment of pesticide chemicals that have a common mechanism of toxicity. Washington (DC): Office of Pesticide Programs. Available from: http://www.epa.gov/oppfead1/trac/science/cumulative_guidance.pdf
- [USEPA] US Environmental Protection Agency. 2002c. Organophosphate pesticides: revised cumulative risk assessment. Available from: <http://www.epa.gov/pesticides/cumulative/rra-op/>
- [USEPA] US Environmental Protection Agency. 2002d. Methods for measuring the acute toxicity of effluents and receiving waters to freshwater and marine organisms. 5th ed., EPA-821-R-02-012. Washington (DC): US Environmental Protection Agency.
- [USEPA] US Environmental Protection Agency. 2002e. Short-term methods for estimating the chronic toxicity of effluents and receiving waters to freshwater organisms 4th ed., EPA-821-R-02-013. Washington (DC): Office of Water: US Environmental Protection Agency.
- [USEPA] US Environmental Protection Agency. 2002f. Short-term methods for estimating the chronic toxicity of effluents and receiving waters to marine and estuarine organisms. 3rd ed., EPA-821-R-02-014. Washington (DC): Office of Water: US Environmental Protection Agency.
- [USEPA] US Environmental Protection Agency. 2003a. The feasibility of performing cumulative risk assessments for mixtures of disinfection by-products in drinking water. EPA/600/R-03/051, ORD/NCEA. Cincinnati (OH): US Environmental Protection Agency.

- [USEPA] US Environmental Protection Agency. 2003b. Developing relative potency factors for pesticide mixtures: biostatistical analyses of joint dose–response. EPA/600/R-03/052, ORD/NCEA. Cincinnati (OH): US Environmental Protection Agency.
- [USEPA] US Environmental Protection Agency. 2004. National whole effluent toxicity (WET) implementation guidance under the NPDES program [Draft]. EPA 832-B-04-003. Washington (DC): US Environmental Protection Agency. Office of Wastewater Management.
- [USEPA] US Environmental Protection Agency. 2005. ECOTOX database. US Environmental Protection Agency. Available from: <http://cfpub.epa.gov/ecotox/>
- [USEPA] US Environmental Protection Agency. 2006a. Exposures and internal doses of trihalomethanes in humans: multi-route contributions from drinking water. EPA/600/R-06/087, ORD/NCEA. Cincinnati (OH): US Environmental Protection Agency.
- [USEPA] US Environmental Protection Agency. 2006b. Available from: <http://www.epa.gov/iaq/voc.html>; <http://www.epa.gov/iaq/pubs/insidest.html>
- [USEPA] US Environmental Protection Agency. 2008. Integrated Risk Information System (IRIS). Available from: <http://cfpub.epa.gov/ncea/iris/index.cfm>
- [US PCCRARM] Presidential/Congressional Commission on Risk Assessment and Risk Management. 1997. Framework for environmental health risk management. Washington (DC): US Presidential/Congressional Commission on Risk Assessment and Risk Management.
- Van Brummelen TC, Van Gestel CAM, Verweij RA. 1996a. Long-term toxicity of five polycyclic aromatic hydrocarbons for the terrestrial isopods *Oniscus asellus* and *Porcellio scaber*. Environ Toxicol Chem 15:1199–1210.
- Van Brummelen TC, Verweij RA, Wedzinga SA, Van Gestel CAM. 1996b. Polycyclic aromatic hydrocarbons in earthworms and isopods from contaminated forest soils. Chemosphere 32:315–341.
- Van den Berg M, Birnbaum L, Bosveld ATC, Brunström B, Cook P, Feeley M, Giesy JP, Hanberg A, Hasegawa R, Kennedy SW, Kubiak T, Larsen JC, van Leeuwen FXR, Liem AKD, Nolt C, Peterson RE, Poellinger L, Safe S, Schrenk D, Tillitt D, Tysklind M, Younes M, Waern F, Zacharewski T. 1998. Toxic equivalency factors (TEFs) for PCBs, PCDDs, PCDFs for humans and wildlife. Environ Health Perspect 106:775–792.
- Van den Berg M, Birnbaum LS, Denison M, DeVito M, Farland W, Feeley M, Fiedler H, Hakansson H, Hanberg A, Haws L, Rose M, Safe S, Schrenk D, Tohyama C, Tritscher A, Tuomisto J, Tysklind M, Walker N, Peterson RE. 2006. Review: the 2005 World Health Organization reevaluation of human and mammalian toxic equivalency factors for dioxins and dioxin-like compounds. Toxicol Sci 93:223–241.
- Van den Brink PJ, Roelsma J, Van Nes EH, Scheffer M, Brock TCM. 2002. PERPEST model, a case-based reasoning approach to predict ecological risks of pesticides. Environ Toxicol Chem 21:2500–2506.
- Van der Geest HG, Greve GD, Boivin ME, Kraak MHS, Van Gestel CAM. 2000. Mixture toxicity of copper and diazinon to larvae of the mayfly (*Ephoron virgo*) judging additivity at different effect levels. Environ Toxicol Chem 19:2900–2905.
- Van der Oost R, Beyer J, Vermeulen NPE. 2003. Fish bioaccumulation and biomarkers in environmental risk assessment: a review. Environ Toxicol Pharmacol 13:57–149.
- Van Ewijk PH, Hoekstra JA. 1993. Calculation of the EC50 and its confidence interval when subtoxic stimulus is present. Ecotoxicol Environ Safety 25:25–32.
- van Gestel CAM, Hensbergen PJ. 1997. Interaction of Cd and Zn toxicity for *Folsomia candida* Willem (Collembola: Isotomidae) in relation to bioavailability in soil. Environ Toxicol Chem 16:1177–1186.
- Van Leeuwen CJ, Hermens JLM, editors. 1995. Risk assessment of chemicals: an introduction. 2nd ed. Dordrecht (NL): Kluwer Academic Publishers.

- Van Leeuwen CJ, Vermeire T, editors. 2007. Risk assessment of chemicals: an introduction. Dordrecht (NL): Kluwer Academic Publishers.
- Van Leeuwen IMM, Zonneveld C, Kooijman SALM. 2003. The embedded tumour: host physiology is important for the evaluation of tumour growth. *Br J Cancer* 89:2254–2263.
- Van Meeuwen JA, van den Berg M, Sanderson JT, Verhoef A, Piersma AH. 2007. Estrogenic effects of mixtures of phyto- and synthetic chemicals on uterine growth of prepubertal rats. *Toxicol Lett* 170:165–176.
- Van Vlaardingen PL, Traas TP, Wintersen AM, Aldenberg T. 2004. ETX 2.0. A program to calculate risk limits and fraction affected, based on normal species sensitivity distributions. Report 601501028/2004. Bilthoven (NL): National Institute for Public Health and the Environment (RIVM).
- Van Wezel AP, De Vries DAM, Sijm DTHM, Opperhuizen A. 1996. Use of the lethal body burden in the evaluation of mixture toxicity. *Ecotoxicol Environ Safety* 35:236–241.
- Van Wijk RJ, Postma JF, Van Houwelingen H. 1994. Joint toxicity of ethyleneamines to algae, daphnids and fish. *Environ Toxicol Chem* 13:167–171.
- Venkatakrisnan K, Von Moltke LL, Greenblatt DJ. 2000. Effects of the antifungal agents on oxidative drug metabolism: clinical relevance. *Clin Pharmacokinet* 38:111–180.
- Venkatakrisnan K, Von Moltke LL, Greenblatt DJ. 2001. Human drug metabolism and the cytochromes P450: application and relevance of *in vitro* models. *J Clin Pharmacol* 41:1149–1179.
- Verhaar HJM, Van Leeuwen CJ, Hermens J. 1992. Classifying environmental pollutants. 1. Structure–activity relationships for prediction of aquatic toxicity. *Chemosphere* 25:471–491.
- Vijver MG, Van Gestel CAM, Lanno RP, Van Straalen NM, Peijnenburg WJGM. 2004. Internal metal sequestration and its ecotoxicological relevance: a review. *Environ Sci Technol* 38:4705–4712.
- Vijver MG, Vink JPM, Jager T, Wolterbeek HT, Van Straalen NM, Van Gestel CAM. 2005. Biphasic elimination and uptake kinetics of Zn and Cd in the earthworm *Lumbricus rubellus* exposed to contaminated floodplain soil. *Soil Biol Biochem* 37:1843–1851.
- Vink K, Dewi L, Bedaux J, Tompot A, Hermans M, Van Straalen NM. 1995. The importance of exposure route when testing the toxicity of pesticides to saprotrophic isopods. *Environ Toxicol Chem* 14:1225–1232.
- Voet D, Voet JG. 2004. Biochemistry. 3rd ed. Toronto (CA): John Wiley & Sons.
- VROM. 1989. Premises for risk management. Risk limits in the context of environmental policy. Parliament session 1988–1989, 21137, no 5. The Hague (NL): Ministry of Housing, Spatial Planning and the Environment (VROM).
- Wade MG, Foster WG, Younglai EV, McMahon A, Leingartner K, Yagminas A, Blakey D, Fournier M, Desaulniers D, Hughes CL. 2002. Effects of subchronic exposure to a complex mixture of persistent contaminants in male rats: systemic, immune, and reproductive effects. *Toxicol Sci* 67:131–143.
- Walker CH, Hopkin SP, Sibly RM, Peakall DB. 2001. Principles of ecotoxicology. London: Taylor & Francis.
- Walker CH, Johnston GO. 1993. Potentiation of pesticide toxicity in birds: role of cytochrome P-450. *Biochem Soc Trans* 21:1066–1068.
- Walter H, Consolaro F, Gramatica P, Scholze M, Altenburger R. 2002. Mixture toxicity of priority pollutants at no observed effect concentrations (NOECs). *Ecotoxicology* 11:299–310.
- Wania F, Mackay D. 1996. Tracking the distribution of persistent organic pollutants. *Environ Sci Technol* 30:A390–A396.

- Warne MStJ. 2003. A review of the ecotoxicity of mixtures, approaches to, and recommendations for, their management. In: Langley A, Gilbey M, Kennedy B, editors, Proceedings of the Fifth National Workshop on the Assessment of Site Contamination. Adelaide (AU): National Environment Protection Council Service Corporation, p 253–276.
- Wassenberg DM, Di Guilio RT. 2004. Synergistic embryotoxicity of polycyclic aromatic hydrocarbons aryl hydrocarbon receptor agonists with cytochrome P4501A inhibitors in *Fundulus heteroclitus*. *Environ Health Perspect* 112:1658–1664.
- Watts AW, Ballesterio, TP, Gardener KH. 2006. Uptake of polycyclic aromatic hydrocarbons (PAHs) in salt marsh plants *Spartina alterniflora* grown in contaminated sediments. *Chemosphere* 62:1253–1260.
- Weis BK, Balshaw D, Barr JR, Brown D, Ellisman M, Liov P, Omenn G, Potter JD, Smith MT, Sohn L, Suk WA, Sumner S, Swenberg J, Walt DR, Watkins S, Thompson C, Wilson SH. 2005. Personalized exposure assessment: promising approaches for human environmental health research. *Environ Health Perspect* 113:840–848.
- White JC, Pignatello JJ. 1999. Influence of bisolute competition on the desorption kinetics of polycyclic aromatic hydrocarbons in soil. *Environ Sci Technol* 33:4292–4298.
- Whitfield J. 2001. Vital signs. *Nature* 411:989–990.
- WHO. 2001. Integrated risk assessment. Report prepared for the WHO/UNEP/ILO International Programme on Chemical Safety. WHO/IPCS/IRA/01/12. Geneva (CH): World Health Organization, International Programme on Chemical Safety.
- Whyatt RM, Camann DE, Kinney PL, Reyes A, Dietrich J, Diaz D, Holmes D, Perera FP. 2002. Residential pesticide use during pregnancy among urban minority women. *Environ Health Perspect* 110:507–514.
- Willett KL, Wassenberg D, Lienesch L, Reichert W, Di Guilio RT. 2001. *In vivo* and *in vitro* inhibition of CYP1A-dependent activity in *Fundulus heteroclitus* by the polynuclear aromatic hydrocarbon fluoranthene. *Toxicol Appl Pharmacol* 177:264–271.
- Winters D, Cleverly D, Meier K, Dupuy A, Byrne C, Deyrup C, Ellis R, Ferrario J, Harless R, Lesse W, Lorber M, McDaniel D, Schaum J, Walcott J. 1996. A statistical survey of dioxin-like compounds in the United States beef. *Chemosphere* 32:469–478.
- Wintersen A, Posthuma L, De Zwart D. 2004. The RIVM e-toxBase. A database for storage, retrieval and export of ecotoxicity data. Bilthoven (NL): National Institute for Public Health and the Environment (RIVM).
- Witherow LE, Houston JB. 1999. Sigmoidal kinetics of CYP3A substrates: an approach for scaling dextromethorphan metabolism in hepatic microsomes and isolated hepatocytes to predict *in vivo* clearance in rat. *J Pharmacol Exp Ther* 290:58–65.
- Woo YT, Lai D, McLain JL, Manibusan MK, Dellarco V. 2002. Use of mechanism-based structure–activity relationships analysis in carcinogenic potential ranking for drinking water disinfection by-products. *Environ Health Perspect* 110:75–87.
- Wright JF, Sutcliffe DW, Furse MT, editors. 2000. Assessing the biological quality of fresh waters RIVPACS and other techniques. Ambleside (UK): The Freshwater Biological Association.
- Yang RSH, editor. 1994. Toxicology of chemical mixtures: case studies, mechanisms, and novel approaches. San Diego (CA): Academic Press.
- Yang RSH. 1997. Toxicologic interactions of chemical mixtures. In: Bond JA, editor, Comprehensive toxicology. Vol. 1. Oxford (UK): Elsevier Science Ltd. p 189–203.
- Yang RSH, Andersen ME. 2005. Physiologically based pharmacokinetic modeling of chemical mixtures. In: Reddy MB, Yang RSH, Clewell HJ III, Andersen ME, editors, Physiologically based pharmacokinetics: science and applications. New York: John Wiley & Sons, p 349–373.
- Yang RSH, El-Masri HA, Thomas RS, Dobrev ID, Dennison JE, Bae DS, Campaign JA, Liao KH, Reisfeld B, Andersen ME, Mumtaz M. 2004. Chemical mixture toxicology: from descriptive to mechanistic, and going on to *in silico* toxicology. *Environ Toxicol Pharmacol* 18:65–81.

- Yang RSH, Mayeno AN, Liao KH, Reardon KF, Reisfeld B. 2006. Integration of PBPK and reaction network modeling: predictive xenobiotic metabolomics. *ALTEX* 23(Special Issue 2):373–379.
- Yang RSH, Mayeno AN, Lyons M, Reisfeld B. 2010. The application of physiologically-based pharmacokinetics (PBPK), Bayesian population PBPK modeling, and biochemical reaction network (BRN) modeling to chemical mixture toxicology. In: Mumtaz M, editor, *Principles and practices of mixture toxicology*. Hoboken (NJ): John Wiley & Sons.
- Young JF, Wosilait WD, Luecke RH. 2001. Analysis of methylmercury disposition in humans utilizing a PBPK model and animal pharmacokinetic data. *J Toxicol Environ Health A* 63:19–52.
- Yu X, Johanson G, Ichihara G, Shibata E, Kamijima M, Ono Y, Takeuchi Y. 1998. Physiologically based pharmacokinetic modeling of metabolic interactions between n-hexane and toluene in humans. *J Occup Health* 40:293–301.
- Zhang H. 2004. *In-situ* speciation of Ni and Zn in freshwaters: comparison between DGT measurements and speciation models. *Environ Sci Technol* 38:1421–1427.
- Zhang H, Zhao FJ, Sun B, Davison W, McGrath SP. 2001. A new method to measure effective soil solution concentration predicts copper availability to plants. *Environ Sci Technol* 35:2602–2607.
- Zhao Y, Newman MC. 2004. Shortcomings of the laboratory-derived median lethal concentration for predicting mortality in field populations: exposure duration and latent mortality. *Environ Toxicol Chem* 23:2147–2153.
- Zhao Y, Newman MC. 2007. The theory underlying dose–response models influences predictions for intermittent exposure. *Environ Toxicol Chem* 26:543–547.
- Zhou S, Kestell P, Paxton JW. 2002. Predicting pharmacokinetics and drug interactions in patients from *in vitro* and *in vivo* models: the experience with 5,6-dimethylxanthene-4-acetic acid (DMXAA), an anti-cancer drug eliminated mainly by conjugation. *Drug Metab Rev* 34:751–790.
- Zwick M, Renn O. 1998. *Wahrnehmung und Bewertung von Technik in Baden-Württemberg. Eine Präsentation der Akademie für Technikfolgenabschätzung in Baden-Württemberg*. Stuttgart (DE): Akademie für Technikfolgenabschätzung in Baden-Württemberg. Available from: <http://elib.uni-stuttgart.de/opus/volltexte/2004/1765/>